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## CLINICAL SCIENCE

Interaction between the *STAT4* rs11889341(T) risk allele and smoking confers increased risk of myocardial infarction and nephritis in patients with systemic lupus erythematosus

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**ABSTRACT**

**Objective** To investigate how genetics influence the risk of smoking-related systemic lupus erythematosus (SLE) manifestations.

**Methods** Patients with SLE (n<sub>discovery cohort</sub>=776, n<sub>replication cohort</sub>=836) were genotyped using the 200K Immunochip single nucleotide polymorphisms (SNP) Array (Illumina) and a custom array. Sixty SNPs with SLE association (p<5.0×10<sup>-8</sup>) were analysed. Signal transducer and activator of transcription 4 (*STAT4*) activation was assessed in *in vitro* stimulated peripheral blood mononuclear cells from healthy controls (n=45).

**Results** In the discovery cohort, smoking was associated with myocardial infarction (MI) (OR 1.96 (95% CI 1.09 to 3.55)), with a greater effect in patients carrying any rs11889341 *STAT4* risk allele (OR 2.72 (95% CI 1.24 to 6.00)) or two risk alleles (OR 8.27 (95% CI 1.48 to 46.27)).

Smokers carrying the risk allele also displayed an increased risk of nephritis (OR 1.47 (95% CI 1.06 to 2.03)). In the replication cohort, the high risk of MI in smokers carrying the risk allele and the association between the *STAT4* risk allele and nephritis in smokers were confirmed (OR 6.19 (95% CI 1.29 to 29.79) and 1.84 (95% CI 1.05 to 3.29), respectively).

The interaction between smoking and the *STAT4* risk allele resulted in further increase in the risk of MI (OR 2.14 (95% CI 1.01 to 4.62)) and nephritis (OR 1.53 (95% CI 1.08 to 2.17)), with 54% (MI) and 34% (nephritis) of the risk attributable to the interaction. Levels of interleukin-12-induced phosphorylation of *STAT4* in CD8+ T cells were higher in smokers than in non-smokers (mean geometric fluorescence intensity 1063 vs 565, p=0.0063).

Lastly, the *IL12A* rs564799 risk allele displayed association with MI in both cohorts (OR 1.53 (95% CI 1.01 to 2.31) and 2.15 (95% CI 1.08 to 4.26), respectively).

**Conclusions** Smoking in the presence of the *STAT4* risk gene variant appears to increase the risk of MI and nephritis in SLE. Our results also highlight the role of the IL12–*STAT4* pathway in SLE-cardiovascular morbidity.

**Key messages****What is already known about this subject?**

► Neither traditional nor systemic lupus erythematosus (SLE)-related risk factors can fully account for the excess cardiovascular disease risk seen in patients with SLE, but interactions between traditional and SLE-specific risk factors have been scarcely investigated.

**What does this study add?**

► Our results show that the signal transducer and activator of transcription 4 (*STAT4*) risk allele rs11889341 enhances the effect of smoking on the risk of myocardial infarction and nephritis and that smoking is associated with increased interleukin (IL)-12-induced phosphorylation of *STAT4* in CD8+ T cells.  
► We further demonstrate that the *IL12A* SLE risk variant rs564799 is associated with an increased risk of myocardial infarction, which further highlights the importance of the IL12–*STAT4* pathway in the aetiology of cardiovascular morbidity in SLE.

**How might this impact on clinical practice or future developments?**

► Our results suggest that genetic profiling of patients with SLE may be useful for predicting comorbidities of the disease, impact of environmental factors and for targeted smoking cessation interventions.

**INTRODUCTION**

Systemic lupus erythematosus (SLE) is a disease characterised by loss of tolerance to self-antigens, formation of immune complexes and an activated type I interferon (IFN) system.<sup>1</sup> A widely accepted view of the aetiology of SLE is that environmental factors trigger the disease in genetically susceptible individuals. The genetic background is complex, with more than 100 single nucleotide polymorphisms (SNPs) associated with risk

for SLE.<sup>2</sup> Exposure to certain environmental factors, including ultraviolet radiation and viral infections, is associated with SLE development and flare-ups of the disease.<sup>3,4</sup> Several studies have evaluated smoking as a risk factor for SLE, with the largest meta-analysis to date showing a modest risk increase.<sup>5</sup> While the results are not confirmed in prospective studies, both Cozier and Barbhuiya *et al* observed a trend of increased risk in smokers.<sup>6,7</sup> The most extensive prospective study involving 286 cases with SLE demonstrated an association between smoking and development of SLE with increased anti-dsDNA, but no risk of overall SLE.<sup>7</sup>

Although death from active SLE has decreased since the 1950s,<sup>8</sup> the mortality rate still exceeds that of the general population, with cardiovascular morbidity remaining considerably high and a strong risk factor for premature mortality.<sup>9–11</sup> Both traditional and SLE-related risk factors, such as hypertension, nephritis and high disease activity have been identified as risk factors, but cannot fully account for the excess cardiovascular disease (CVD) risk seen in patients with SLE.<sup>12,13</sup> To fully explain the aetiology of SLE or its comorbidities such as CVD, gene–gene or gene–environment interactions may be essential to consider. In rheumatoid arthritis, there is compelling evidence of a strong interaction between the HLA-DRB1\*04 shared epitope and smoking on the development of anticitrullinated protein

autoantibodies<sup>14</sup> and a high prevalence of cardiovascular events (CVE).<sup>15</sup>

In SLE, a few studies have investigated the interaction between genetic risk factors and smoking on the development of the disease.<sup>16,17</sup> Recently, Cui *et al* demonstrated that an additive interaction between smoking and the cumulative genetic risk of SLE increases the risk of the disease.<sup>18</sup> However, gene–smoking interactions on the development of specific manifestations or co-morbidities of SLE have been scarcely studied. This study, therefore, aims to investigate the effect of smoking on the development of specific manifestations of SLE, including CVE, end-stage renal disease (ESRD) and nephritis, and examine how the effect is modulated by the presence of genetic variants associated with an increased risk of SLE development.

