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EXTENDED REPORT

Comparison of lipid and lipid-associated cardiovascular risk marker changes after treatment with tocilizumab or adalimumab in patients with rheumatoid arthritis

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ABSTRACT

Objective Compare changes in lipids and lipid-associated cardiovascular (CV) risk markers in patients with rheumatoid arthritis (RA) treated with tocilizumab or adalimumab.

Methods Post-hoc analysis was performed in patients with RA who received tocilizumab intravenously every 4 weeks or adalimumab subcutaneously every 2 weeks for 24 weeks in the ADACTA trial. Lipid and lipid-associated CV risk biomarkers, including high-density lipoprotein-associated serum amyloid-A (HDL-SAA), secretory phospholipase A₂ IIA (sPLA₂ IIA) and lipoprotein (a) (Lp(a)), were measured at baseline and at week 8.

Results The study included 162 patients treated with tocilizumab and 162 patients treated with adalimumab; HDL-SAA and sPLA₂ IIA were measured in a subpopulation of 87 and 97 patients, respectively. Greater increases in mean low-density lipoprotein cholesterol (LDL-C) (0.46 mmol/L (95% CI 0.30 to 0.62)), high-density lipoprotein cholesterol (HDL-C) (0.07 mmol/L (0.001 to 0.14)), total cholesterol (TC) (0.67 mmol/L (0.47 to 0.86)), triglycerides (0.24 mmol/L (0.10 to 0.38)) and TC:HDL ratio (0.27 (0.12 to 0.42)) occurred with tocilizumab from baseline to 8 weeks. HDL-SAA, sPLA₂ IIA and Lp(a) decreased more with tocilizumab than adalimumab. Median changes from baseline to week 8 were -3.2 and -1.1 mg/L (p=0.0077) for HDL-SAA and -4.1 and -1.3 ng/mL (p<0.0001) for sPLA₂ IIA; difference in adjusted means was -7.12 mg/dL (p<0.0001) for Lp(a). Similar results were observed in efficacy responders and non-responders per American College of Rheumatology and European League against Rheumatism criteria.

Conclusion LDL-C and HDL-C increased more with tocilizumab than adalimumab. HDL-SAA, sPLA₂ IIA and Lp(a) decreased more with tocilizumab. Lipid change effects of interleukin-6 and tumour necrosis factor (TNF) inhibition, manifest by their net impact on lipids and lipoproteins, are not synonymous; the clinical significance is unclear and requires further study.

Trial registration number NCT01119859; post-results

INTRODUCTION

Patients with rheumatoid arthritis (RA) are at increased risk of cardiovascular disease (CVD) compared with the general population.^{1 2} Traditional

risk factors for CVD do not appear to fully explain this increased risk,³ and additional factors, including inflammation, may contribute to CVD risk in RA.^{4 5} The impact of inflammation on lipid levels is complex and may manifest as changes in total cholesterol (TC) levels and in lipid particle-associated proteins, such as serum amyloid A (SAA) and secretory phospholipase A₂ IIA (sPLA₂ IIA); both are identified biomarkers of increased cardiovascular (CV) risk.^{6–8} Patients with severe, untreated RA may have very low lipid levels, which is paradoxical when considering their increased risk of CVD.⁹ In contrast, treatment of active disease can lead to elevated levels of TC, low-density lipoprotein cholesterol (LDL-C) and high-density lipoprotein cholesterol (HDL-C) in conjunction with reduced levels of inflammation.⁹

Lipoprotein(a) (Lp(a)) levels are increased in patients with RA.¹⁰ The association of Lp(a) with CVD in the general population has been assessed through genetic and Mendelian randomisation studies.^{11–13} These studies strongly point to Lp(a) as a causal agent in the process of atherogenesis.^{11–14}

