TRANSLATIONAL SCIENCE

Methotrexate hampers immunogenicity to BNT162b2 mRNA COVID-19 vaccine in immune-mediated inflammatory disease

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ABSTRACT

Objective To investigate the humoral and cellular immune response to messenger RNA (mRNA) COVID-19 vaccines in patients with immune-mediated inflammatory diseases (IMIDs) on immunomodulatory treatment.

Methods Established patients at New York University Langone Health with IMID (n=51) receiving the BNT162b2 mRNA vaccination were assessed at baseline and after second immunisation. Healthy subjects served as controls (n=26). IgG antibody responses to the spike protein were analysed for humoral response. Cellular immune response to SARS-CoV-2 was further analysed using high-parameter spectral flow cytometry. A second independent, validation cohort of controls (n=182) and patients with IMID (n=31) from Erlangen, Germany, were also analysed for humoral immune response.

Results Although healthy subjects (n=208) and patients with IMID on biologic treatments (mostly on tumour necrosis factor blockers, n=37) demonstrate robust antibody responses (over 90%), those patients with IMID on background methotrexate (n=45) achieve an adequate response in only 62.2% of cases. Similarly, patients with IMID on methotrexate do not demonstrate an increase in CD8+ T-cell activation after vaccination.

Conclusions In two independent cohorts of patients with IMID, methotrexate, a widely used immunomodulator for the treatment of several IMIDs, adversely affected humoral and cellular immune response to COVID-19 mRNA vaccines. Although precise cut-offs for immunogenicity that correlate with vaccine efficacy are yet to be established, our findings suggest that different strategies may need to be explored in patients with IMID taking methotrexate to increase the chances of immunisation efficacy against SARS-CoV-2 as has been demonstrated for augmenting immunogenicity to other viral vaccines.

INTRODUCTION

Patients with immune-mediated inflammatory diseases (IMIDs) have an inherently heightened susceptibility to infection and may thus be considered high risk for developing COVID-19. Importantly, however, the strength of response to viral vaccines (ie, influenza and hepatitis B) and their long-lasting protective effects in patients with IMID taking conventional disease-modifying antirheumatic drugs (DMARDs), such as methotrexate, or biologic DMARDs, such as tumour necrosis factor inhibitors (TNFis), may not be as robust...
as it is in the general population following immunisation.\textsuperscript{1-5} Data regarding messenger RNA (mRNA) COVID-19 vaccines’ safety, immunogenicity and efficacy are rapidly emerging for the immunocompetent adult population,\textsuperscript{6} where more than 90% of subjects achieve a satisfactory humoral response. However, the ability of patients with IMID to adequately respond to these vaccines and the differences in humoral and cellular immune response to SARS-CoV-2 vaccine are not known, leaving a significant gap in knowledge that prevents optimal management of this patient population.

Given the experience with seasonal influenza vaccine immunogenicity,\textsuperscript{2,7} we hypothesised that patients with IMID treated chronically with certain conventional synthetic DMARDs (ie, methotrexate) would have an attenuated response to mRNA COVID-19 vaccines compared with patients with IMID receiving anticytokine treatment or non-IMID participants. To achieve this, we obtained preimmunisation and postimmunisation peripheral blood monocyte cells (PBMCs) and sera from IMID participants (n=82) in two independent cohorts (SAGA (Serologic Testing and Genomic Analysis of Autoimmune, Immune-Mediated and Rheumatic Patients with COVID-19) cohort in New York City, USA, and Erlangen, Germany) and analysed SARS-CoV-2 spike-specific antibody titres compared with non-IMID controls (n=208). Cellular immune responses were further investigated using high-dimensional spectral flow cytometry in the New York City cohort.

METHODS

Participants

Established patients with IMID (n=51) receiving methotrexate, anticytokine biologics or both participating in the SAGA study at New York University Langone Health in New York City,\textsuperscript{8} who were receiving BNT162b2 mRNA vaccination were assessed at baseline and after the second dose during the period from 23 December 2020 through 31 March 2021. Healthy subjects served as controls (n=26). IgG antibody responses to the S protein were analysed for humoral immune response. A second independent validation cohort of controls (n=182) and patients with IMID (n=31) on either TNFi or methotrexate monotherapy from the Erlangen validation cohort in New York City, USA, and Erlangen, Germany, was also analysed for humoral response. Cellular immune responses to the vaccine were also studied for the New York SAGA participants using high-parameter spectral flow cytometry.

