EXTENDED REPORT

Single cell cloning and recombinant monoclonal antibodies generation from RA synovial B cells reveal frequent targeting of citrullinated histones of NETs

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ABSTRACT

Objectives Rheumatoid arthritis (RA) is characterised by breach of self-tolerance towards citrullinated antigens with generation of anti-citrullinated peptide/proteins antibodies (ACPA). Currently, the nature and source of citrullinated antigens driving the humoral autoimmune response within synovial ectopic lymphoid structures (ELS) is a crucial unknown aspect of RA pathogenesis. Here we characterised the autoreactive B-cell response of lesional B cells isolated from ELS+RA synovium.

Methods Single synovial tissue CD19+cells were Fluorescence Activated Cell Sorting (FACS)-sorted and Vh/Vl Ig genes cloned to generate recombinant monoclonal antibodies (rmAbs) from patients with ELS+/ACPA+RA.

Results RA-rmAbs immunoreactivity analysis provided the following key findings: (1) in a chIP-based array containing 300 autoantigens and in a ‘citrulline’ multiplex assay, a strong reactivity against citrullinated histones H2A/H2B (citH2A/citH2B) was observed in ~40% of RA-rmAbs, followed by cit-fibrinogen and cit-vimentin; (2) anti-citH2A/citH2B-reactive RA-rmAbs (but not anti-citH2A/citH2B negative) selectively recognised neutrophil extracellular traps (NETs) from peripheral blood and/or RA joint neutrophils; (3) anti-citH2A/citH2B and anti-NET immunobinding was dependent on affinity maturation and was completely abrogated following reversion of hypermutated IgVhe/vlh genes to germline sequences; (4) ELS+ (but not ELS–) RA synovial tissues engulfed into Severe Combined ImmunoDeficiency (SCID) mice released human anti-citH2A/citH2B and anti-NET antibodies in association with the intra-graft expression of CXCL13 and lymphotoxin (LT)-β, two master regulators of ELS.

Conclusion We provided novel evidence that B cells differentiated within synovial ELS in the RA joints frequent target deaminated proteins which could be generated during NETosis of RA synovial neutrophils including histones. Thus, NETs could represent a source of citrullinated antigens fuelling the ACPA autoimmune response within the RA synovium.

INTRODUCTION

Rheumatoid arthritis (RA) is characterised by breach of self-tolerance towards citrullinated proteins (anti-citrullinated peptide/proteins antibodies (ACPA)), which can occur years prior to clinical onset of RA at extra-articular sites.1–6 Several posttranslationally deaminated proteins have been indicated as a potential source of citrullinated antigens in the RA joints, but to date their cellular source and specific contribution to the lesional ACPA response is unknown.

About 40%–50% of patients with RA display synovial ectopic lymphoid structures (ELS) characterised by B-cell follicles supporting a germinal centre (GC) response.7–9 Synovial ELS are self-sustained niches whereby autoreactive B cells undergo antigen-driven selection/differentiation with local antibody diversification through Ig genes somatic hypermutation (SHM) and class switching.10 Citrullination, or arginine deimination, is catalysed by the enzyme peptidyl-arginine deiminate (PAD). In the RA synovium, monocyte–macrophages are the main source of this enzyme.11–12 As a result, citrullination of fibrinogen, vimentin and α-enseola, among others, has been observed within the RA joints and associated with circulating ACPA.13–15 Accordingly, monoclonal antibodies generated from synovial fluid B cells frequently react against citrullinated antigens.16

PAD-mediated deamination of core histones (H2A/H2B/H3/H4) has been described in neutrophils during the neutrophil extracellular traps (NETs) formation, or NETosis, a form of cell death which enhances the antimicrobial properties of activated neutrophils.17–18 Interestingly, RA synovial fluid neutrophils display an enhanced NETosis in the absence of microbial stimuli due to the RA proinflammatory milieu19 and RA sera react against citrullinated H4 from NETs.2

At present, direct evidence that synovial B cells from ELS+RA recognise citrullinated proteins and the specific contribution of different citrullinated antigens in fuelling the lesional ACPA production is missing. To this aim, we investigated the immunoreactivity of recombinant monoclonal antibodies (rmAbs) generated from single synovial B-cell clones obtained from patients with ELS+/ACPA+RA.

MATERIALS AND METHODS

A full list of methods is reported in the online supplementary methods.

