LUNG TARGETED DELIVERY OF EVEROLIMUS AS A NEW TREATMENT OF SCLERODERMA-RELATED INTERSTITIAL LUNG DISEASE (SSc-ILD) DEVELOPED BY PSGL-1 KO MICE

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Background: Interstitial lung disease (ILD), the main cause of mortality in scleroderma (SSc) patients (1), has no treatment (2). P-selectin glycoprotein ligand 1 (PSGL-1), the main ligand for P-Selectin, is expressed on leukocytes and responsible for the initial steps of extravasation (3). The absence of PSGL-1 in mice spontaneously develops an autoimmune syndrome similar to human SSc with fibrosis, vascular damage, autoantibodies and pulmonary arterial hypertension in females, and almost 60% of animals older than 12 months develop ILD with aging (4). In this work, the therapeutic action of everolimus-loaded nanomedicine given by local administration as a treatment for ILD was evaluated. The intratracheal administration of everolimus loaded into liposomes decorated with hyaluronic acid (HA) is studied as an administration strategy to reach the inflammatory areas. We aimed to target these cells and avoiding systemic effects and possible toxicity on epithelial cells.

Objectives: 1) To study the effect of everolimus on bronchoalveolar lavage (BAL) cell populations and in lung pathology in SSc-ILD PSGL-1 KO mice 2) To analyze the intratracheal application of everolimus included in empty liposomes (Lip+Ev) vs. liposomes decorated with hyaluronic acid (Lip-HA+Ev) as an administration strategy to decrease drug toxicity and increase drug effectiveness.

Methods: In an observational study, PSGL-1−/− C57BL/6 males older than 12 months (n=4) were treated intratracheally with 4 doses of Lip or Lip-HA (with or without everolimus included), once a week (Lip+Ev 295.67µg/mL; Lip+Ev 827.39g/mL; Lip-HA+Ev 82.73g/mL). Then, animals were euthanized and BAL and lungs were obtained. BAL cells were stained for flow cytometry analysis. Lungs were embedded in paraffin blocks for blind histological analysis by a pathologist and evaluated for interstitial inflammation and fibrosis degree. Lip-HA was selected as the treatment of choice for a second experiment (n=8) following the same experimental design (86.22µg/mL).

Results: The observational study showed an increase in CD45+, alveolar macrophages (AM), eosinophils (Eos), granulocytes (Gr1+) and T cells in the BAL of untreated PSGL-1−/− compared with WT mice. Everolimus reduced these populations to WT levels in all cases. Lip-HA+Ev administration was chosen for further experiments because a lower dose of the drug gave a better result than the high dose in undecorated liposomes. Reduction of CD45+, AM, eosinophils, and CD45+ cells populations by Lip-HA+Ev was confirmed. Lip-HA treatment increased the number of neutrophils and T cells, but this effect is controlled by the everolimus administration. Histological lung analysis showed an increase in interstitial inflammation and fibrosis in untreated PSGL-1−/− and empty Lip-HA experimental groups. Treatment with everolimus included in Lip-HA reduced the fibrotic and inflammatory interstitial lung lesions, reaching values similar to those observed in WT mice.

Conclusion: PSGL-1 KO mice present ILD associated with scleroderma (SSc-ILD) with an increase of CD45+, Gr1+, Eos, T cells and AM populations in the BAL. Intratracheal treatment with everolimus included in liposomes decorated with hyaluronic acid reduces immune cell infiltration and fibrosis once SSc-ILD is established.


Long non-coding RNAs (IncRNA) are a class of non-coding trancripts which modulate many biological processes. Our previous studies showed that IncRNA H19X is pivotal in the regulation of TGFβ-driven fibrosis in systemic sclerosis (SSc).

Objectives: We aimed to investigate whether H19X plays a functional role in the treatment of endothelial cell (EC) activation, which is crucial in SSc vasculopathy.