Moderate early elevations in LDL-C, HDL-C and triglyceride levels were reported in Phase II and Phase III clinical trials of patients with RA treated with the interleukin-6 (IL-6) receptor inhibitor tocilizumab (TCZ); the TC:HDL-C ratio either decreased or remained unchanged.¹⁵ In contrast, a decline in Lp(a) with TCZ treatment and a change in HDL protein composition occurred.¹⁶ Lipid changes have also been reported in patients with RA treated with tumour necrosis factor (TNF)- α inhibitors.¹⁷ Patients with RA treated with adalimumab had increased HDL-C and apolipoprotein A1 levels, with no change in LDL-C or triglyceride levels, and improvement in atherogenic ratios.^{18 19} Data on the effect of TNF- α blockers on Lp(a) are mixed, though most did suggest a reduction.^{19–23} Described here is a post-hoc analysis of data from a clinical trial that compared IL-6 and TNF- α signalling inhibition to assess the impact of these therapeutic strategies on lipid-associated CV risk biomarkers and their relationship to treatment response. The dearth of such comparator trials despite an urgent need for better understanding of any differential effects of these agents on CV risk parameters makes this analysis important.



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PATIENTS AND METHODS

Patients

This post-hoc study included patients from the ADACTA trial (ClinicalTrials.gov number NCT01119859). ADACTA was a Phase IV study that assessed the efficacy of TCZ as monotherapy compared with adalimumab as monotherapy in adults who had RA for ≥ 6 months and who were intolerant of or not good candidates for continued use of methotrexate (MTX).²⁴ A total of 326 patients were randomly assigned 1:1 to receive either TCZ 8 mg/kg monotherapy intravenously every 4 weeks plus subcutaneous placebo every 2 weeks or adalimumab 40 mg monotherapy subcutaneously every 2 weeks plus intravenous placebo every 4 weeks for 24 weeks. Patients had to discontinue all synthetic disease-modifying antirheumatic drugs (DMARDs) within an appropriate washout period before baseline; any patient requiring treatment with a synthetic or biological DMARD was withdrawn from the study.²⁴ Analyses of core lipids and Lp(a) were performed in the ADACTA safety population, which included all patients who received at least one dose of study medication and had at least one post-dose safety assessment. Additional analyses of lipid-associated CV risk biomarkers were performed in 184 patients (97 adalimumab, 87 TCZ) who consented to donate serum bio-repository samples for further exploratory analysis and who provided both baseline and week 8 samples. Week 8 samples provide a larger sample size than do later time points, and previous studies have shown that lipid changes observed after 6 weeks remain stable with continued TCZ treatment.²⁵

Assessments

Core lipid panel (LDL-C, HDL-C, TC and triglycerides) and lipid-associated CV risk biomarker levels (Lp(a), HDL-SAA and sPLA₂ IIA) were analysed in serum samples obtained at baseline and at week 8. Week 8 was selected because of the limited availability of bio-repository samples later in the study although data for Lp(a) and core lipids were available for later time points. With the exception of LDL, which was calculated using the formula TC-HDL-C-(triglycerides/5), the core lipid panel was measured using the β -quantification method. The core lipid panel and Lp(a) was analysed by Covance Central Laboratory Services (Indianapolis, Indiana, USA). Assays for HDL-SAA and sPLA₂ IIA levels were performed at Pacific Biometrics (Seattle, Washington, USA).

sPLA₂ IIA was determined with the use of an enzyme immunoassay (EIA) kit (Cayman Chemical, Ann Arbor, Michigan, USA) by coupling a monoclonal capture antibody with detection by acetylcholinesterase/Fab conjugate and 5'-dithiobis 2-nitrobenzoic acid (DTNB). EDTA serum Lp(a) (Denka Seiken, Tokyo, Japan) was quantified using immunoturbidimetric assay kits. These assays were performed with a Roche Modular P autoanalyzer (Roche Diagnostics, Indianapolis, Indiana, USA).