Patients

Three synovial tissues from total joint replacement were obtained after informed consent
Anti-Rheumatic Drug (DMARD) therapy including methotrexate (MTX) was previously described. This board specifically approved the collection of the synovial tissue. Synovial tissue was dissected and processed as previously described.

**Synovial mononuclear cell isolation and CD19+ cell FACS sorting**

Mononuclear cells were isolated from fresh synovial tissue specimens obtained as above. Detailed method is reported in the online supplementary methods.

**Generation of recombinant monoclonal antibodies**

Single-cell real-time-PCR reactions and IgV gene amplification were performed as described in refs. 21 and 22. Briefly, cDNA from CD3-CD19+B cells was amplified using reverse primers that bind the Cμ/Cγ or Cα constant region in three independent nested-PCR. The complete sequence of primers is reported in online supplementary table S1. Aliquots of Variable Heavy (VH)/Vs/Vα, chains second PCR products were sequenced with the respective reverse primer and analysed by IgBlot. IgH complementary determining region (CDR3) amino acids and lengths were determined as described.21 The V gene somatic mutations analysis was performed using IMmunoGeneTics/Variable (IMGT/V)-QUeRy and StAndarization (QUEST) to characterise the silent versus non-silent mutation in each Framework Region (FR)/CDR region to determine the R:S ratio. The expression vector cloning strategy and the monoclonal antibody production were performed as described in ref. 21. Immunoglobulin Analysis Tool (IgAT) software was used to calculate the probability of antigen-driven selection within the Ig repertoire of the RA-rmAbs, as previously described.

**Multiplex autoantibody assay**

The multiplex autoantibody assays containing 20 citrullinated RA-associated antigens (see online supplementary table S2) was performed as previously published.5 Briefly, rmAbs were added at 10 μg/mL to custom Bio-Plex beads associated with RA putative autoantigens and incubated at room temperature (RT) for 1 h. After washing, Phycoerythrin (PE)-anti-human-IgG antibody was added to the beads and incubated at RT. The fluorescence of PE detected (Luminex200, Bio-Plex-Software V5.0, Bio-Rad) reflects the amount of antibodies that bind to the beads.

**ELISA assay for anti-citrullinated H2A and H2B**

ELISA plates were coated with citrullinated or unmodified histones H2A or H2B at 10 μg/mL in Phosphate Buffered Saline (PBS). Samples at 10 μg/mL were transferred into the ELISA plate and incubated for 2 h (SCID serum diluted 1:10). Unbound antibodies were removed by washing before incubation for 1 h with horseradish peroxidase coupled goat-anti-human-IgG. Assays were developed using TetraMethylBenzidine (TMB) Substrate Reagent Set (Becton Dickinson Optical Enzyme ImmunoAssay (BDOptEIA)). Optical densities (ODs) were measured at 450 nm.

**Stimulation of NETosis and immunofluorescence microscopy on NETs**

Neutrophils were isolated from peripheral blood (PB) of healthy donors or synovial fluid of two patients with RA using discontinuous gradient centrifugation according to ref. 24 and seeded onto cell culture cover slides at 2×10⁵ cells/well. Cells were either fixed with 4% paraformaldehyde or before fixation activated with 100 nM Phorbol Myristate Acetate (PMA) for 4 h at 37°C. NETs were stained with RA-rmAbs or Sjögren’s syndrome (SS) control rmAbs diluted in PBS for 1 h (RT). After washing with Tris Buffered Saline (TBS), Alexa488-goat-anti-human-IgG (Invitrogen, 1:200) was added for 30 min (RT). 4’, 6-Diamidino-2-Phenylindole (DAPI) (Invitrogen) was added to visualise the NETs. All sections were visualised using an Olympus BX60 microscope. All rmAbs have been tested at 10 μg/mL.

**RA synovial tissue transplantation into SCID mice**

RA SCID chimaeras were established as previously described.10 Sera and synovial grafts were harvested and analysed for autoantibodies and gene expression profiling, respectively, as previously reported.10 Detailed method is reported in the online supplementary methods.

**Statistical analysis**

Differences in quantitative variables were analysed by the Mann–Whitney U test (two groups) and by the Kruskal–Wallis with Dunn’s post-test (multiple groups). χ² test with Yates’s correction or Fisher’s exact test when required were used to evaluate associations of qualitative variables in the different groups. All the statistical analyses were performed using GraphPad-Prism5.01. A p value <0.05 was considered statistically significant. For array reactivity, the Significance Analysis of Microarrays algorithm to normalise array median fluorescence intensity values was applied.