## METHODS

### Patients of the discovery and replication cohort

The discovery cohort included 774 patients with SLE from Sweden. The replication cohort included 836 patients from Norway and Denmark. All subjects fulfilled  $\geq 4$  American College of Rheumatology (ACR)–82 and ACR-97 classification criteria for SLE and were of European descent. Clinical characteristics of the cohorts are described in table 1 and online supplemental

**Table 1** Prevalence of clinical manifestations in smokers (n=371) and non-smokers (n=387) in the discovery cohort

	Smokers, n (%)	Non-smokers, n (%)	OR (95% CI)	P value
Age at last follow-up, mean (SD)	55 (15)	50 (17)		<b>0.000025</b>
Disease duration, mean (SD)	17 (11)	16 (12)		<b>0.016</b>
Male sex	49 (13)	48 (13)	0.94 (0.61 to 1.45)	0.79
Deceased at follow-up	55 (15)	39 (11)	1.19 (0.75 to 1.88)	0.45
ACR 1982 classification criteria <sup>40</sup>				
1 Malar rash	205 (55)	211 (57)	1.00 (0.74 to 1.34)	0.97
2 Discoid rash	83 (22)	80 (21)	0.96 (0.68 to 1.36)	0.81
3 Photosensitivity	262 (71)	246 (65)	1.27 (0.93 to 1.74)	0.13
4 Oral ulcer	103 (28)	96 (25)	1.20 (0.86 to 1.67)	0.27
5 Arthritis	306 (82)	301 (81)	1.11 (0.76 to 1.61)	0.59
6 Serositis	179 (48)	165 (45)	1.10 (0.82 to 1.47)	0.52
7 Renal disorder	129 (35)	132 (36)	1.10 (0.80 to 1.50)	0.56
8 Neurological disorder	33 (9)	40 (10)	0.86 (0.52 to 1.40)	0.54
9 Haematological disorder	213 (57)	258 (70)	<b>0.62 (0.46 to 0.84)</b>	<b>0.0021</b>
10 Immunological disorder	245 (66)	256 (69)	0.97 (0.71 to 1.33)	0.86
Anti-dsDNA	224 (61)	231 (63)	1.02 (0.76 to 1.38)	0.88
11 ANA	364 (98)	367 (99)	0.69 (0.19 to 2.49)	0.57
Renal variables				
WHO class I–II	12 (5)	20 (8)	0.71 (0.33 to 1.50)	0.37
WHO class III–IV	62 (21)	71 (22)	1.13 (0.75 to 1.69)	0.57
WHO class V	15 (6)	16 (6)	1.05 (0.50 to 2.20)	0.90
Other*	11 (4)	6 (2)	1.24 (0.50 to 3.06)	0.65
ESRD	14 (4)		1.41 (0.62 to 3.24)	0.42
Cardiovascular events				
MI	39 (11)	19 (5)	<b>1.96 (1.09 to 3.55)</b>	<b>0.025</b>
ICVD	45 (12)	30 (8)	1.37 (0.84 to 2.24)	0.21
VTE	61 (16)	52 (14)	1.14 (0.76 to 1.71)	0.52
Clinical APS	68 (20)	61 (18)	1.08 (0.73 to 1.60)	0.68
Anti- $\beta$ 2GP-I IgG	58 (19)	57 (18)	0.98 (0.69 to 1.39)	0.91
Anti- $\beta$ 2GP-I IgM	8 (11)	11 (12)	0.96 (0.36 to 2.55)	0.93
LA	62 (23)	57 (21)	1.19 (0.79 to 1.80)	0.40
aCL-IgG	86 (26)	90 (28)	1.05 (0.70 to 1.59)	0.080
aCL-IgM	34 (14)	33 (13)	0.96 (0.36 to 2.55)	0.93

Logistic regression models were used to assess differences between smokers and non-smokers. All analyses were adjusted for age at last follow-up and disease duration.  $p < 0.05$  (unadjusted for multiple comparisons) in bold.

\*Patients with biopsies displaying signs of nephritis but not meeting the criteria for any of the above classes were classified as *other*.

ACR, American College of Rheumatology; ANA, antinuclear antibodies; Anti- $\beta$ 2GP-I, anti- $\beta$ 2-glycoprotein-I; APS, antiphospholipid syndrome; dsDNA, double-stranded DNA; ESRD, end-stage renal disease; ICVD, ischaemic cerebrovascular disease; LA, lupus anticoagulant; MI, myocardial infarction; VTE, venous thromboembolism.

table 1, respectively. Clinical data, including smoking status (ever-smoker, including current or a history of smoking, versus never-smoker), the ACR-82 classification criteria, antiphospholipid syndrome diagnosis, ESRD, renal biopsy data and CVE, defined as myocardial infarction (MI), ischaemic cerebrovascular disease (ICVD) or venous thromboembolism, was collected from medical records. For definitions, see online supplemental file. In the replication cohort, data regarding ESRD were not available, and data on smoking were available from 503 patients.

### Genotyping and selection of SNPs

Genotyping of the discovery cohort was performed using the Illumina 200K ImmunoChip SNP array, for details, see online supplemental file. SNPs previously associated with SLE at genome-wide significance in the European population<sup>2</sup> were selected. For SNPs not included on the ImmunoChip, the SNP-proxy with the highest linkage disequilibrium (LD) ( $r^2 \geq 0.96$ ) was selected. All SNPs were filtered for independent signals, removing the variant with the lowest SLE-OR for SNPs in LD ( $r^2 > 0.2$ ). In total, 4 HLA and 56 non-HLA SNPs were investigated for associations with MI (online supplemental table 2). Individuals in the replication cohort were genotyped for three single nucleotide variants using a custom assay on the MassARRAY system (see online supplemental file).

### Interleukin-12-induced phosphorylation of STAT4

Interleukin 12 (IL-12)-induced phosphorylation of signal transducer and activator of transcription 4 (pSTAT4) was previously determined in 72 healthy blood donors from Uppsala Bioresource using flow cytometry.<sup>19</sup> Smoking data were available from 45 of these donors, of which 20 were past or current smokers and 25 were non-smokers.

