antibody, 28H1, to which the PS, IRDye700DX, for tPDT is attached. Here we investigated the feasibility of using FAP-tPDT to induce cell death in murine arthritic synovium *ex vivo*.

Methods: After conjugation of the IDrY700DX to 28H1 (28H1-700DX), binding and specificity of the conjugate was determined. Subsequently, tPDT efficiency *in vitro* was established using a 3T3 fibroblast cell line stably transfected with FAP. Biodistribution using an [111 In] In-DTPA-28H1 conjugate with and without IRDye700DX was performed in healthy C57BL/6N mice as well as in C57BL/6N mice with antigen induced arthritis (AIA). Finally, the potential of FAP-tPDT to induce targeted cell death in the synovial lining was determined by treating knee joints from mice with AIA *ex vivo*.

Results: Conjugation of IRDye700DX to the antibody did not negatively influence the immunoreactive fraction or binding capacity of the conjugate (94.7% for 28H1-700DX). 28H1-700DX was able to efficiently induce FAP-specific cell death *in vitro*. At 17.6 J/cm² radiant exposure, 89.24% $\pm$ 3.67% of fibroblasts died in the group incubated with antibody compared to control incubated with buffer only ($p < 0.001$). Biodistribution of the compound with the PS showed increased accumulation in the liver compared to the antibody without PS (31.46 $\pm$ 5.49% injected dose per gram tissue (%ID/g) versus 5.32 $\pm$ 1.17 %ID/g for the antibody with or without PS, respectively ($p < 0.001$)). However, despite this increased clearance to the liver, accumulation in the inflamed joints was increased in the group injected with the antibody-PS construct (1.61 $\pm$ 0.08 %ID versus 1.13 $\pm$ 0.06 %ID for the antibody with or without PS ($p < 0.001$)). Interestingly, *ex vivo* FAP-tPDT of knee joints of arthritic mice caused significant photo-bleaching of the PS (19.69 $\pm$ 2.02% fluorescent signal remaining versus 96.00 $\pm$ 25.98% compared to the unexposed control at baseline, $p = 0.047$). Furthermore FAP-tPDT induced marked apoptosis as was indicated by an increased staining of the markers caspase-3 and yH2AX evident in the synovium of treated knee joints.

Conclusion: Here we demonstrated the feasibility of conjugating a PS to an antibody targeting FAP on activated SF without negatively impacting the binding capacity thereof. Furthermore we showed that this construct can then be used to deliver cell specific cytotoxicity through tPDT both *in vitro* and *ex vivo* in a mouse model of arthritis. This approach may have therapeutic potential in the treatment of RA.

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SAT0053 GLYCOSYLATION IN MAMMALS PROTECTS CITRULLINATED CHEMOKINES FROM PARTIAL DEGRADATION

Olexandr Korchynskyi^{1,2,3}, ¹Rzeszow University, Medical Faculty, Rzeszow, Poland; ²Institute of Cell Biology, Natl.Acad.Scie. of Ukraine, Lviv, Ukraine; ³Academic Medical Centre, University of Amsterdam, Amsterdam, Netherlands

Background: Citrullination is a posttranslational modification of specific proteins by peptidylarginine deiminase (PAD) activities. Nowadays citrullination is recognized as a hallmark of rheumatoid arthritis and other autoimmune diseases. In our recent study we have shown a presence of citrullinated chemokines epithelial-derived monocyte chemoattractant protein-1 (MCP-1/CCL2), macrophage inflammatory protein-1 α (MIP-1 α /CCL3) and neutrophil-activating peptide 78 (ENA-78/CXCL5) in biological fluids collected from patients suffering from rheumatoid arthritis (RA) versus non-autoimmune arthritis diseases and demonstrated citrullinated ENA-78/CXCL5 as an efficient macrophage chemoattractant in a contrast with non-citrullinated ENA-78/CXCL5. In our current work we found that citrullinated *in vitro* bacterially produced chemokines cannot be efficiently used as the standards in modified enzyme linked immunosorbent assay (ELISA) assays designed to detect citrullinated chemokines.

Objectives: In our work we aimed generate a procedure for preparation of stable citrullinated chemokines suitable for research applications and to validate a hypothesis that posttranslational modifications occurring in mammalian cells that can stabilize chemokines can also protect citrullinated chemokines from quick degradation.

Methods: MCP-1/CCL2 and MIP-1 α /CCL3 were cloned from total RNA isolated from synovial fibroblasts obtained from RA patient. Both bacterially-produced and mammalian cells-produced recombinant human chemokines were citrullinated by commercial rabbit PAD2. Success in

citrullination was confirmed with Western blotting and mass-spectrometry. Citrullinated chemokine concentrations were measured by modified sandwich ELISA assays.