Methods: Single-cell RNA sequencing (scRNA-seq) data from 27 dcSSc and 10 healthy control (HC) skin biopsies, following 10X Genomics partitioning and cdDNA preparation, were analyzed for H19X expression in skin ECs, using Seurat package in R. A total of 4,981 ECs, of which 1,583 cells originated from HC and 3,396 cells originated from SSc patients, ranging from 59 to 342 ECs per subject, characterized by enrichment of EC markers of CD31, VWF and PECAM1. Expression of H19X in Human Dermal Microvascular ECs (HDMEC) was analyzed by qPCR. HDMEC were stimulated with different proinflammatory cytokines including IFNγ, IFNβ, TGFβ, TNFα, IL-6, IL-11 and IL-4 at biologically relevant concentrations. In order to ascertain its effect in ECs, H19X was silenced in HDMECs using locked nucleic acid antisense oligonucleotides (LNA Gapmers). IC50, EC50 and IC80 for Westergen stimulate were used to analyze the effects of H19X down-regulation on EC activation biomarkers.

Results: scRNA-seq data showed that H19X was significantly upregulated in SSc compared to healthy ECs (p=0.0095). Based on the differentially expressed gene profiles among subclusters, ECs were further annotated as arterial (SEMA3G, HEY1), capillary (CA4, RGCC), venous (ACKR1, VCAM1), lymphatic (PROX1, VWF) ECs, as well as two aberrant clusters, proliferating (TOP2A, MK Rex7) and injured (HSGP2, APLMR) ECs, which were dominated by the SSc ECs. Specifically, the highest expression of H19X was found in injured SSc ECs and capillary SSc ECs. Overall, 15% SSc EC, about 51 cells, expressed detectable levels of H19X. In HDMEC (n=3), H19X was consistently induced by IFNγ, IFNβ and IFNα. Time curve analysis demonstrated that the strongest induction was observed at 48h (1.5±0.2, 1.6±0.4 and 2.1±0.3 – fold increase respectively). The combination of different IFNαs determined stronger H19X induction after 48h stimulation, with a 2.4±0.1 increase with the combination of all IFNs and a 2.4±0.1 increase after the combination of IFNα+γ. Importantly, H19X knockdown lead to consistent and significant decrease of mRNAs of several adhesion molecules, including VCAM1, E- Selectin and P-Selectin, both in untreated HDMEC and after IFN stimulation. A decrease of VCAM and P-Selectin could be also demonstrated with WB analysis. No change was seen in other EC activation markers, including endothelin-1 and angiogenesis markers including VEGF, VEGFRA, Tie2 and thrombospondin.

Conclusion: This is the first report analyzing a potential role of IncRNA H19X in SSc vasculopathy. Our results suggest that IncRNA H19X could act as a regulator of adhesion molecules expression in EC, possibly mediated by IFNs, and be involved in EC activation.


Disclosure of Interests: Francesco Trelle: None declared, Elena Pacher: None declared, Robert Lafayatis Consultant of: RL served as a consultant with Bristol Meyers Squib, Formation, Sanofi, Boehringer-Ingelheim, Merck, and Genentech/Roche, Acceleron, Grant/research support from: RL received grants form Bristol Meyers Squib, Corbus, Formation, Moderna, Regeneron, Pfizer, and Kiniksa, Merci Huang: None declared, Francia Zarock: None declared, Olivier Distler Speakers bureau: Speaker fee on Scleroderma treatments for systemic sclerosis and its complications (last three years):Abbvie, Actelion, Acceleron Pharma, Amgen, AnaMar, Arxx Therapeutics, Bayer, Baecon Discovery, Blade Therapeutics, Boehringer, CSL Behring, ChemomAb, Corbus Pharma, Curzion Pharmaceuticals, Ergonex, Galapagos NV, GSK, Genmack Pharmaceuticals, Inventiva, Italfarmaco, iQvia, Kymera, Medac, Medscape, Mitsubishi Tanabe Pharma, MSD, Roche, Sanofi, UCBI, Lilly, Target BioScience, Pfizer. Consultant relationship for rheumatology topic other than Scleroderma:
Abbvie, Amgen, Lilly, Target BioScience, Pfizer. In addition, he holds a patent issued "mir-29 for the treatment of systemic sclerosis" (US8247389, EP2331143). Grant/research support from: Hss received grants/research support in the area of potential treatments for systemic sclerosis from Actelion, Bayer, Boehringer Ingelheim, Kymera Therapeutics, Mitsubishi Tanabe Pharma.