### Statistical analysis

To investigate associations between smoking and clinical manifestations, logistic regression models were used. As smoking was associated with longer disease duration and higher age at follow-up (table 1), these variables were included as covariates. In analysis of associations between genetic variants and MI, SNPs were first analysed separately. Next, all variants demonstrating a positive association with MI were included in a forwards conditional multiple regression model. All analyses were adjusted for age and disease duration. Results considered statistically significant (unadjusted  $p < 0.05$ ) were reanalysed in the replication cohort using the same statistical model and covariates. Meta-analyses were performed on the two datasets and multiplicative and additive interactions between the *STAT4* risk allele and smoking were studied in a combined dataset through addition

of a *STAT4*\*smoking interaction term in the logistic models and by calculating the attributable proportion due to interaction, respectively.<sup>20, 21</sup> Differences in levels of pSTAT4 were assessed by Student's t-test and by a linear regression model allowing adjustment for age and the *STAT4* risk allele.  $R^{22}$  was used for all analyses except the meta-analyses which were performed in PLINK.<sup>23</sup>

## RESULTS

### Smoking is modestly associated with MI

Initially, we assessed the association between smoking and clinical manifestations (table 1). We found no evidence of any associations between smoking and the ACR criteria, except the haematological criterion, which was less prevalent in smokers (table 1). Elevated levels of red and white blood cells in smokers is a well-known phenomenon.<sup>24</sup> Smoking was not associated with a history of DVT or ICVD, however, a significant association between smoking and MI was observed (OR 1.96 (95% CI 1.09 to 3.55),  $p = 0.025$ ) (table 1).

### Increased risk of MI in SLE-smokers with the *STAT4* risk allele

Next, we asked whether there are sub-groups of patients in which smoking plays a more prominent role in MI development. We initially examined 60 SNPs with established association with SLE ( $p < 5.0 \times 10^{-8}$ ) for association with MI (online supplemental table 2). We found that the Neutrophil Cytosolic Factor 2 (*NCF2*), Interleukin-12A (*IL12A*) and *STAT4* risk alleles displayed independent, positive association with MI (table 2). In addition, patients carrying two alleles of both the *STAT4* and the *IL12A* risk variants ( $n = 37$ , 4.9% of the patients) displayed a substantially higher prevalence of MI compared with those with any other allele combination (27% vs 7%) (OR 5.88 (95% CI 2.44 to 14.17),  $p = 7.9 \times 10^{-5}$ ) (figure 1A).

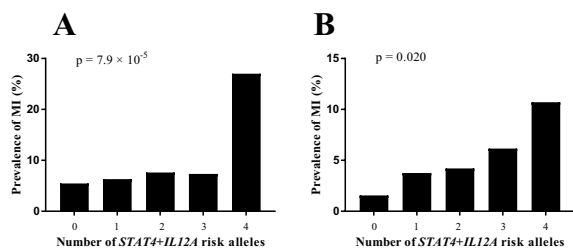
Next, we stratified patients by smoking status to determine whether each of the three SNPs displayed stronger association with MI in smokers. No significant associations were found for the *NCF2* or *IL12A* risk alleles (OR 1.58 (95% CI 0.89 to 2.78),  $p = 0.12$  and OR 1.36 (95% CI 0.89 to 2.08),  $p = 0.15$ , respectively). However, the *STAT4* risk allele demonstrated a stronger association in smokers (OR 2.45 (95% CI 1.46 to 4.19),  $p = 0.00086$ ) (figure 2A). Next, we assessed the association between smoking and MI in patients carrying the *STAT4* risk allele and observed an almost 3-fold increase in risk for the smokers compared with the non-smokers (OR 2.72 (95% CI 1.24 to 6.00),  $p = 0.013$ ). In patients carrying two risk alleles, the risk was more than eightfold higher for smokers (OR 8.27 (95% CI 1.48 to 46.27),  $p = 0.016$ ). In contrast, we could not demonstrate a significant association between smoking and MI

**Table 2** Associations between SLE risk SNPs and myocardial infarction in the discovery and replication cohort

SNP	Gene name	Discovery cohort (n=763)				P value	Replication cohort (n=836)				
		HWE p	Risk allele frequency				HWE p	Risk allele frequency			P value
			MI+	MI-	OR			MI+	MI-	OR	
rs17849502	<i>NCF2</i>	0.013	0.12	0.08	<b>2.00 (1.08 to 3.68)</b>	<b>0.027</b>	$8.80 \times 10^{-7}$	0.18	0.11	1.94 (0.94 to 4.04)	0.075
rs11889341	<i>STAT4</i>	0.81	0.44	0.34	<b>1.76 (1.18 to 2.63)</b>	<b>0.0054</b>	0.53	0.40	0.31	1.81 (0.94 to 3.47)	0.075
rs564799	<i>IL12A</i>	0.29	0.68	0.59	<b>1.53 (1.01 to 2.31)</b>	<b>0.042</b>	1.00	0.74	0.61	<b>2.15 (1.08 to 4.26)</b>	<b>0.029</b>

Using a forward conditional multiple logistic regression model, 60 genetic variants with previously established association with SLE ( $p < 5 \times 10^{-8}$ , online supplemental table 2) were analysed for associations with myocardial infarction in the discovery cohort. The table shows SNPs included in the final model. These SNPs were subsequently analysed in the replication cohort. Age at follow-up and disease duration were included as covariates.  $P < 0.05$  (unadjusted for multiple comparisons) in bold. HWE was tested on all patients in the discovery and replication cohorts, respectively.

HWE, Hardy-Weinberg equilibrium; IL12a, Interleukin12A; MI, myocardial infarction; NCF2, neutrophil cytosolic factor 2; SLE, systemic lupus erythematosus; SNPs, single-nucleotide polymorphisms; STAT4, signal transducer and activator of transcription 4.



**Figure 1** Prevalence of myocardial infarction (MI) in patients with different numbers of *STAT4* and *IL12A* risk alleles. The prevalence of MI for patients in the discovery (A) and the replication (B) cohorts, stratified by number of risk alleles of *STAT4* (rs11889341) and *IL12A* (rs564799). Patients carrying two risk alleles of both *STAT4* and *IL12A* ( $n_{\text{discovery cohort}}=37$  (4.9%),  $n_{\text{replication cohort}}=28$  (3.4%)) were compared with the remaining patients using logistic regression, adjusting for age at follow-up and disease duration. IL12, interleukin-12; *STAT4*, Signal Transducer and Activator of Transcription 4.

in patients without the risk allele (OR 1.20 (95% CI 0.49 to 2.96)  $p=0.55$ ).