Humoral and cellular immune response to BNT162b2 mRNA vaccine

Humoral immune response was assessed by testing IgG antibodies against the spike protein of SARS-CoV-2.\textsuperscript{9} In the New York City cohort, direct ELISA was used to quantify antibody titres on serum as previously described.\textsuperscript{10} Titre of 5000 units or greater was used as the cut-off to determine an adequate response to vaccination. IgG antibodies against the S1 domain of the spike protein of SARS-CoV-2 were tested in Erlangen participants using the commercial ELISA from Euroimmun (Lübeck, Germany) on the EUROMMUN Analyzer I platform and according to the manufacturer’s protocol.\textsuperscript{11} Adequate response was defined as greater than 5.7 nm OD. Immune cell phenotyping before and after immunisation in New York participants was performed by 35-colour spectral flow cytometry on PBMCs. Further details on methodology and analysis can be found in the online supplemental appendix.

Statistical analysis

Patient characteristics were summarised using means, medians, SD, ranges and percentages as appropriate. \( \chi^2 \) tests of independence and Fisher’s exact tests were used for categorical data. Mann-Whitney U and Kruskal-Wallis tests were used for unpaired continuous data, and Wilcoxon signed-rank tests were used for paired continuous data. A p value of less than 0.05 was considered significant. All analyses were done using R V.3.6.0 software (R Foundation for Statistical Computing) and GraphPad Prism V9 (GraphPad Software).

Patient and public involvement

This study was designed in response to frequent questions asked by patients with IMID but did not contain any direct public involvement.

RESULTS

The New York City cohort comprised 26 healthy individuals, 25 individuals with IMID receiving methotrexate monotherapy or in combination with other immunomodulatory medications, and 26 individuals with IMID on anticytokine therapy and/or other oral immunomodulators (table 1). Healthy individuals and those with IMID not on methotrexate were similar in age (49.2±11.9 years and 49.1±14.9 years, respectively), whereas patients with IMID receiving methotrexate were generally older (63.2±11.9 years). IMID diagnoses were predominantly psoriasis/psoriatic arthritis and rheumatoid arthritis. The Erlangen cohort consisted of 182 healthy subjects, 11 subjects with IMID receiving TNFi monotherapy and 20 subjects with IMID on methotrexate monotherapy (online supplemental table 1). Individuals on methotrexate monotherapy were on average older than healthy individuals and those with IMID not on methotrexate (54.5±19.2 vs 40.8±12.0 and 45.0±15.5, respectively).

Decreased antibody response to mRNA COVID-19 vaccine in patients with IMID on methotrexate

Immunogenicity was characterised by testing IgG antibodies against the spike protein of SARS-CoV-2. In the New York City cohort, of the healthy participants, 25 (96.1%) of 26 demonstrated adequate humoral immune response. Patients with IMID not on methotrexate achieved a similar rate of high antibody titres (24/26, 92.3%), whereas those on methotrexate had a lower rate of adequate humoral response (18/25, 72.0%) (figure 1A; table 1). This remains true even after the exclusion of patients who had evidence of previous COVID-19 infection (p=0.045). Median titres were 104354 (range, 141–60185), 113608 (25–737310) and 46901 (25–694528) for participants who were healthy, for those with IMID not on methotrexate and for those with IMID on methotrexate, respectively. Similarly, in the Erlangen validation cohort, 179 (98.3%) of 182 healthy controls, 10 (90.9%) of 11 patients with IMID receiving no methotrexate and 10 (50.0%) of 20 receiving methotrexate...
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achieved adequate immunogenicity (figure 1B). Median ODs for this cohort were 9.4 (range, 1.2–14), 7.8 (2.3–11.3) and 5.9 (0.95–13.5) for participants who were healthy, for those with IMID not on methotrexate and for those with IMID on methotrexate, respectively. Furthermore, when looking at the two cohorts in conjunction (n=290), 204 (98.1%) of 208 healthy controls, 34 (91.9%) of 37 patients with IMID receiving no methotrexate and 28 (62.2%) of 45 receiving methotrexate achieved adequate immunogenicity (p<0.001) (online supplemental figure S1).