For determination of HDL-SAA, serum HDL particles were isolated by polyethylene glycol 8000 (PEG-8000; Promega, Madison, Wisconsin, USA) precipitation, as described by Chiba *et al.*²⁶ Briefly, equal volumes of 13.0% PEG (P-4463; Sigma-Aldrich, St Louis, Missouri, USA) were mixed to precipitate non-HDL proteins and lipoproteins. After centrifugation for 5 min at 18 000 g, supernate SAA was determined by EIA (Abazyme; Needham, Massachusetts, USA). An anti-SAA monoclonal capture and a horseradish peroxidase-conjugated polyclonal antibody were used with 3,3',5,5'-tetramethylbenzidine (TMB) detection.

Statistical analyses

Differences between TCZ and adalimumab in change from baseline to week 8 in LDL-C, HDL-C, TC, triglycerides, TC:HDL

ratio and Lp(a) were assessed using a post-hoc analysis of covariance model adjusted for baseline laboratory parameters.

Differences in change from baseline to week 8 in HDL-SAA and sPLA₂ IIA were assessed using the Kruskal-Wallis test. Log transformation of the data was investigated because of the skewed distribution of these two parameters. The transformation did not allow the parameters to be normally distributed; therefore, the non-parametric statistical method of the Kruskal-Wallis test was used to account for the skewed distribution of the data. Change from baseline to week 8 was summarised and was also split by efficacy responders. Week 24 American College of Rheumatology (ACR) 20, ACR50 and European League against Rheumatism (EULAR) good/moderate responders and non-responders were used to analyse HDL-SAA, sPLA₂ IIA and Lp(a) levels by efficacy. Change in Lp(a) from >50 mg/dL at baseline to ≤ 50 mg/dL at week 8 was summarised based on recommendations that suggest Lp(a) levels should ideally be <50 mg/dL in patients at intermediate or high risk of CVD/coronary heart disease (CHD).²⁷ All statistical analyses were post-hoc; therefore, no adjustments were made for multiple testing, and no statistical significance can be claimed.

RESULTS

The ADACTA safety population consisted of 324 patients who were intolerant of MTX or for whom MTX was inappropriate. The bio-repository population consisted of 184 patients from the ADACTA safety population. Baseline demographics and disease characteristics were similar between the two treatment arms and between the safety and bio-repository populations (see online supplementary table S1). Core lipid parameters, including LDL-C, HDL-C, TC, triglyceride levels and TC:HDL ratio, increased from baseline to week 8 with TCZ monotherapy; numerically smaller changes were observed for these lipids with adalimumab monotherapy.

Differences in mean (95% CI) change from baseline between arms were as follows: 0.46 mmol/L (0.30 to 0.62; $p < 0.0001$) for LDL-C; 0.07 mmol/L (0.001 to 0.14; $p = 0.0453$) for HDL-C; 0.67 mmol/L (0.47 to 0.86; $p < 0.0001$) for TC; 0.24 mmol/L (0.10 to 0.38; $p = 0.0008$) for triglycerides and 0.27 (0.12 to 0.42; $p = 0.0005$) for TC:HDL ratio (table 1).

Reductions in HDL-SAA levels were observed from baseline to week 8 in patients treated with TCZ monotherapy and in those treated with adalimumab monotherapy. A greater treatment effect was observed in the TCZ group; median (IQR) change from baseline to week 8 was -3.2 mg/L (-11.1 , -1.0) and -1.1 mg/L (-7.1 , 0.6) for TCZ and adalimumab, respectively ($p = 0.0077$) (table 2 and figure 1A). Reductions in sPLA₂ IIA levels were also observed with both TCZ and adalimumab monotherapy, with greater reductions seen in the TCZ group; median (IQR) change from baseline to week 8 was -4.1 ng/mL (-7.8 , -1.1) and -1.3 ng/mL (-2.9 , 0.8) for TCZ and adalimumab, respectively ($p < 0.0001$) (table 2 and figure 1B). Greater reductions in Lp(a) were observed in patients treated with TCZ monotherapy than with adalimumab monotherapy; the difference in adjusted means (95% CI) was -7.12 mg/dL (-9.9 to -4.4) ($p < 0.0001$) (table 3 and figure 1C). Higher proportions of patients who had baseline Lp(a) levels >50 mg/dL improved (defined as achieving Lp(a) levels ≤ 50 mg/dL) by week 8 in the TCZ monotherapy group (11/21 patients (52.4%)) than in the adalimumab monotherapy group (6/24 patients (25.0%)).