As patients with nephritis have previously been shown to have a higher prevalence of both MI and the *STAT4* risk allele.<sup>25–27</sup> we hypothesised that the results would be similar if using nephritis, rather than MI, as the outcome variable. Without stratifying for smoking, the association between the *STAT4* risk allele and nephritis reached suggestive significance (OR 1.23 (95% CI 0.98 to 1.54),  $p=0.072$ ). The effect was more pronounced in the smokers only (OR 1.47 (95% CI 1.06 to 2.03),  $p=0.020$ ). In addition, we found moderate evidence that patients with nephritis carrying the *STAT4* risk allele were at a greater risk of developing ESRD (OR 1.85 (95% CI 0.96 to 3.59),  $p=0.068$ ), and this risk was enhanced in smokers (OR 2.52 (95% CI 1.04 to 6.10),  $p=0.040$ ) (figure 2B). Of note, despite the non-smoking group including more patients with nephritis ( $n=140$  vs  $n=129$ ), no evidence of an association between the *STAT4* risk allele and nephritis or ESRD could be demonstrated in this group (OR 1.07 (95% CI 0.77 to 1.46),  $p=0.70$  and OR 1.10 (95% CI 0.38 to 3.16),  $p=0.86$ , respectively).

To validate our significant findings, we performed the same analyses in an independent cohort of patients with SLE (online supplemental table 1). Analysis of the genetic variants demonstrated that the *IL12A* risk allele was the only gene variant significantly associated with MI when not accounting for smoking

(table 2). Patients with two risk alleles of both *STAT4* and *IL12A* ( $n=28$ , 3.2% of the patients) were found to have a significantly higher prevalence of MI than patients without this combination of risk alleles (OR 7.21 (95% CI 1.36 to 38.27,  $p=0.020$ ) (figure 1B). Similar to in the discovery cohort, we found a significant association between the *STAT4* risk allele and MI in smokers (OR 2.11 (95% CI 1.04 to 4.26),  $p=0.038$ ). In addition, smoking was associated with MI in patients carrying the *STAT4* risk allele (OR 6.19 (95% CI 1.29 to 29.79),  $p=0.023$ ). No evidence of these associations could be observed in the non-smoking group (OR 0.58 (95% CI 0.09 to 3.90),  $p=0.58$  and OR 1.32 (95% CI 0.23 to 7.34),  $p=0.75$ , respectively). In patients carrying two risk alleles ( $n=51$ ), the logistic regression could not be performed due to a ‘perfect separation’ between groups, with 9% of smokers having had a MI compared with 0% of never-smokers. As in the discovery cohort, we found an association between the *STAT4* risk allele and nephritis in smokers (OR 1.84 (95% CI 1.05 to 3.29),  $p=0.035$ ), whereas the effect size was non-significant in non-smokers (OR 0.78 (95% CI 0.41 to 1.45),  $p=0.44$ ).

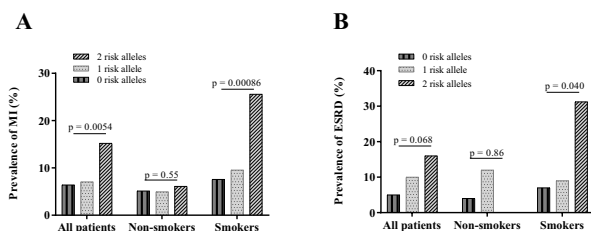
Meta-analysis of the two cohorts showed the risk of MI to more than double with each additional *STAT4* risk allele in smokers (OR 2.28,  $p=0.00010$ ). In contrast, no association could be detected in the never-smoker group (OR 0.80,  $p=0.52$ ). Similarly, in meta-analysis of patients with 2 *STAT4* risk alleles ( $n=282$ , 22%), smoking was found to be a strong risk factor for MI (OR 8.27,  $p=0.016$ ). For nephritis, each *STAT4* risk allele increased the risk by ~50% in smokers (OR 1.52,  $p=0.00051$ ), whereas no increase in risk was found in the non-smoker group (OR 0.82,  $p=0.33$ ).

### Interaction between *STAT4* risk allele and smoking results in a higher risk of MI and nephritis

We subsequently performed interaction analyses on all patients and found a significant multiplicative interaction between the *STAT4* risk allele and smoking on the development of MI (OR 2.14 (95% CI 1.01 to 4.62),  $p=0.049$ ) as well as nephritis (OR 1.53 (95% CI 1.08 to 2.17),  $p=0.020$ ) (online supplemental table 3). Next, we examined additive interaction and observed an attributable proportion due to interaction of 0.54 (95% CI 0.24 to 0.83,  $p=0.00019$ ) and 0.34 (95% CI 0.080 to 0.61,  $p=0.0051$ ) for MI and nephritis, respectively.

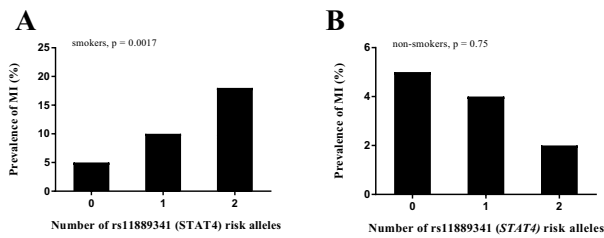
To determine whether the effect of the *STAT4* risk allele on nephritis in smokers could explain the association with MI, we performed stratification of the combined dataset and investigated the association between the *STAT4* risk allele and MI in smokers and non-smokers without nephritis. In the smokers, the association between the *STAT4* risk allele and MI remained significant ( $n=428$ , OR 2.43 (95% CI 1.40 to 4.27),  $p=0.0017$ ) (figure 3). Similarly, the association between the *STAT4* risk allele and nephritis in smokers remained significant after excluding patients with MI from the analysis ( $n=623$ , OR 1.54 (95% CI 1.20 to 1.98),  $p=0.00076$ ).