Because of the imbalance in age between groups, we further analysed immunogenicity based on a cut-off age of 55. In both

Table 1  Baseline characteristics and spike-specific SARS-CoV-2 antibody titres in the New York City cohort

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Healthy (n=26)</th>
<th>IMID No MTX (n=26)</th>
<th>IMID Yes MTX (n=25)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, mean (range, SD)</td>
<td>49.2 (28–74, 11.9)</td>
<td>49.1 (29–79, 14.9)</td>
<td>63.2 (22–77, 11.9)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Female, n (%)</td>
<td>16 (61.5)</td>
<td>18 (69.2)</td>
<td>18 (66.7)</td>
<td>0.352</td>
</tr>
<tr>
<td>Race, n (%)</td>
<td>0.220</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>White</td>
<td>16 (61.5)</td>
<td>20 (76.9)</td>
<td>17 (63.0)</td>
<td></td>
</tr>
<tr>
<td>Black</td>
<td>1 (3.8)</td>
<td>2 (7.7)</td>
<td>3 (11.1)</td>
<td></td>
</tr>
<tr>
<td>Asian</td>
<td>9 (34.6)</td>
<td>3 (11.5)</td>
<td>3 (11.1)</td>
<td></td>
</tr>
<tr>
<td>Other</td>
<td>0 (0.0)</td>
<td>1 (3.8)</td>
<td>2 (7.4)</td>
<td></td>
</tr>
<tr>
<td>Hispanic ethnicity, n (%)</td>
<td>1 (3.8)</td>
<td>3 (11.5)</td>
<td>5 (18.5)</td>
<td>0.200</td>
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<tr>
<td>Primary IMID, n (%)</td>
<td>0.107</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Psoriasis and/or psoriatic arthritis</td>
<td>--</td>
<td>15 (57.7)</td>
<td>9 (36.0)</td>
<td></td>
</tr>
<tr>
<td>Rheumatoid arthritis</td>
<td>--</td>
<td>10 (38.5)</td>
<td>12 (48.0)</td>
<td></td>
</tr>
<tr>
<td>Other*</td>
<td>--</td>
<td>1 (3.8)</td>
<td>4 (16.0)</td>
<td></td>
</tr>
<tr>
<td>Long-term medication, n (%)</td>
<td>--</td>
<td>0 (0.0)</td>
<td>25 (100.0)</td>
<td></td>
</tr>
<tr>
<td>Methotrexate</td>
<td>--</td>
<td>11 (42.3)</td>
<td>9 (36.0)</td>
<td>0.776</td>
</tr>
<tr>
<td>Tumour necrosis factor inhibitor</td>
<td>--</td>
<td>9 (34.6)</td>
<td>1 (4.0)</td>
<td>0.011</td>
</tr>
<tr>
<td>Other anticytokines/Janus kinase inhibitors†</td>
<td>--</td>
<td>7 (26.9)</td>
<td>6 (24.0)</td>
<td>1.00</td>
</tr>
<tr>
<td>Other oral immunomodulators†</td>
<td>--</td>
<td>25 (96.1)</td>
<td>18 (72.0)</td>
<td>0.023</td>
</tr>
<tr>
<td>Methotrexate dose, mean (SD)</td>
<td>--</td>
<td>25 (100.0)</td>
<td>25 (100.0)</td>
<td>1.00</td>
</tr>
<tr>
<td>COVID-19 infection before vaccination, n (%)</td>
<td>4 (15.4)</td>
<td>5 (19.2)</td>
<td>2 (8.0)</td>
<td>0.509</td>
</tr>
<tr>
<td>Days from first vaccination dose, mean (range, SD)</td>
<td>29.0 (23–44, 4.6)</td>
<td>32.5 (25–45, 5.0)</td>
<td>34.6 (21–73, 9.9)</td>
<td>0.002</td>
</tr>
<tr>
<td>Number receiving second vaccination dose, n (%)</td>
<td>26 (100.0)</td>
<td>26 (100.0)</td>
<td>25 (100.0)</td>
<td>1.00</td>
</tr>
<tr>
<td>Adequate humoral response§¶, n (%)</td>
<td>25 (96.1)</td>
<td>24 (92.3)</td>
<td>18 (72.0)</td>
<td>0.023</td>
</tr>
<tr>
<td>Spike-specific SARS-CoV-2 antibody titres¶</td>
<td>0.294</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Vasculitis, dermatomyositis, adult-onset Still’s disease, sarcoidosis and polymyalgia rheumatica.
†For IMID No MTX: IL-17i (3), IL-23i (2), abatacept (1), rituximab (1), JAKi (2). For IMID Yes MTX: IL-17 (1).‡For IMID No MTX: leflunomide (2), oral steroid (1), sulfasalazine (2), apremilast (1), hydroxychloroquine (1). For IMID Yes MTX: oral steroid (2), sulfasalazine (2), hydroxychloroquine (2).
§Adequate humoral response defined as greater than 5000 units.
¶All values 1 week after second vaccination.
IMID, immune-mediated inflammatory disease; MTX, methotrexate.