Changes in HDL-SAA, sPLA₂ IIA and Lp(a) levels by efficacy response

Changes from baseline to week 8 in Lp(a), sPLA₂ IIA and HDL-SAA were also evaluated relative to efficacy response.

Table 1 Mean change from baseline to week 8* in lipid parameters (safety population)

Statistic	ADA 40 mg SC q2w, n=162	TCZ 8 mg/kg IV q4w, n=162
Total cholesterol, mmol/L		
Baseline		
n	145	150
Mean (SD)	4.94 (1.06)	5.13 (1.11)
Change from baseline to week 8		
n	129	138
Mean (SD)	0.17 (0.65)	0.79 (0.97)
Difference in adjusted means (95% CI)†		0.67 (0.47 to 0.86)
p Value‡		<0.0001
Triglycerides, mmol/L		
Baseline		
n	145	150
Mean (SD)	1.39 (0.69)	1.48 (0.97)
Change from baseline to week 8		
n	129	138
Mean (SD)	0.07 (0.47)	0.29 (0.68)
Difference in adjusted means (95% CI)†		0.24 (0.10–0.38)
p Value‡		0.0008
HDL-C, mmol/L		
Baseline		
n	145	149
Mean (SD)	1.52 (0.38)	1.56 (0.48)
Change from baseline to week 8		
n	129	137
Mean (SD)	0.07 (0.25)	0.14 (0.31)
Difference in adjusted means (95% CI)†		0.07 (0.001 to 0.14)
p Value‡		0.0453
LDL-C, mmol/L		
Baseline		
n	144	146
Mean (SD)	2.78 (0.89)	2.88 (0.87)
Change from baseline to week 8		
n	128	133
Mean (SD)	0.07 (0.53)	0.52 (0.79)
Difference in adjusted means (95% CI)†		0.46 (0.30 to 0.62)
p Value‡		<0.0001
Total cholesterol/HDL ratio		
Baseline		
n	145	149
Mean (SD)	3.40 (0.97)	3.51 (1.11)
Change from baseline to week 8		
n	129	137
Mean (SD)	-0.01(0.51)	0.24 (0.71)
Difference in adjusted means (95% CI)†		0.27 (0.12 to 0.42)
p Value‡		0.0005

*Includes only patients with both baseline and week 8 worst fasting values.

†Difference and 95% CI were based on adjusted least square means (TCZ-ADA).

‡ANCOVA model was adjusted for baseline laboratory parameter. p Values were unadjusted for multiple testing; therefore, no statistical significance can be claimed.

ADA, adalimumab; ANCOVA, analysis of covariance; HDL, high-density lipoprotein; HDL-C, high-density lipoprotein cholesterol; IV, intravenous; LDL, low-density lipoprotein; LDL-C, low-density lipoprotein cholesterol; q2w, every 2 weeks; q4w, every 4 weeks; SC, subcutaneous; TCZ, tocilizumab.

Greater reductions were seen in all three lipid-associated CV risk biomarkers at week 8 in both efficacy responders and non-responders who received TCZ monotherapy than in those who received adalimumab monotherapy. In ACR20 responders and non-responders, median IQR changes from baseline to week 8 in HDL-SAA were -3.3 mg/L (-10.0, -1.0) and -3.1 mg/L (-12.95, -1.15), respectively, with TCZ and -1.0 mg/L (-6.8, 0.55) and -1.1 mg/L (-8.7, 1.2), respectively, with adalimumab. Similar trends were observed for ACR50 and EULAR good/

moderate responders and non-responders for HDL-SAA and for sPLA₂ IIA (table 4) and Lp(a) (table 5).