Lastly, as both SLE risk alleles in *STAT4* and smoking have shown association with the development of aPL in previous studies,<sup>28 29</sup> we assessed the association between the *STAT4* risk allele and aPL in smokers, however, it was not significant (OR 1.56 (95% CI 0.71 to 3.72,  $p=0.26$ ). Next, we performed a multiple regression model in the smoking group including the *STAT4* risk allele, any aPL, nephritis, age at follow-up, and disease duration as covariates. We found the association between the *STAT4* risk allele and MI to remain significant (OR 3.26 (95% CI 1.15 to 9.20),  $p=0.026$ ), whereas neither aPL nor



**Figure 2** Prevalence of myocardial infarction (MI) and end-stage renal disease (ESRD) in the discovery cohort. Prevalence of MI (A) and ESRD (B) in the discovery cohort in all patients ( $n=776$ ), smokers only ( $n=371$ ) and non-smokers only ( $n=405$ ), stratified by the number of *STAT4* (rs11889341) risk alleles. Patients with 0, 1 or 2 risk alleles in each group were compared using logistic regression, adjusting for age at follow-up and disease duration. *STAT4*, Signal Transducer and Activator of Transcription 4.



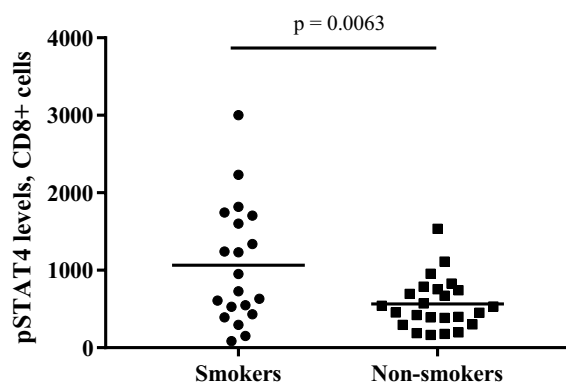


**Figure 3** The prevalence of myocardial infarction (MI) in patients without nephritis. To investigate whether the association between the rs11889341 (*STAT4*) allele and MI was dependent on the association between the *STAT4*–nephritis association, all patients with nephritis were excluded and the prevalence of MI was subsequently plotted for smoking (A) and non-smoking (B) patients with 0, 1 or 2 of the *STAT4* risk allele. Patients with 0, 1 or 2 risk alleles in each group were compared using logistic regression, adjusting for age at follow-up and disease duration. *STAT4*, signal transducer and activator of transcription 4.

nephritis displayed significant association with MI ( $p=0.87$  and  $p=0.50$ , respectively), (online supplemental table 4).

#### Levels of activated *STAT4* are increased in smokers

The *STAT4* risk allele is associated with higher levels of p*STAT4* in CD8+ T cells from SLE patients on stimulation with IL-12.<sup>19</sup> Furthermore, higher levels of *STAT4* expression have demonstrated association with increased cardiovascular damage in patients with SLE.<sup>30</sup> To investigate whether smoking also increases the levels of p*STAT4*, we analysed IL-12 stimulated CD8+ T cells from healthy blood donors who were smokers ( $n=20$ ) or never-smokers ( $n=25$ ). We found the levels of p*STAT4* to be higher in smokers ( $p=0.0063$ ), with a mean value of 1063 compared with 565 in non-smokers (figure 4). When adjusting for age and the *STAT4* risk allele, which is in LD ( $r^2=1.00$ ) with a *STAT4* risk variant previously shown to influence levels of p*STAT4* in these individuals,<sup>19,31</sup> the association between smoking and levels of p*STAT4* remained significant ( $\beta=396$ ,  $p=0.023$ ).



**Figure 4** Levels of p*STAT4* in CD8+ T cells after stimulation with IL-12. The levels of IL-12–induced p*STAT4* were compared between healthy blood donors who were smokers (current or past) ( $n=20$ ) and never-smokers ( $n=25$ ) by Student's t-test. IL-12, interleukin-12; p*STAT4*, phosphorylated Signal Transducer and Activator of Transcription 4.

## DISCUSSION

In the present study, we demonstrate that smoking substantially increases the risk of MI in a subset of patients with SLE carrying a variant of the *STAT4* SLE-risk gene. In both the discovery and replication cohorts, the effect size increased with an increasing number of *STAT4* risk alleles, with smoking giving rise to a more than 8-fold risk of MI in homozygous individuals. We believe that our results add important knowledge in the understanding of how SLE-risk alleles can modulate the effect of traditional risk factors.

The prevalence of MI is higher in SLE patients with nephritis than patients without renal manifestations<sup>26,32</sup> and SLE-risk alleles in *STAT4* have previously been linked to both nephritis and severe renal insufficiency.<sup>25,27</sup> We, therefore, speculated that the smoking-*STAT4* risk allele interaction did not directly affect MI, but rather, was a consequence of an interaction between the *STAT4* risk allele and smoking on the development of nephritis. Indeed, we found that this gene-environment combination also results in a higher risk of nephritis, as well as ESRD. Interestingly, however, the *STAT4* risk allele/smoking effect on MI did not decrease when adjusting the model for nephritis or when completely removing the patients with nephritis from the analysis. Similarly, the effect of the *STAT4* risk allele/smoking on nephritis remained significant after excluding all patients with MI from the analysis, indicating that the associations were independent.

