not only professional bone-resorbing cells but also directly involved in controlling inflammatory responses. Depending on the pathophysiological context, OC can polarize the immune response towards tolerance or inflammation. Aiming at identifying key regulators of inflammatory OC functions, we have defined a miRNA-based signature, which includes miR-342-3p. MiRNAs are key regulators of gene expression that control cellular processes, including osteoclastogenesis, and few miRNAs have been described in the differentiation of myeloid precursors into mature OC.

**Objectives** To study the role of miR-342-3p in inflammatory OC.

**Methods** OC were derived from the murine monocyte RAW264.7 cells. The expression levels of miR-342-3p and OC-specific genes were monitored by qRT-PCR. RAW264.7 cells were transfected with either miR-342-3p mimics, neutralizing molecules or control miRNAs. Cell survival and proliferation were assessed at 48 hour after RANKL incubation using quantification of the caspase3-7 activity, ATP production and BrDU incorporation. Motility of OC precursors was monitored using time-laps during the course of OC differentiation. The K/BxN serum-transfer arthritis (STA) model was performed in 8 weeks old C57BL/6 males and bone marrow was flushed. Primary OC were generated from either total bone marrow or sorted CD11b + and CD11c + cell subsets of healthy and arthritic mice.

**Results** The expression of miR-342-3p was transiently up-regulated in the early phase of OC generation and was down-regulated after 24–48 hour in OC precursors. While pre-miR-342-3p promoted the maturation of RAW264.7 cells, anti-miR-342-3p inhibited all motility parameters recorded (p<0.0001, ANOVA test). Anti-miR-342-3p reduced the proliferation (p<0.01) and cell survival of OC precursors through a pro-apoptotic effect, as assessed by increased caspase3-7 activity (p<0.01). Overall, miR-342-3p neutralization in OC precursors reduced OC numbers (p<0.001) compared to the pre-miR-342-3p condition. In primary cells, miR-342-3p was up-regulated in bone marrow-derived mature OC from arthritic mice compared to healthy controls (p=0.03; STA n=5/group).

**Conclusions** Our data suggest that miR-342-3p promotes the early phase of osteoclastogenesis by enhancing the cell survival and motility of OC precursors. The up-regulation of miR-342-3p in OC isolated from arthritic mice may reflect the increased osteoclastogenic potential of inflammatory precursors in arthritis.

**Disclosure of Interest** None declared.

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**P067 TOFACITINIB IMPAIRS MONOCYTE-DERIVED DENDRITIC CELL DIFFERENTIATION IN RHEUMATOID ARTHRITIS AND PSORIATIC ARTHRITIS**

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**Career situation of first and presenting author** Post-doctoral fellow.

**Introduction** Tofacitinib is an oral Janus kinase inhibitor, recently approved for the treatment of rheumatoid arthritis (RA) and psoriatic arthritis (PsA). Although its mechanism of action has been explored in circulating cells, including neutrophils and lymphocyte, its effect on dendritic cells development and function remains still to be elucidated. Monocyte-derived dendritic cells (Mo-DC) are a subset of inflammatory DC derived from circulating monocytes with a key role in inflammation and infection.

**Objectives** The aim of this project is to evaluate the effect of Tofacitinib on inflammatory Mo-DC differentiation from RA and PsA patients, an important step in innate immunity.

**Methods** Monocytes were isolated from blood of healthy donor (HC), RA and PsA patients by magnetic separation and differentiated in the presence of GM-CSF/IL-4 cocktail for 7 days. To evaluate the effect of Tofacitinib on Mo-DC differentiation, monocyte were pre-treated with 1 μM Tofacitinib (or DMSO as control). CD209 (immature DC marker) was evaluated by flow cytometry in the CD11c + population. Non-specific macrophagocytosis (using Lucifer Yellow) and receptor-mediated endocytosis (using DQ™ Ovalbumin) were investigated by flow cytometry. The effect of Tofacitinib on NADPH oxidases (NOX) 5 and 2 expression, known players in Mo-DC differentiation, was evaluated by Western blot analysis. Finally, the frequency of CD209 + cells and their chemokine receptor expression (CXCR3/4/5 and CCR6/7) were evaluated by flow cytometry in peripheral blood (PBMC), synovial fluid (SFMC) mononuclear cells and synovial tissue cell suspensions from RA and PsA patients.

**Results** Pre-treatment of Mo-DC with Tofacitinib inhibited Mo-DC differentiation in RA and PsA patients, as evident by reduced CD209 marker expression. The decreased ability of monocytes to differentiate into DC in the presence of Tofacitinib was translated into a functional impairment of endocytic ability, in particular in PsA patients, as observed by the decreased uptake of both DQ™ Ovalbumin and Lucifer Yellow. In addition, Tofacitinib decreased NOX5 and increased NOX2 protein expression in Mo-DC of PsA and RA Mo-DC, altering the NOX2/NOX5 balance. Finally, we identified CD209 + cells in the periphery of RA and PsA patients, which were enriched in SFMC and synovial tissue cell suspension, and presented with an increased expression of CCR7 and CXCR3/5.

**Conclusions** Together, these observations suggest a novel mechanism of action of Tofacitinib in RA and PsA, by inhibiting Mo-DC development, which may alter migration of DC to the joint and subsequent activation of the immune response.

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**P068 RHEUMATOID ARTHRITIS PERIPHERAL CD14+ MONOCYTES ARE HYPER-INFLAMMATORY, HYPERGLYCOLYTIC AND RETAIN A MEMORY BIAS TOWARD M1 MACROPHAGES**

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**Career situation of first and presenting author** Post-doctoral fellow.

**Introduction** Myeloid cells with a monocyte/macrophage phenotype are present in large numbers in the rheumatoid arthritis (RA) joint, significantly contributing to disease.