DISCUSSION

It was previously demonstrated that though LDL-C levels increase in patients treated with TCZ, Lp(a) levels may decline, and favourable changes in HDL-C level and HDL composition may occur.¹⁶ Published data regarding the effect of TNF- α blockers on Lp(a) are conflicting.^{19–22} Data collected in the

Table 2 Change from baseline to week 8* in HDL-SAA and sPLA₂ IIA (bio-repository population)

	ADA 40 mg SC q2w, n=97	TCZ 8 mg/kg IV q4w, n=87	p Value†
HDL-SAA (mg/L)			
Baseline			
Median (IQR)	7.8 (4.0, 16.8)	6.9 (3.0, 13.9)	
Mean (SD)	16.8 (23.6)	13.0 (19.2)	
	n=64	n=58	
Change from baseline to week 8			
Median (IQR)	-1.1 (-7.1, 0.6)	-3.2 (-11.1, -1.0)	0.0077
Mean (SD)	-4.4 (12.3)	-9.3 (18.3)	
	n=62	n=55	
sPLA₂ IIA (ng/mL)			
Baseline			
Median (IQR)	6.8 (5.1, 10.2)	7.2 (4.7, 12.2)	
Mean (SD)	10.7 (12.4)	2.1 (14.8)	
	n=90	n=77	
Change from baseline to week 8			
Median (IQR)	-1.3 (-2.9, 0.8)	-4.1 (-7.8, -1.1)	<0.0001
Mean (SD)	-1.8 (9.15)	-7.8 (12.8)	
	n=86	n=73	

n represents number of patients contributing to summary statistics.
 *Includes only patients with both baseline and week 8 values.
 †Comparison between medians (ADA vs TCZ) using the Kruskal–Wallis test.
 ADA, adalimumab; HDL-SAA, high-density lipoprotein-associated serum amyloid A; IV, intravenous; q2w, every 2 weeks; q4w, every 4 weeks; SC, subcutaneous; sPLA₂ IIA, secretory phospholipase A₂ II; TCZ, tocilizumab.

ADACTA study allowed, for the first time, a direct comparison of the effect of anti-TNF (aTNF) and anti-IL-6 therapies on lipids and lipid-associated CV risk biomarkers in the context of a single study in which other trial parameters were balanced between groups by strict randomisation and blinding. Post-hoc analyses showed a greater increase in LDL-C and greater reductions in Lp(a), HDL-SAA and sPLA₂ IIA levels over the first 8 weeks of treatment in patients with RA who received TCZ monotherapy compared with those who received adalimumab monotherapy. Reductions in HDL-SAA, sPLA₂ IIA and Lp(a) were observed in efficacy responders and non-responders, suggesting that the cytokines themselves—as opposed to, or at least in addition to, other disease activity-related pathways—mediate some of the effects observed. Although reductions for both responders and non-responders treated with TCZ monotherapy appear to be larger than for those treated with adalimumab monotherapy, the small sample size for each group precludes statistical comparison.