DOI: 10.1136/annrheumdis-2021-eular.3028

**POS0333 ALTERED MACROPHAGE POLARIZATION PHENOTYPES IN SYSTEMIC SCLEROSIS**

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Background: Fos-like 2 (Fosl-2) is a transcription factor of the AP-1 family and has a broad range in inducing cellular changes affecting fibrosis and inflammatory responses. Pathological effects of Fosl-2 have been associated with systemic sclerosis (SSc). In addition, increased expression of Fosl-2 has been detected in human SSc monocyte-derived macrophages [1]. Monocytes and macrophages play a central role in activating and propagating acute inflammatory diseases following pathological fibrosis and organ dysfunction. The classification of the macrophage polarization phenotype can be assigned based on the stimulus, for example, classically-activated M(1) (M1) or alternatively-activated M(2) (M2) macrophages [2]. However, the role of the Fosl-2 transcription factor in macrophage polarization remains elusive.

**Objectives:** To investigate the role of Fosl-2 in macrophage polarization in SSc using Fosl-2 overexpressing transgenic (Fosl-2 tg) mice and human blood-derived macrophages from SSc patients.

**Methods:** Thioglycolate-elicited peritoneal macrophages were isolated from wild-type (wt) and Fosl-2 tg mice. Human peripheral CD14+ blood-derived monocytes were isolated and differentiated to macrophages (hMDM) from healthy controls and SSc patients. Murine and human macrophages were polarized with LPS (10 ng/ml), LPS + recombinant mouse IFN-γ (10 ng/ml), recombinant mouse, resp, human IL-4 (10 ng/ml) or remained untreated. Macrophage surface marker expression was assessed by flow cytometry using a mouse (F4/80, CD11b, CD86, CD163, MHCIІІ, CD206, PD-L1, PD-L2, CD36) or human (CD38, CD40, CD86, PD-L2, PD-L1, CD163, CD206) determined polarization panel. Phagocytic activity was determined with phthodo Red E-cell particles by flow cytometry. Gene expression and secretion of pro- and anti-inflammatory markers were measured by RT-qPCR, standard ELISAs and Griess Assay for nitric oxide production.

**Results:** After LPS stimulation, mRNA levels of IL-1β (p<0.01, n=11-12), TNF-α (p<0.05, n=11-12) and IFN-γ (p<0.05, n=7) were reduced, whereas expression of IL-10 (p<0.05, n=11-12) was enhanced in Fosl-2 tg peritoneal macrophages in comparison to wt cells. Secretion of TNF-α (p<0.01, n=9-11) and nitric oxide (p<0.01, n=9) was impaired in Fosl-2 tg peritoneal macrophages compared to wt cells after LPS stimulation. Peritoneal macrophages were analyzed directly after isolation for macrophage polarization cell surface marker expression. Fosl-2 tg peritoneal macrophages showed an increase in the F4/80+PD-1+CD236+ cell population (p<0.01, n=3-6) compared to peritoneal macrophages from wt mice. The expression of cell surface markers of non-polarized and L-4 stimulated SSc hMDM (n=17) showed an increased percentage of CD40-CD86-CD206+PD-L2+CD163- cells (p<0.05) compared to healthy control hMDM (n=7). Phagocytic activity was enhanced in SSc hMDM (n=7) compared to healthy untreated (p<0.05), LPS (p<0.05) and IL-4 (p<0.05) hMDM (n=5).

**Conclusion:** Our animal data indicates a role of Fosl-2 in regulating macrophage polarization with a shift from a classically-activated to an alternatively-activated phenotype. Similarly, SSc hMDM resembles a functional M(IL-4) alternative macrophage phenotype. Thus, maintaining a balanced proportion of classically- and alternatively-activated macrophage phenotypes may be an effective tool to control macrophage function in SSc.

**REFERENCES:**


**Disclosure of Interests:** Amela Hukara: None declared, Michal Rudnik: None declared, Chantal Brigitte Ruffer: None declared, Oliver Distler Speakers bureau: Actelion, Bayer, Boehringer Ingelheim, Medpace, Novartis, Roche, Mepha, MSD, IQone, Pfizer, Consultant: of: Abbvie, Actelion, Acceleron Pharma, Amgen, AnaMar, Anxx Therapeutics, Bayer, Baecoe Discovery, Blade Therapeutics, Boehringer, CSL Behring, ChemobAb, Corpupharma, Curzon Pharmaceuticals, EndoGenex, Galapagos NV, GSK, Glenmark Pharmaceuticals, Inventiva, Italfarmaco, IQvia, Kymera, Medac, Medpace, Mitsubishi Tanabe Pharma, MSD, Roche, Sanofi, UCB, Lilly, Target BioScience, Pfizer, Grant/research support from: Actelion, Bayer, Boehringer Ingelheim, Kymera Therapeutics, Mitsubishi Tanabe, Przemyslaw Blyszczuk: None declared, Gabriela Kania: None declared.