**RESULTS**

*Ig gene analysis demonstrates intra-synovial antigen-driven B-cell affinity maturation and clonal diversification in ELS + RA synovium*

We sorted single CD19+B cells from synovial cell suspension obtained from three joint replacements of patients with ELS +/ACPA+RA (figure 1A, B). Sequence analysis of different VH/JH regions (n=139) and Vιιιι regions (Vιιιι=94;Vιιιι=81) demonstrated that the Vιιιι/Vιιιι gene repertoire of the synovial B cells was similar to PB CD5-IgM+B cells of healthy donors9 25 (figure 1E–G). We observed statistically different JH gene usage with JH2/JH3 and JH5 over-represented in the IgM and IgG synovial B cells, respectively, and JH4 less frequent in the IgG compartment (figure 1E). Generally, we observed a similar frequency in the distribution of IgM (33%), IgG (40%) and IgA (27%) isotypes across all sequences analysed (figure 1C, D). IgG and IgA synovial B-cell clones showed significantly higher number of SHM in their VH region compared with IgM, ~50% of which displayed germline sequences (see figure 2A and online supplementary figure S1A); additionally, SHM in Variable Light (Vιιιι) was higher in χ than λ chains (figure 2B). Switched B-cell clones also showed (1) high ratios of replacement (R) to silent (S) mutations in CDR1-2 compared with the FR1-3 regions (see figure 2C and online supplementary figure S1B), (2) a shorter CDR3 length compared with unswitched unmutated IgM +clones (figure 2D) and (3) a higher frequency of positively charged aa, which have been shown to be frequently used by autoreactive B cells*6 (figure 2E). We estimated that between 24% and 30% of the sequences (probability 0.05 and 0.1, respectively) from the synovial B cells display evidence of antigen selection (see online supplementary figure S2). Full antibodies characteristics are provided in online supplementary table S4. Clonal relationship analysis of the VH gene sequences of RA
synovial B cells showed evidence of intra-synovial diversification, as previously reported9 (figure 2f).

RA synovial rmAbs display frequent immunoreactivity towards citrullinated histones

To investigate the autoantigenic immunoreactivity of the synovial B-cell clones, we cloned matching VH+VL Ig genes from individual B cells into specific expression vectors and produced 66 in vitro whole rmAbs as complete IgG1 displaying identical VH+VL pairing and specificity of the parental B cells.21 26 Sufficient yield (>5 μg/mL) was obtained from 59 rmAbs (RA015/11=12; RA056/11=26; RA057/11=21), which were used for downstream analysis. First, we screened the RA-rmAbs in a synovial autoantigen microarray platform27 and in a multiplex RA-associated citrullinated antigen assay.5 Strikingly, RA-rmAbs (~40%) showed strong immunoreactivity towards citrullinated histones H2A (citH2A) and citH2B by multiplex assay (figure 3A) with reactivity to histones H2A and H2B also frequently observed in the protein array heatmap (see online supplementary figure S3). Quantitative analysis confirmed that the strongest reactivity was directed against citH2A and citH2B followed by citrullinated vimentin and fibrinogen (figure 3B). Additionally, 5 rmAbs displayed binding to different citrullinated antigens, highlighting the existence of clones with multiple citrullinated reactivity. In particular, reactivity to citrullinated vimentin was demonstrated by ELISA in three RA-rmAbs (see online supplementary figure S4).

Overall, 41% (24/59) and 34% (20/59) of the RA-rmAbs were above the threshold of citH2A and citH2B reactivity, respectively (figure 3C). Such reactivity was confirmed to be disease-specific as it was not detectable in 30 control rmAbs from circulating naïve and memory B-cells of five patients with ANA+/ENA+ SS (see online supplementary figure S5). To confirm the importance of citrullination, we tested the RA-rmAbs towards the native versus citrullinated form of H2A and H2B by ELISA. A significant increase was detected in the binding to citH2A/H2B compared with native H2A/H2B histones in a large proportion of rmAbs from ELS+ACPA+RA synovial B cells but not in either