Increased inflammation in patients with autoimmune disorders such as RA is associated with quantitative and qualitative modifications of circulating lipids. Recent data suggest that decreased LDL-C and HDL-C levels in patients with active RA are associated with increased cholesterol catabolism rather than decreased synthesis, with the reverse true following treatment of inflammation.²⁸ These data are somewhat reassuring. Nevertheless, it should be noted that parallel lipid kinetic data do not exist for TCZ or for aTNF therapies. In addition to

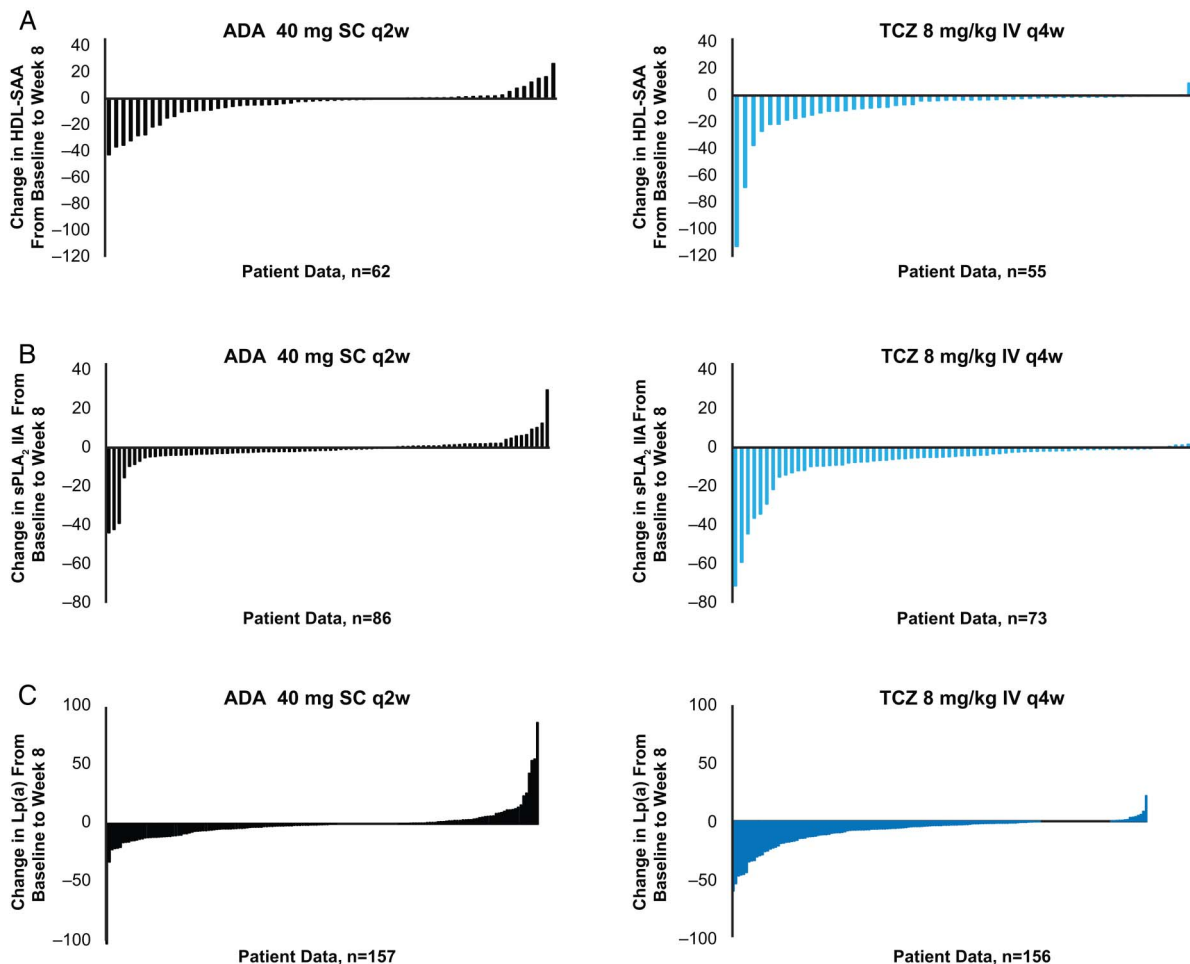


Figure 1 Change from baseline to week 8 in HDL-SAA (A, bio-repository population), sPLA₂ IIA (B, bio-repository population) and Lp(a) (C, safety population)—patient-level data. ADA, adalimumab; HDL-SAA, high-density lipoprotein-associated serum amyloid A; IV, intravenous; Lp(a), lipoprotein (a); sPLA₂ IIA, secretory phospholipase A₂ IIA; q2w, every 2 weeks; q4w, every 4 weeks; SC, subcutaneous; TCZ, tocilizumab.