Based on these results, we hypothesised that the increased risk in individuals who smoke and carry the risk allele is connected with the levels of activated *STAT4* in these individuals. Hagberg *et al* have shown that the rs7574865 *STAT4* risk allele—which is in perfect LD ( $r^2=1.00$ ) with the SNP used in this study<sup>31</sup>—is associated with increased levels of p*STAT4* in activated CD8+T cells of SLE patients.<sup>19</sup> Therefore, we assessed whether smoking elevates p*STAT4* in this cell type and found that the levels were almost twofold higher in smokers. This observation is in line with previous findings by Di Stefano *et al*, who demonstrated higher levels of p*STAT4* in bronchial T cells from healthy smokers compared with non-smokers.<sup>33</sup> When *STAT4* is activated and phosphorylated, it homodimerises and translocates to the nucleus where it induces expression of hundreds of genes, resulting in production of IFN- $\gamma$ , T-helper type 1 and 17 differentiation and activation of monocytes.<sup>34</sup> Increased *STAT4* mRNA expression is associated with increased cardiovascular damage in patients with SLE,<sup>30</sup> and several studies on animal models indicate a link between *STAT4* and the development of atherosclerosis.<sup>35</sup> The mechanism of how smoking leads to increased levels of activated *STAT4* is unclear, however, we speculate that epigenetics may constitute the bridge between smoking, genetics and SLE. It is well known that smoking affects both overall DNA methylation and specific gene promoters.<sup>36,37</sup> Epigenetic regulation is further believed to play an important role both in cardiovascular biology and in SLE development.<sup>36,38</sup> Whether smoking is associated with epigenetic changes in SLE-specific genes, and if such changes are associated with specific manifestations of SLE, deserves further studies.

The analyses of individual SLE risk alleles identified the SLE-risk SNP *IL12A* to be associated with an increased risk of MI, and that patients in the discovery and replication cohort carrying two alleles of both the *IL12A* and *STAT4* risk SNPs had a more than fivefold and eightfold risk of MI, respectively. The *IL12A* SNP is located within the fourth intron of the *IL12A* gene, which encodes the p35 subunit of the IL-12 protein. On binding to its receptor, IL-12 induces phosphorylation of *STAT4*.<sup>34</sup> We

believe that the association of the *IL12A* risk allele, in addition to the *STAT4* risk allele, with MI points to the importance of this pathway in the development of the comorbidity. Previous work has demonstrated that JAK-inhibitors efficiently block the increase in pSTAT4 levels, and ameliorate murine lupus as well as its associated vascular dysfunction.<sup>19,39</sup> Due to the potential therapeutic strategy of JAK-inhibitors for patients with SLE displaying an altered activity in this particular pathway, we believe that further studies of the effect of this pathway on development of CVE are warranted.

This study's strength is the large, well-characterised discovery cohort, that results were validated in a second large cohort, and the analysis of healthy control cells, which confirmed that pSTAT4 levels are higher in smokers. In addition, the quality control of genetic data was rigorous, and the patients' long mean follow-up time of 17 years allowed for many outcome variables such as MI to be recorded. There are, however, some limitations. First, our study is based on retrospective data and we lacked data on the year of smoking cessations. As patients who were past smokers at the last follow-up may have been active smokers at the time of their CVE, we could not analyse previous and current smoking separately. Second, we did not have data on number of pack-years, which may have generated more precise results. Third, the study includes only Scandinavian patients with SLE, and whether the associations are generalisable to patients of other ethnicities needs further investigation.

## CONCLUSION

We demonstrate that smokers carrying the *STAT4* risk allele are at an increased risk of MI and nephritis and that the *IL12-STAT4* pathway may be important for the development of MI. Our results stress the importance of smoking cessation in SLE and particularly among those carrying this risk allele.

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**Supplementary table 1. Prevalence of clinical manifestations in the replication cohort (n=836)**

Clinical Variable	n (%)
Male sex	44 (12)
<b>ACR 1982 classification criteria[1]</b>	
1 Malar rash	216 (59)
2 Discoid rash	40 (11)
3 Photosensitivity	265 (72)
4 Oral ulcer	109 (30)
5 Arthritis	301 (82)
6 Serositis	151 (41)
7 Renal disorder	144 (39)
8 Neurological disorder	40 (11)
9 Hematologic disorder	245 (68)
10 Immunologic disorder	305 (83)
Anti-dsDNA	175 (76)
11 ANA	371 (100)
<b>Cardiovascular events</b>	
MI	21 (6)
ICVD	33 (9)
VTE	61 (16)
<b>Antiphospholipid antibodies</b>	
Clinical APS	32 (14)
Anti-β2GP-I IgG	35 (18)
Anti-β2GP-I IgM	25 (13)
LA	86 (39)
aCL-IgG	65 (29)
aCL-IgM	55 (24)

OR: odds ratio. CI: Confidence interval. ACR: American College of Rheumatology[1]. dsDNA: double-stranded DNA. ANA: Antinuclear antibodies. MI: Myocardial infarction. ICVD: Ischemic cerebrovascular disease. VTE: venous thromboembolism. APS: antiphospholipid syndrome. LA: Lupus anticoagulant. Anti-β2GP-I: anti-β<sub>2</sub> Glycoprotein-I.

**Supplementary table 2. The 60 SLE susceptibility SNPs investigated for associations with myocardial infarction in the discovery cohort.**

Location	Lead SNP in publication[2]	Gene	Discovery cohort		Location	Lead SNP in publication[2]	Gene	Discovery cohort	
			OR	P				OR	p
1p13.2	rs2476601	<i>PTPN22</i>	1.53 (0.93-2.51)	0.097	6q23	rs6932056	<i>TNFAIP3</i>	0.73 (0.28-1.90)	0.52
1q23.3	rs1801274	<i>FCGR2A</i>	1.18 (0.80-1.74)	0.40	6q23	rs2327832	<i>OLIG3-LOC100130476</i>	1.15 (0.75-1.77)	0.52
1q25.1	rs2205960	<i>TNFSF4</i>	1.36 (0.89-2.09)	0.16	7p15.1	rs849142	<i>JAZF1</i>	1.10 (0.75-1.62)	0.61
1q25.3	rs17849502	<i>NCF2</i>	1.92 (1.05-3.53)	0.035	7p12.2	rs4917014	<i>IKZF1</i>	0.76 (0.49-1.18)	0.23
1q25.3	rs10911363	<i>NCF2</i>	0.88 (0.56-1.40)	0.59	7q11.23	rs73366469	<i>GTF2IRD1-GTF2I</i>	1.05 (0.62-1.79)	0.85
1q31.3	rs34889541 <sup>a</sup>	<i>CD45</i>	1.01 (0.40-2.54)	0.99	7q32.1	rs4728142	<i>TNPO3-IRF5</i>	1.00 (0.67-1.51)	0.99
1q32.1	rs3024505	<i>IL10</i>	0.92 (0.56-1.53)	0.76	7q32.1	rs2070197	<i>TNPO3-IRF5</i>	0.63 (0.37-1.04)	0.073



