

The level of peripheral regulatory T cells is linked to changes in gut commensal microflora in patients with systemic lupus erythematosus

Systemic lupus erythematosus (SLE) is a multisystem autoimmune disorder mainly mediated by lymphocytes and autoantibodies, which have a pervasive negative impact on the majority of organs.^{1,2} It deserves more attention to further explore the pathogenesis because of the unclear complex pathogenesis and the limited clinical efficacy of SLE treatment.

Recently, a cross-sectional discovery cohort in the USA conducted by Azzouz *et al*, published in *Annals of Rheumatic Diseases*, suggested that specific gut commensal strains, especially *Ruminococcus gnavus*, may contribute to the disease activity and autoantibody production in SLE patients,³ which put forward a novel concept for the immune pathogenesis of SLE. Their study was well performed and analysed the abundance of intestinal flora and sera profiled for antibacterial and autoantibody responses between SLE patients with the different disease activity index scores and matched healthy controls. However, the faecal microbiota is lacking a precise link with lymphocytes in peripheral blood of the patients, which are the important participants of immune mechanism of SLE.

In this study, we studied the correlation between the changes in faecal microbial diversity and the absolute numbers of peripheral lymphocyte subgroups and CD4+ T subsets in SLE patients, especially regulatory T cells (Tregs) that mediate immune tolerance and maintain immunological homeostasis.⁴ The blood and stool samples were collected from 92 patients with SLE and 217 matched healthy adults. The 16S rRNA in the stool specimens were sequenced using the Roche/45 high-throughput sequencing platform. The absolute numbers of circulating lymphocytes and CD4+ T subgroups of these individuals were detected by flow cytometry combined with standard absolute counting beads.⁵

Patients with SLE, regardless male or female, had different taxonomic diversity and abundance of specific strains of a gut commensal at the level of the phylum, family and genus ($p < 0.05$) from healthy controls. They had higher levels of Proteobacteria, Bacteroidetes and Actinobacteria and a lower level of Firmicutes as compared with those of healthy controls at the phylum level ($p < 0.05$). In addition, at the family or genus level, the proportion or abundance of gut bacteria in SLE patients, including Bacteroidaceae, Veillonellaceae, *Klebsiella*, Streptococcaceae and Erysipelotrichaceae, differed from those in healthy individuals with statistically significant difference ($p < 0.05$) (table 1). It was noteworthy that patients with SLE had significantly lower proportion of Ruminococcaceae at family level than healthy controls ($p < 0.001$). Interestingly, the percentage of *Ruminococcus* at genus level was higher in SLE patients ($p < 0.05$), which was confirmed to be involved in the incidence of lupus in Azzouz *et al*'s study, suggesting that we should study the related bacterial flora and its mechanism of SLE patients at multiple levels.

We found that the proportion of *Ruminococcus* was correlated with the absolute counts of lymphocytes, suggesting that the changed of intestinal flora was involved in the imbalance of proinflammation and anti-inflammation T cells in SLE. Particularly, the proportion of *Ruminococcus* was significantly correlated with the absolute counts of Tregs (its dysfunction was one of the crucial immune mechanisms in the onset of lupus)⁶ and the ratio of Th1/Th2 and Th17/Treg (figure 1D–F), but not with the numbers of Th1, Th2 and Th17 cells (figure 1A–C), which may be one of the reasons for the changes in intestinal

Table 1 Shifts in taxonomic abundance between SLE and healthy controls (HC) (mean±SD)

Taxonomy	SLE (%)	HC (%)	P value
Phylum			
Proteobacteria	13.26±21.77	2.65±3.56	<0.001
Firmicutes	40.14±25.58	59.20±19.19	<0.001
Bacteroidetes	42.69±25.26	35.75±18.58	0.018
Actinobacteria	2.25±3.49	1.26±3.28	0.002
Family			
Lachnospiraceae	16.35±13.09	25.09±12.89	<0.001
Ruminococcaceae	12.72±12.68	23.50±13.57	<0.001
Bacteroidaceae	35.12±25.56	26.67±18.03	0.021
Veillonellaceae	2.63±5.42	5.68±9.30	<0.001
Streptococcaceae	1.26±5.56	0.24±1.49	<0.001
Genus			
<i>Ruminococcus</i>	2.63±6.59	1.41±2.04	0.016
<i>Haemophilus</i>	0.15±0.53	0.17±0.59	0.035
<i>Faecalibacterium</i>	7.58±9.80	19.99±13.87	<0.001
<i>Bacteroides</i>	38.75±26.96	30.61±19.8	0.016
<i>Clostridium</i> IV	0.17±0.40	0.27±0.51	<0.001
<i>Klebsiella</i>	3.04±9.39	0.30±1.54	0.001
Erysipelotrichaceae	0.06±0.32	0.02±0.13	<0.001

SLE, systemic lupus erythematosus.

microbial population are involved in SLE patients. However, there was no obvious correlation between the abundance of *Ruminococcus* and the absolute numbers of total T, B, natural killer (NK), CD4+ T and CD8+ T cells ($p > 0.05$).

In conclusion, the changes in taxonomic diversity and the abundance of partial specific flora, such as *Ruminococcus* genus, may regulate the absolute number of Tregs in peripheral blood to participate in the pathogenesis of SLE. This study on the correlation between the intestinal flora abundance and levels

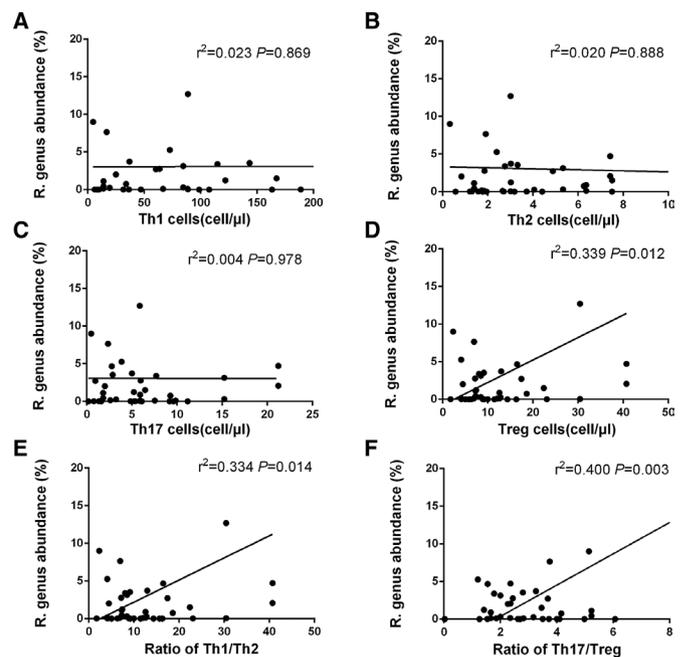


Figure 1 Correlation of absolute numbers of CD4+ T subsets and the ratio of Th1/Th2 and Th17/Treg with the proportion of *Ruminococcus* at genus level in SLE patients by Spearman coefficient. All p values reported herein are two-tailed. $P < 0.05$ was taken as statistical significance. SLE, systemic lupus erythematosus.

of lymphocyte subsets reconfirmed the involvement of gut commensal microflora in the pathogenesis of SLE.

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