Table 3 Change from baseline to week 8* in Lp(a) for all patients (safety population)

Subgroup	Statistic	ADA 40 mg SC q2w, n=162	TCZ 8 mg/kg IV q4w, n=162
Lp(a) (mg/dL)			
Baseline			
n		162	162
Mean (SD)		25.5 (30.2)	22.4 (25.5)
min, max		4.0, 194.0	4.0, 131.0
Change from baseline to week 8			
n		157	156
Mean (SD)		-1.1 (15.1)	-7.6 (12.0)
min, max		-102.5, 86.4	-60, 21.9
Difference in adjusted means (95% CI)†		-7.12 (-9.9 to -4.4)	
p Value‡		<0.0001	

*Includes only patients with both baseline and week 8 values.

†Difference and 95% CI based on adjusted least square means (TCZ-ADA).

‡ANCOVA model was adjusted for baseline Lp(a). p Values were unadjusted for multiple testing; therefore, no statistical significance can be claimed.

ADA, adalimumab; ANCOVA, analysis of covariance; IV, intravenous; Lp(a), lipoprotein (a); q2w, every 2 weeks; q4w, every 4 weeks; SC, subcutaneous; TCZ, tocilizumab.

quantitative changes, inflammation may be accompanied by changes in lipid particle size and protein composition that may result in a potentially pro-atherogenic lipid profile. In particular, SAA, sPLA₂ IIA and cholesteryl ester transfer protein can remodel the lipid and protein composition of HDL particles.²⁹ Association of HDL with SAA and sPLA₂ IIA impairs antioxidative and atheroprotective capacity of HDL.³⁰ Displacement of HDL cargo proteins that have antioxidant activity, such as apoA1 and paraoxonase 1, with these acute-phase reactants results in HDL particles with pro-oxidant capacity.³¹ SAA-containing HDL may be retained in the vascular matrix by vascular proteoglycans and may therefore not be available for the reverse cholesterol transport.³²

Elevated Lp(a) levels can contribute to atherogenesis by depositing LDL-C in the intima of the vascular wall by the recruitment of inflammatory cells or the binding of proinflammatory oxidised phospholipids. Lp(a) shares structural homology with plasminogen and plasmin and has potentially prothrombotic and anti-fibrinolytic properties, which could promote clot stabilisation and thrombosis.²⁷ Recent data suggest that Lp(a) is an independent genetic risk marker of CVD and is potentially causal.¹³ It has been recommended that desirable levels of Lp(a) for patients at intermediate or high risk of CVD/CHD are <50 mg/dL.²⁷

The observation that reductions in HDL-SAA, sPLA₂ IIA and Lp(a) occurred in non-responders suggests that the reduction in these potentially pro-atherogenic proteins with TCZ may be, at least partially, independent of the RA treatment response, reflecting the effect of IL-6 blockade outside the joint. IL-6 is known to have an effect on the release of acute-phase proteins from hepatocytes; SAA is increased and apoA1 is decreased.^{31 33} Synthesis of Lp(a) is increased in response to IL-6 in cultured human hepatocytes through direct regulation of a response element in the Lp(a) promoter, an effect that was inhibited by TCZ.³⁴ Therefore, the results of the current analysis may be explained by an effect of IL-6 inhibition that is independent of its anti-arthritis effect, similar to its effect on C reactive protein production.³⁵ Decreases in HDL-SAA, sPLA₂ IIA and Lp(a) levels also occurred in adalimumab non-responders; although they appeared to have occurred to a lesser extent than in TCZ