2p23.1	rs7579944	LBH	0.96 (0.62-1.47)	0.85	8p23.1	rs2980512 <sup>a</sup>	FAM86B3P	0.81 (0.54-1.22)	0.32
2p14	rs6740462	SPRED2	0.82 (0.51-1.31)	0.40	8p23.1	rs2736340	BLK	0.80 (0.52-1.24)	0.32
2q24.2	rs2111485	IFIH1	0.76 (0.49-1.17)	0.21	8q12	rs7829816	LYN	0.54 (0.34-0.85)	0.0087
2q24.2	rs10930046	IFIH1	1.44 (0.40-5.14)	0.58	8q21	rs1966115	PKIA-ZC2HC1A	1.09 (0.72-1.65)	0.69
2q32.3	rs11889341	STAT4	1.71 (1.15-2.55)	0.0084	10q11.23	rs877819	WDFY4	1.12 (0.74-1.71)	0.58
3p14.3	rs6445972	ABHD6-PXK	1.20 (0.74-1.95)	0.46	11p15.5	rs4963128	IRF7-PHRF1	1.06 (0.68-1.66)	0.78
3q13.33	rs1132200	TMEM39A	1.57 (0.80-3.06)	0.19	11p13	rs2732552	CD44	1.07 (0.72-1.57)	0.75
3q25.33	rs564799	IL12A	1.43 (0.95-2.15)	0.085	11q13.1	rs1308020 <sup>a</sup>	RNASEH2C	1.29 (0.83-1.99)	0.26
3q26.2	rs10936599	MYNN	0.59 (0.39-0.91)	0.016	11q13.4	rs3794060 <sup>a</sup>	DHCR7-NADSYN1	0.87 (0.57-1.32)	0.52
4q24	rs10028805	BANK1	1.05 (0.70-1.57)	0.83	11q24.3	rs7941765 <sup>a</sup>	ETS1-FLI1	1.03 (0.69-1.52)	0.90
4q27	rs907715	IL21	0.80 (0.52-1.24)	0.32	12q24.12	rs10774625	SH2B3-ATXN2	0.87 (0.58-1.30)	0.49
5q31.1	rs7726414	TCF7-SKP1	1.12 (0.38-3.32)	0.84	12q24.32	rs1059312	SLC15A4	0.86 (0.58-1.29)	0.47
5q33.1	rs7708392 <sup>a</sup>	TNIP1	1.31 (0.87-1.98)	0.19	16p13	rs9652601	CLEC16A-CIITA-SOCS1	1.33 (0.84-2.09)	0.23
5q33.3	rs2431697	PTTG1-MIR146A	0.85 (0.57-1.28)	0.44	16p11.2	rs34572943	ITGAM-ITGAX	0.77 (0.42-1.39)	0.38
6p23	rs17603856	ATXN1	1.20 (0.79-1.81)	0.39	16q13	rs223881	CCL22	1.00 (0.65-1.56)	0.99
6p21.31	rs11755393	UHRF1BP1	0.90 (0.61-1.32)	0.59	16q22.1	rs1170426 <sup>a</sup>	ZPF90	0.89 (0.55-1.45)	0.65
6p21.31	rs2762340	ANKS1A	1.10 (0.74-1.64)	0.65	16q24.1	rs2280381	IRF8	1.11 (0.72-1.70)	0.64
6p21.32-33	rs3906272 <sup>a</sup>	HLA-C-HLA-B	0.85 (0.49-1.48)	0.57	17q12	rs2941509	IKZF3	1.49 (0.68-3.24)	0.31
6p21.32-33	rs1269852	MSH5-ATF6B	0.59 (0.35-1.02)	0.058	17q25	rs930297	GRB2	0.55 (0.25-1.22)	0.14
6p21.32-33	rs9270984 <sup>a</sup>	HLA-DRB1	1.73 (0.99-3.03)	0.053	19p13	rs3093030	ICAM1-ICAM4-ICAM5	1.34 (0.91-1.99)	0.14
6p21.32-33	rs2051549 <sup>a</sup>	HLA-DQA2	0.76 (0.51-1.14)	0.18	19p13	rs2304256	TYK2	0.85 (0.55-1.32)	0.46
6q15	rs597325	BACH2	0.86 (0.57-1.29)	0.46	20q13.13	rs11697848	RNF114	0.83 (0.28-2.45)	0.74
6q21	rs6568431	PRDM1-ATG5	0.83 (0.54-1.27)	0.38	22q11.21	rs7444	UBE2L3-YDJC-HIC2	0.75 (0.46-1.22)	0.25

SNP: single nucleotide polymorphism. We selected SNPs with a  $p < 5.0 \times 10^{-8}$  from Chen *et al.* [2]. All SNPs have previously reported genome-wide levels of significance for SLE ( $p < 5 \times 10^{-8}$ ). ORs for MI are based on comparisons between the 61 patients with MI and 713 patients without MI in the discovery cohort. <sup>a</sup>For SNPs not occurring on the ImmunoChip, the proxy SNP with the highest linkage disequilibrium [3] was used; for rs34889541, rs16843520 was used ( $r^2=1.00$ ); for rs7708392, rs6889239 was used ( $r^2=0.99$ ); for rs3906272, rs28732109 was used ( $r^2=1.00$ ); for rs9270984, rs9270986 was used ( $r^2=1.00$ ); for rs2051549, rs7453920 was used ( $r^2=1.00$ ); for rs2980512, rs2948286 was used ( $r^2=0.96$ ); for rs1308020, rs489574 was used ( $r^2=1.00$ ); for rs3794060, rs4944062 was used ( $r^2=1.00$ ); for rs7941765, rs6590343 was used ( $r^2=0.99$ ); for rs1170426, rs1170427 was used ( $r^2=1.00$ ).