DOI: 10.1136/annrheumdis-2021-eular.3124

**POS0334 INTERSTITIAL LUNG INFLAMMATION WITH ALVEOLAR HEMORRHAGE IN AUTOIMMUNE MRL/Mp-lpr/lpr MICE**

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**Background:** Anti-melanoma differentiation-associated gene 5 (MDA5) antibody is associated with interstitial lung disease (ILD) in patients with juvenile dermatomyositis (JDM). Although the mechanisms leading to pulmonary involvement remain uncertain, both inflammatory cytokines and autoimmune responses between MDA5 and anti-MDA5 antibody could be inferred. The present study examined the roles of MDA5 in an inducible form of lung involvement that develops in autoimmune mice treated with the pristane.

**Objectives:** MRL/Mp-lpr/lpr mice and wild type controls (+/+) at 5 weeks of age. They received 0.5ml of pristane or an equal volume of PBS IP at the age 7 week (day 1). And 1.25g of recombinant human MDA5 protein (rMDA5) was instilled IP or PBS at day2 and Day9. The mice were sacrificed on 8 weeks after the intraperitoneal injection. Lung tissue was harvested for histological assessment.

**Methods:** MRL/Mp-lpr/lpr mice and wild type controls (+/+) at 5 weeks of age. They received 0.5ml of pristane or an equal volume of PBS IP at the age 7 week (day 1). And 1.25g of recombinant human MDA5 protein (rMDA5) was instilled IP or PBS at day2 and Day9. The mice were sacrificed on 8 weeks after the intraperitoneal injection. Lung tissue was harvested for histological assessment.

**Results:** Lung involvements did not develop in PBS-treated MRL/lpr mice and WT mice. And Lung involvements did not develop in MRL/lpr mouse treated with pristane. H&E staining of lung tissue from MRL/lpr mice and WT mouse with pristane that showed induced bland alveolar hemorrhage. H&E staining of lung from WT mouse compared with lung from a MRL/lpr mouse treated with pristane + rMDA5 protein showing mild thickening of the alveolar septa despite the alveolar hemorrhage. And perivascular lymphocytes infiltrate in a MRL/lpr mouse rather than a WT mouse treated with pristane + rMDA5 protein. CD163 staining of alveolar macrophages were present in the alveolar spaces was more intense in mouse treated with pristane + rMDA5 protein than in mouse treated with only pristane. The Lymphocyte infiltrations around alveolar macrophages was more prominent in MRL/lpr mouse treated with pristane + rMDA5 protein than other mouse.

**Conclusion:** These results suggest that lung involvements such as the alveolar hemorrhage, are caused by pristane and MDA5 protein in the pathogenesis of interstitial pneumonia. MRL/lpr mouse treated with pristane + rMDA5 protein showed more alveolar macrophages that had lymphocyte infiltrations. After the alveolar hemorrhage by pristane, the antigen exposure of MDAS might induce continuously inflammatory response to lymphocytes and macrophages in the alveolar lesions.

**REFERENCES:**


**Figure 1. Lung disease (A-D). Light microscopy. Lung tissue from mice 8 weeks after treatment were stained with H&E. (A) Pristane-treated WT mice with almost no alveolar lesions. (B) Pristane + MDA5 protein-treated WT mice with alveolar hemorrhage and slight lymphocyte infiltrations. (C) Pristane-treated MRL/lpr mouse with almost no alveolar lesions and slight lymphocyte infiltrations. (D) Pristane-treated MRL/lpr mouse with alveolar hemorrhage and lymphocyte infiltrations, thickening of the alveolar septa.**

**Disclosure of Interests:** None declared.

DOI: 10.1136/annrheumdis-2021-eular.3411