**Figure 1** Histological characterisation of synovial ectopic lymphoid structures (ELS), single synovial CD19+ cell sorting and VH/VL Ig gene analysis. (A) Representative immunohistological characterisation of synovial tissue samples from patients with rheumatoid arthritis (RA) used in this study (RA015/11, RA056/11 and RA057/11). To assess the presence of ELS, sequential paraffin tissue sections were stained for CD20 (B cells, left-panel), CD3 (T cells, central-panel) and CD138 (plasma cells, right-panel), respectively. (B) Isolation strategy of single CD19+ RA synovial B cells is shown. Mononuclear cells were surface labelled with fluorochrome-coupled anti-CD19 and anti-CD3 antibodies; the sorting gate strategy for single CD19+CD3− B cell is shown. A total of 50 000 events is shown in the FACS plot. (C) The frequencies of μ, γ and α heavy chain among all CD19+ B cells for which VH sequences were obtained are shown. (D) Frequencies of μ, γ and α heavy chain among all CD19+ B cells for each synovial tissue are shown. (E) The VH and JH gene repertoire of single CD19+ synovial B cells for each individual chain isotype, μ (white), γ (grey) and α (black) is shown. (F) The Vκ and (G) Vλ gene repertoire are shown. The red bars indicate the VH+L gene repertoire of peripheral blood-naïve B cells. The absolute number of sequences analysed is reported over each graph. Error bars in bar graph indicate SEM for individual patients. p Values compare data from RA synovial B cells with peripheral blood-naïve B cells; *p<0.05; **p<0.001.

naïve or memory SS B cells or flu control rmAb (figure 3D, E). Interestingly, using as substrate synthetic citrullinated H2A peptides spanning the whole histone H2A length, we showed that different antibodies each recognised different citrullinated H2A epitopes (figure 3F), suggesting the occurrence of in situ ‘epitope spreading’. The immunoreactivity observed against citrullinated...
histones or multiple citrullinated antigens was not due to poly-reactivity, a phenomenon frequently observed in rmAbs generated from naïve B cells as only 1/59 clones displayed polyreactivity against multiple structurally unrelated antigens (see online supplementary figure S6).

RA synovial rmAbs with cit-histones specificity bind neutrophils NETs

We next investigated the biological significance of the observed immunoreactivity towards citrullinated histones. First, we showed that the RA SF-neutrophils spontaneously undergo NETosis, as previously described (figure 4D, top). We assessed whether synovial RA-rmAbs reactive against citrullinated histones could specifically target NETs. A large proportion of RA-rmAbs displayed strong binding to NETs generated from either PB-neutrophils of healthy donors (figure 4A) or from RA SF-neutrophils (figure 4D) with 33%, 42% and 19% of the total synovial antibody response of patients RA015/11, RA056/11 and RA057/11, directed against NETs, respectively (figure 4E). Conversely, none of the control SS-rmAbs displayed anti-NET reactivity (figure 4B–D). Of relevance, the immunoreactivity of the RA-rmAbs was restricted to neutrophils undergoing NETosis with negligible or no binding to the nucleus of resting neutrophils.

Of relevance, the reactivity towards NETs was strongly associated with the level of immunobinding to citrullinated histones in the multiplex assay (see multiplex tiles in figure 4C) and in ELISA (figure 4F).

Synovial B cells anti-NET immunoreactivity is dependent on SHM and is lost after reversion to germline

A progressive increase in the mutational load within the VH Ig genes was associated with higher reactivity to citrullinated histones in all the isotypes obtained, with the strongest difference observed in IgG-switched B-cell clones (figure 5A). Therefore, to address the importance of affinity maturation and clonal
diversification via SHM in the anti-NET reactivity, we reverted selected highly mutated RA-rmAbs with strong NETs reactivity to the corresponding \( V_H + V_L \) Ig germline sequences by overlapping PCR. The germline RA-rmAbs invariably lost their reactivity towards NETs (figure 5B). These data demonstrate that intra-synovial antigen-driven SHM is required for the immunoreactivity of RA synovial B-cell clones to NET-associated autoantigens.