Figure 1  Anti-SARS-CoV-2 IgG levels in cohorts from New York City (A) and Erlangen (B) in healthy participants without IMID (blue), patients with IMID not receiving MTX (green) and patients with IMID treated with MTX (yellow). Solid lines represent mean titre of each group. For the New York City cohort (A), adequate response is defined as greater than 5000 units, and for the Erlangen cohort (B), adequate response is defined as greater than 5.7 (OD, 450 nm), 2 SDs of the mean of controls. Percentages and group comparisons using χ² test of independence reflect proportion of those achieving an adequate response within each group. * indicates p value less than .05 and ** indicates p value less than .001. IMID, immune-mediated inflammatory disease; MTX, methotrexate.
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Figure 2 Immune cell populations from the New York City cohort by high spectral flow in healthy controls (blue, n=20), patients with immune-mediated inflammatory disease (IMID) not on methotrexate (MTX; green, n=24) and patients with IMID on MTX (yellow, n=18), at baseline and after the second dose of BNT162b2 mRNA vaccine. Prevaccination and postvaccination comparisons were performed using Wilcoxon signed-rank tests. Y-axes presented as a logarithmic scale. NS indicates no statistical significance. * indicates p value less than .05. ** indicates p value less than .01. *** indicates p value less than .001. Tfh, T follicular helper.

Lack of CD8+ T-cell activation in patients with IMID on methotrexate following mRNA COVID-19 vaccine

In the New York City cohort, 20 healthy controls, 24 patients with IMID not receiving methotrexate and 18 patients with IMID who were receiving methotrexate underwent immune cell phenotyping before and after vaccination. The proportions of spike-specific B cells, circulating T follicular helper (cTfh; CD4+ ICOS+ CD38+ subset) cells, activated CD4+ T cells and HLA-DR+ CD8+ T cells increased significantly in all groups after immunisation (figure 2A–D). Activated CD8+ T cells, defined as CD8+ T cells expressing Ki67 and CD38, and the granzyme B-producing (GZMB) subset of these activated CD8+ T cells were induced in healthy adults and participants with IMID not on methotrexate, but not induced in patients receiving methotrexate (figure 2E,F).

DISCUSSION

In two geographically independent cohorts of patients with IMID, we found that methotrexate, a widely used immunomodulator for the treatment of several IMIDs, adversely affected humoral and cellular immunogenicity to COVID-19 mRNA vaccines.

For humoral immunity, the BNT162b2 mRNA vaccines did not induce adequately elevated SARS-CoV-2 spike-specific IgG antibody titres in up to a third of the patients on methotrexate, compared with patients with IMID on other DMARDs, who demonstrated a response as robust as that of healthy controls. This finding was analogous to the previously described effects of methotrexate on influenza vaccine immunogenicity.5 12–14 While a recent report has shown no differences in immunogenicity for patients with IMID, none of the included participants were on methotrexate.15 A second study in patients with self-reported rheumatic and musculoskeletal diseases recruited via social media showed that 10 of 13 participants on background methotrexate had detectable antibody levels after only one dose of SARS-CoV-2 mRNA vaccine,16 although this was both underpowered and used a semiquantitative ELISA measuring antibodies against SARS-CoV-2 receptor-binding domain. Therefore, the findings from our work looking at antibody responses in patients with IMID after full vaccination regimen are of potentially high clinical relevance because it was recently shown that a temporary discontinuation of methotrexate for 2 weeks significantly improved influenza vaccine immunogenicity in patients with rheumatoid arthritis.7