Results: Both commercially available and self-made bacterially produced chemokines MCP-1/CCL2, MIP-1 α /CCL3 and ENA-78/CXCL5 undergo quick partial degradation upon their *in vitro* citrullination by PAD2 and cannot be detected with either Western blotting or mass-spectrometry. At the same time mammalian cells-produced properly glycosylated MCP-1/CCL2, MIP-1 α /CCL3 and ENA-78/CXCL5 can be efficiently citrullinated and successfully used as the standards in modified ELISA assays as well as in bioassays.

Conclusion: Glycosylation that is lacking in bacterially-produced proteins but occurs in mammalian cells stabilizes citrullinated chemokines thus protecting them from quickly ongoing partial degradation.

Bioethics Committee Approval: All human subject samples were collected after approval by the Institutional Review Board of the Academic Medical Center/University of Amsterdam, Amsterdam, The Netherlands (Protocol MEC 07/079 #10.17.0708) and provision of informed consent by the patients.

REFERENCE

- [1] Yoshida K., Korchynskyi O., Tak P.-P., Isozaki T., Ruth J.H., Campbell P. L., Baeten D.L., Gerlag D.M., Amin M.A., and Koch A.E. (2014) *Arthritis and Rheumatism* 66, No 10, P.2716-2727.

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SAT0054 INVESTIGATING MECHANISMS OF AUTOANTIBODY INDUCED PAIN, BONE LOSS AND ARTHRITIS DEVELOPMENT

Akila Krishnamurthy¹, Yogan Kisten¹, Katalin Sandoz², Alexandra Circumaru¹, Gustaf Wigerblad¹, Peter Damberg³, Koji Sakurabas¹, Heidi Wähämaa³, Patrik Jarvölli⁴, Juan Jimenez Jimenez Andrade⁵, Vivianne Malmström¹, Lars Klareskog¹, Camilla Svensson², Bence Réthi¹, Anca Catrina¹. ¹Karolinska Institutet, Rheumatology unit, Medicine, Stockholm, Sweden; ²Karolinska Institutet, Physiology and Pharmacology, Stockholm, Sweden; ³Karolinska Institutet, Karolinska Experimental Research and Imaging Centre (KERIC), Stockholm, Sweden; ⁴Karolinska Institutet, Stockholm, Sweden; ⁵Universidad Autónoma de Tamaulipas, Reynosa, Tamaulipas, Mexico

Background: In rheumatoid arthritis (RA), autoantibodies against citrullinated proteins (ACPAs) have been reported to be associated with bone loss, pain and tenosynovitis prior to disease onset.

Objectives: We aimed to investigate if transfer of human ACPAs into mice could reproduce these clinical observations.

Methods: Monoclonal ACPA (1325:04C03 and 1325:01B09) and control (1362:01E02) antibodies (mAbs) were generated from synovial plasma or memory B cells of RA patients. 2mg of combination of monoclonal ACPAs or control antibody were injected in BALB/c female mice (12-16 Weeks) with or without a consecutive intra-articular injection of LPS after 8 days. Pain-like behavior was monitored by measuring mechanical hypersensitivity using von Frey filaments every 3 days and estimation by up-down Dixon method. Bone mineral density was measured by micro-CT. Using specially designed mobilization casts, dedicated mouse MRI coils, and gadolinium enhanced contrast medium, the hind limbs of these mice were scanned and evaluated for any signs of soft tissue joint inflammation. Blinded to ACPA and controls, the MRI images were scored for the presence of synovial thickening, effusion and tendon inflammatory changes by 3 readers in consensus.

Results: ACPAs (1325:04C03 and 1325:01B09) induced significantly more pronounced pain-like behavior (lasting for at least 4 weeks) and reduction of the trabecular bone thickness in the hind limbs, whereas no such effect was seen with the control mAbs generated in the same way as the monoclonal ACPAs. While no macroscopic sign of joint inflammation could be detected, preliminary MRI data shows that sub-clinical joint inflammation (such as tenosynovitis) in mice injected with ACPAs but not those injected with control mAb. Intra-articular LPS injection resulted in significantly increased prolonged mechanical hypersensitivity in mice initially receiving sub-optimal doses of monoclonal ACPA as compared to those receiving control mAb. This was associated with higher levels of sub-clinical inflammation (as shown by MRI scans) in ACPA injected mice.

Conclusion: We show that ACPA induces pain-like behavior, bone loss and sub-clinical inflammation in mice, a model that mimics the pre-clinical state of ACPA positive RA.