ELS+RA synovia sustain anti-NET and anti-citrullinated histone antibodies in vivo in the Hu-RA/SCID chimeric mouse model

Finally, we investigated whether the anti-NET and anti-citrH2A/ H2B reactivity of the RA-rmAbs were reproduced in whole RA synovial tissue obtained from the same joints. For this purpose we used an in vivo chimeric human RA/SCID mouse transplantation model (figure 6A). As we had previously shown,\(^{10}\) RA synovial ELS were self-maintained for several weeks in the absence of recirculating immune cells (figure 6B) and released IgG-ACPA autoantibodies (measured as total anti-CCP-IgG, not shown). Mouse sera from mice transplanted with RA015/11 or RA056/11 synovial grafts contained autoreactive human anti-NET IgG (figure 6C) and/or anti-citrH2A/citrH2B histone antibodies (figure 6D). Additionally, in the synovial grafts of mice producing anti-citrullinated histones/NETs we observed significantly higher binding in anti-NET\(^+\) versus anti-NET\(^-\) clones.
A common feature of RA is the formation of discrete clusters of infiltrating lymphomononuclear cells forming lymphoid aggregates or ELS. These structures resembling secondary lymphoid organs sustain a functional ectopic-GC response and support B-cell differentiation into high affinity autoantibody-producing cells. ELS formation is dependent on the activation of the LT-β/lymphoid chemokines pathway in ectopic sites and is driven by chronic antigenic stimulation. As such, ELS (also observed in non-autoimmune conditions such as chronic infections, allograft rejections and cancer) are unique in their ability to mount disease-specific and antigen-specific immune responses. Indeed, ectopic-GCs produce antibodies against citrullinated proteins in RA, ribonucleoproteins Ro/La in SS, thyroglobulin and thyroperoxidase in Hashimoto’s thyroiditis and acetylcholine receptor in myasthenia gravis. Here, we exploited these unique features of ectopic-GC to unravel the nature and source of the citrullinated antigens driving adaptive immune responses in the RA joints generating full rmAbs from the joints of patients with ACPA+RA containing functional ectopic-GCs. As control, due to the absence of a direct comparator as ELS synovitis do not harbour B cells for cloning, we used rmAbs generated from circulating naïve and Ag-experienced memory B cells of patients with SS to exclude that any observed reactivity was due to non-specific autoimmunity, as circulating SS B cells are frequently autoreactive.

The rmAbs generated in this study have identical specificity of the parental B cells providing us the advantage of combining Ig gene repertoire and B-cell clonal analysis together with the characterisation of humoral immunoreactivity at clonal level. We adopted an unbiased approach sorting single CD19+ synovial tissue cells to have a true representation of the frequency and specificity of the autoimmune response against citrullinated antigens. Previous work generated rmAbs from RA synovial fluid B cells and showed frequent reactivity to selected citrullinated antigens. Of relevance, synovial fluid rmAbs are not clonally related; conversely, the B-cell clones that we isolated displayed highly somatically hypermutated IgVH+VL genes with evidence of intra-synovial clonal diversification compatible with their antigen-driven selection in ectopic-GC, similar to previous studies which microdissected ectopic-GC. This was particularly evident for IgG+ and IgA+ clones, while around 50% of the IgM+ B cells displayed germline sequences. As recently reported, we observed a high prevalence of IgA+ B cells in the RA joints in line with previous evidence that FDC-like synovial stromal cells directly induce IgA class-switch via a B cell Activating Factor of the TNF Family (BAFF) and APRoliation Inducing Ligand (APRIL)-dependent pathway. Importantly, we demonstrated that around 40% of RA synovial B cells from ectopic-GCs displayed reactivity against citrullinated antigens. Although, we observed frequent cross-citrullinated reactivity, using a multiplex ‘citrullinome’ assay, we showed that the strongest reactivity of ACPA-producing B cells was directed towards citrullinated histones, mainly H2A and H2B, followed by cit-vimentin and cit-fibrinogen. The data on histone reactivity were further confirmed (1) in vitro using several immunoenzymatic methods and cell-based NETs co-localisation experiments and (2) in vivo by engrafting ELS+ synovial tissues into SCID mice where we showed that the synovial production of anti-citH2A/citH2B and anti-NET autoantibodies is critically dependent on ELS formation and is sustained by high levels of CXCL13 and LTβ. Although our work clearly...
suggest that cit-histones are critically targeted by synovial-derived ACPA, further work dissecting the fine specificities of the rmAbs towards multiple citrullinated antigens is warranted, also considering that the binding of ACPA can be greatly influenced by immunodominant epitope density and citrullination protocols in different assays.