Supplementary table 3 Interaction models including all patients (n=1277)

	Myocardial infarction		Nephritis <sup>1</sup>	
	OR (95% CI)	p	OR (95% CI)	P
Age at follow-up	<b>1.07 (1.05-1.09)</b>	<b>2.5x10<sup>-13</sup></b>	<b>0.97 (0.96-0.98)</b>	<b>1.0x10<sup>-12</sup></b>
Disease duration	<b>1.03 (1.01-1.05)</b>	<b>0.0038</b>	<b>1.02 (1.01-1.04)</b>	<b>5.4x10<sup>-5</sup></b>
rs11889341 (STAT4)	1.22 (0.58-2.69)	0.61	0.80 (0.57-1.13)	0.20

<b>Smoking<sup>2</sup></b>	1.07 (0.55-2.00)	0.84	1.00 (0.77-1.30)	0.99
<b>rs11889341*smoking</b>	<b>2.14 (1.01-4.62)</b>	<b>0.049</b>	<b>1.53 (1.07-2.17)</b>	<b>0.020</b>

OR: Odds ratio. STAT4: signal transducer and activator of transcription 4. Two separate logistic regression models, including a rs11889341\*smoking interaction term, were performed for MI and nephritis. <sup>1</sup>Defined according to the 1982 ACR criteria[1]. <sup>2</sup>Defined as ever-smoking, including both current and past smoking. p<0.05 (unadjusted) in bold

Supplementary table 4. Multiple regression model for myocardial infarction

	Smokers <sup>1</sup>		Non-smokers <sup>1</sup>	
	OR	p	OR	p
rs11889341 (STAT4) <sup>2</sup>	<b>3.26 (1.15-9.20)</b>	<b>0.026</b>	0.58 (0.09-3.54)	0.55
Nephritis <sup>3</sup>	0.89 (0.23-3.49)	0.87	5.54 (0.56-54.55)	0.14
Any aPL <sup>4</sup>	0.68 (0.21-2.13)	0.50	2.04 (0.18-23.50)	0.60
Age at follow-up	<b>1.09 (1.03-1.15)</b>	<b>0.0031</b>	<b>1.10 (1.02-1.20)</b>	<b>0.021</b>
Disease duration	1.03 (0.97-1.09)	0.38	1.02 (0.95-1.09)	0.64

OR: Odds ratio. STAT4: Signal transducer and activator of transcription 4. aPL: anti-phospholipid antibodies. To test whether the association between STAT4 and MI in smokers remained significant after adjusting for aPL and nephritis, a multiple regression model was performed including all patients with available data for all included variables (n=152). A separate model was performed for the non-smokers (n=170). <sup>1</sup>Defined as ever-smoker, including current and past smokers, or never-smokers. <sup>2</sup>OR per additional risk allele. <sup>3</sup>Defined according to the 1982 ACR criterion (ref). <sup>4</sup>Defined as having at least one positive aPL out of aCL (IgG or IgM), aβ<sub>2</sub>GPI (IgG or IgM) or LA. p<0.05 (unadjusted) in bold

1. Tan EM, Cohen AS, Fries JF, et al. The 1982 revised criteria for the classification of systemic lupus erythematosus. *Arthritis Rheum* 1982;25:1271-7

## Definitions

The world health organization (WHO) 1995 classification system [1] was used for the classification of renal biopsies, where mesangial nephritis was defined as class I-II, proliferative nephritis as class III-IV, membranous nephritis as class V. Patients with biopsies displaying signs of nephritis but not meeting the criteria for any of the above classes, [1] were classified as *other*. Patients who had undergone renal transplantation or were receiving dialysis were considered to have end-stage renal disease (ESRD). dsDNA positivity was defined according to the ACR-82.[2]

A clinical diagnosis of APS was made based on thrombotic events and/or miscarriage in combination with positive tests for IgG aCL, IgG a $\beta$ <sub>2</sub>GPI or LA, and classified according to Miyakis *et al.*[3] Triple-positivity for aPL was defined as having positive tests for aCL (IgG or IgM) and a $\beta$ <sub>2</sub>GPI (IgG or IgM) and LA. Myocardial infarction (MI) required confirmation by electrocardiography and a rise in plasma creatine kinase, muscle and brain fraction (CK-MB) or troponine T, and ischemic cerebrovascular disease (ICVD) by computer tomography or magnetic resonance imaging. Venous thromboembolism (VTE) was defined as deep vein thrombosis, confirmed by venography or ultrasonography, and/or pulmonary embolism, confirmed by radionuclide lung scanning or angiogram.

## Genotyping and quality control

Genomic DNA was extracted from peripheral blood of SLE patients from both the discovery and replication cohorts by using Qiagen Blood Midi Kit (Qiagen, Hilden, Germany). The discovery cohort and healthy controls were genotyped with the Illumina 200K ImmunoChip SNP array at the SNP&SEQ Technology Platform at Science for

Life Laboratory in Uppsala, Sweden. Quality control was performed sample-wise and SNP-wise. Clustering and genotype calling was performed using Illumina's GenCall software. Sample call rate below 95% and SNPs with call rate below 100% were excluded. Samples with abnormal autosomal heterozygosity rate with more than 5SD from the mean of Wright's inbreeding coefficient  $F$  were excluded. A check for mislabelled gender was performed using Wright's inbreeding coefficient  $F$ , calculated from X chromosome data. Annotated females with  $F$  close to one or annotated males with  $F$  close to zero were excluded. Cryptic relatedness was analysed using KING software[4] and one sample from each related pair (up to 2nd degree of relatives) was excluded. Furthermore, principal component analysis (PCA) was performed on 1000 Genomes Project data and then used to project and exclude study samples falling more than 5SD from European populations in each of 5 principal components. SNPs with minor allele frequency (MAF)  $< 1\%$  or with Hardy-Weinberg equilibrium (HWE)  $p$ -values within FDR  $< 5\%$  (based on controls only) were excluded, described in [5].

Genotyping of the replication cohort was performed using the iPLEX chemistry on a MassARRAY system (Agena Bioscience, San Diego, CA, USA). QC included a minimum per sample call rate of 90% and a per variant call rate of 90%. 836 Norwegian and Danish SLE patients passed QC.

## References



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