Importantly, the use of high-dimensional spectral flow cytometry allowed for the interrogation of specific cellular immune responses before and after immunisation. Spike-specific B cells, activated CD4+ T cells and cTfh cells were induced similarly in all groups after mRNA vaccination. In contrast, activated CD8+ T-cell responses were notably attenuated in the methotrexate cohort. Moreover, the poor induction of activated CD8+ T cells expressing granzyme B may indicate reduced cytotoxic functionality of these cells. Indeed, CD8+ T-cell responses were identified to be a correlate of protection in non-human primate studies.
of SARS-CoV-2 infection.17 Thus, reduced induction of cytotoxic CD8+ T-cell responses, combined with inconsistent induction of antibody responses, may further impair the effectiveness of COVID-19 vaccines and render patients with IMID on methotrexate at risk of inadequate vaccine response. However, this finding requires a cautious interpretation as it is quite possible that the use of methotrexate may delay (rather than prevent) adequate cellular mediated immunity against SARS-CoV-2. While spike-specific T-cell immunity has been detected as early as 10 days following one dose of mRNA COVID-19 vaccines in healthy individuals,18 mRNA-1273-specific CD4+ and CD8+ T-cell responses were most robustly elicited 2 weeks after the second dose.19 Therefore, more detailed and comprehensive studies that include long-term characterisation of the dynamics of cellular responses to these vaccines will be required to understand the clinical implications of these findings.

Although our analysis was limited in sample size, followed participants with biosampling for a relatively short period of time without standardised disease activity status metrics and was restricted to one type of mRNA immunisation, our findings were validated in an independent cohort and revealed that methotrexate, which is widely used for many indications, adversely affected the humoral and cellular immunogenicity to COVID-19 mRNA vaccination. Furthermore, because of the inclusion of patients with prior COVID-19 infection, it is possible that results could be biased in favour of those not on methotrexate. However, when excluding all patients with prior infection, the results remained similar. We also acknowledge that there may have been participants with asymptomatic COVID-19 infection that we have not captured.

While immunosenescence may reduce the level of antibody responses to immunisations,20 recent studies on COVID-19 mRNA vaccines have not shown differences in clinical outcomes for the older population.6 In our study, patients with IMID on methotrexate were generally older, which may potentially explain some differences in immunogenicity. However, even when looking at participants younger than 55 years, decreased rates of humoral response were still significant. Further validation in even larger cohorts that address efficacy will be required to understand the interaction between age and methotrexate in the context of COVID-19 vaccination.

Importantly, it is not yet clear what level of immunogenicity is representative of vaccine efficacy (and this includes the arbitrary cut-offs chosen for our measurements). We recognise that the definition of adequate cellular and humoral immune response may need to be refined in the future when correlation with efficacy becomes available. However, even after applying more conservative cut-offs, the hampering effects of methotrexate on immunogenicity are still evident.

Taken together, our results suggest that the optimal protection of patients with IMID against COVID-19 will require further studies to determine whether additional doses of vaccine, dose modification of methotrexate or even temporary discontinuation of this drug can boost immune response as has been demonstrated for other viral vaccines in this patient population.7

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Contributors RHH and JUS designed the New York study, designed the data collection tools, analysed and cleaned the data, and drafted and revised the paper. RH, MS and MM designed the New York study, designed and performed the cellular analysis, and revised the paper. SA designed the New York Study and revised the paper. RB designed the New York study, acquired data and revised the draft. DS, RA, KT, MN and GS designed the Erlangen study, designed the data collection tools, analysed and cleaned the data, and revised the paper. SA aided in original design, statistical analysis and revised the paper. MT, SK, RA and AC analysed data and revised the draft. RC, PR, GS, NA, PR, PI, JS, BG and SMR helped accrue data and revised the draft.

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Competing interests JUS declares that he has served as a consultant for Janssen, Novartis, Pfizer, Sanofi, UCB and Abbvie and has received funding for investigator-initiated studies from Novartis, Sanofi and Janssen. GS has served as a consultant for Abbvie, BMS, Eli Lilly, Gilead, GSK Novartis, Janssen and Roche and has received funding for investigator-initiated studies from BMS, Eli Lilly, GSK, Novartis and UCB. MM declares grants from Eli Lilly, Pfizer and Sanofi and personal fees from Meissa Vaccines. PI has received consulting fees from GSK. RHH has received consulting from Janssen. SA reports grant support from Johnson and Johnson. GS declares consulting fees from Abbvie.

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