Histone citrullination is a critical step in the formation of NETs released from neutrophils in response to infectious or inflammatory stimuli to trap and kill pathogens. Approximately 70% of all NETs proteins are histones in which arginine residues are deiminated to citrulline. Thus, NETosis allows the extracellular exposure of heavily citrullinated histones and, to a much lesser degree, also of other putative RA autoantigens such as citrullinated vimentin which can become the target of the autoimmune ACPA response. Interestingly, the reactivity towards citrullinated vimentin, which was relatively low in our antibodies, was recently described in work using rmAbs derived from synovial fluid B cells, which however did not investigate the reactivity towards citrullinated histones and NETs.

The RA-rmAbs reactive towards citH2A/H2B also strongly and specifically recognised NETs in cell-based immunoassays using either RA-synovial fluid or circulating activated neutrophils as substrate. These data are highly relevant to RA pathophysiology since recent observations showed that (1) RA-synovial fluid neutrophils undergo aberrant NETosis and release citrullinated antigens and (2) patients with ACPA+RA sera recognise citrullinated histones from NETs. Importantly, the RA-rmAbs anti-NET immunoreactivity was entirely dependent on SHM in the IgVH genes since it was completely abrogated when the IgVH+VL genes were reverted into germline sequences. These data demonstrate that the anti-cit-histones/NET immunoreactivity is largely acquired within the synovial microenvironment on affinity maturation and intra-synovial diversification within ectopic-GC. Because these processes in B cells are largely dependent on cognate T-cell help, it is conceivable that Th cells with TCRs specific for processed NET peptides may be present in the RA synovium and engaged by antigen-presenting cells loading citrullinated NETs antigens. In this regard, recent evidence suggests that some NET...
proteins (eg, LL37, HMGB1) promote the activation of professional antigen-presenting cells by facilitating antigen uptake, interaction with endosomal Toll Like Receptor (TLR) and the release of type-I Interferon (IFN) and other proinflammatory cytokines as in the case of plasmacytid DCs in SLE. \(^{42, 43}\) Similarly, in autoimmune vasculitis, NETs are highly immunogenic and mediate the transfer of cytoplasmic neutrophil antigens to myeloid dendritic cells, favouring the formation of anti-neutrophil cytoplasmic antibodies. \(^{44}\)

The clearance of NETs by macrophages is an immunologically silent process. However, in an inflammatory environment, the uptake of NETs can promote proinflammatory cytokines including interleukin (IL)-1β, IL-6, and TNF-α, \(^{45}\) which are critical in RA. Their processing followed by presentation of deiminated antigens could elicit an adaptive immune response against citrullinated histones within GC in the RA joints, as also supported by long-standing evidence that synovial RA macrophages actively engulf neutrophil nuclear fragments. \(^{46}\) Although our and previous work suggests that NETosis is an important source of citrullinated proteins in the joints, several alternative/complementary mechanisms also likely contribute to the generation of citrullinated antigens and ACPR in RA both within and outside the joints. Environmental factors such as bacterial infection in periodontal disease (ie, Porphyromonas gingivalis)\(^ {47, 48}\) and smoking\(^ {49}\) can lead to the formation of citrullinated antigens at mucosal sites which, in susceptible individuals, can lead to the production of ACPR prior to the clinical onset. \(^4\) Within the RA joints, the activation of PADs by macrophages in the RA synovium can bring to the citrullination of intracellular and extracellular proteins such as vimentin and fibrin.\(^ {11-13} \) Based on our data, we propose that NETosis is an additional critical source of citrullinated antigens within the synovium, whereby normally sequestered citrullinated antigens within neutrophils, such as core histones, can be hypercitrullinated, exposed and be targeted by the immune system within the RA joints. Although a formal mechanistic demonstration is still necessary, we propose that the local release of citrullinated histones contribute to fuel the antigen-driven generation of highly mutated B-cell clones within synovial ELS resulting in the generation of high-affinity ACPR displaying anti-NET reactivity (see online supplementary figure S8).

A critical question which remains to be elucidated is whether anti-NET antibodies generated in RA ELS are pathogenic and contribute to chronic inflammation over and above ACPR generated in the peripheral compartment, as the pathogenic role of ACPR remains controversial.\(^ {50, 51} \) Additionally, it is unclear whether anti-NETs are generated in patients with early RA which display a similar prevalence of ELS+ synovitis compared with established RA.\(^ {52, 53} \) The combination of next-generation sequencing followed by single cell cloning and recombinant antibody production together with the availability of ultrasound-guided synovial biopsies could help in clarifying this aspect.\(^ {52, 53} \)

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Competing interests None declared.

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