Table 4 Change from baseline to week 8* in HDL-SAA and sPLA₂ IIA for week 24 ACR20, ACR50 and EULAR good/moderate responders and non-responders (bio-repository population)

Subgroup	Statistic	ADA 40 mg SC q2w, n=97	TCZ 8 mg/kg IV q4w, n=87
HDL-SAA change from baseline to week 8 (mg/L)			
ACR20 responders			
n		32	39
Median (IQR)		-1.0 (-6.8, 0.55)	-3.3 (10.0, -1.0)
Mean (SD)		-5.8 (12.5)	-10.3 (21.1)
ACR20 non-responders			
n		30	16
Median (IQR)		-1.1 (-8.7, 1.2)	-3.1 (-12.95, -1.15)
Mean (SD)		-2.9 (12.1)	-7.1 (7.8)
ACR50 responders			
n		20	26
Median (IQR)		-1.6 (-6.6, 0.25)	-3.3 (-8.6, -1.0)
Mean (SD)		-5.1 (11.2)	-11.8 (25.1)
ACR50 non-responders			
n		42	29
Median (IQR)		-0.9 (-7.10, 0.7)	-3.2 (-12.8, -1.0)
Mean (SD)		-4.0 (12.9)	-7.2 (8.4)
EULAR good/moderate responders			
n		36	45
Median (IQR)		-2.8 (-9.05, 0.5)	-3.2 (-10.0, -1.0)
Mean (SD)		-6.2 (12.3)	-9.6 (19.8)
EULAR non-responders			
n		26	10
Median (IQR)		-0.5 (-5.7, 1.2)	-2.9 (-18.2, -1.3)
Mean (SD)		-1.9 (12.1)	-8.0 (8.9)
sPLA ₂ IIA change from baseline to week 8 (ng/mL)			
ACR20 responders			
n		47	52
Median (IQR)		-1.3 (-3.2, 0.7)	-4.8 (-9.5, -1.3)
Mean (SD)		-1.6 (8.3)	-9.5 (14.8)
ACR20 non-responders			
n		39	21
Median (IQR)		-0.6 (-2.8, 1.9)	-3.1 (-4.9, -1.0)
Mean (SD)		-2.0 (10.2)	-3.7 (2.7)
ACR50 responders			
n		24	36
Median (IQR)		-1.8 (-2.6, 0.2)	-5.0 (-9.5, -0.95)
Mean (SD)		-1.6 (2.5)	-9.7 (13.9)
ACR50 non-responders			
n		62	37
Median (IQR)		-1.0 (-3.2, 1.4)	-4.0 (-6.7, -1.5)
Mean (SD)		-1.9 (10.7)	-6.1 (11.5)
EULAR good/moderate responders			
n		53	62
Median (IQR)		-1.6 (-3.2, 0.7)	-4.7 (-9.1, -1.1)
Mean (SD)		-2.2 (9.8)	-8.7 (13.7)
EULAR non-responders			
n		33	11
Median (IQR)		-0.2 (-2.1, 1.9)	-2.7 (-6.0, -0.8)
Mean (SD)		-1.2 (8.1)	-3.2 (2.4)

n represents number of patients contributing to summary statistics.

The range of data for each parameter is depicted in figure 1.

*Includes only patients with both baseline and week 8 values.

ACR, American College of Rheumatology; ADA, adalimumab; EULAR, European League against Rheumatism; HDL-SAA, high-density lipoprotein-associated serum amyloid-A; IV, intravenous; q2w, every 2 weeks; q4w, every 4 weeks; SC, subcutaneous; sPLA₂ IIA, secretory phospholipase A₂ IIA; TCZ, tocilizumab.

Table 5 Change from baseline to week 8* in Lp(a) for week 24 ACR20, ACR50 and EULAR good/moderate responders and non-responders (safety population)

Subgroup	Statistic	ADA 40 mg SC q2w, n=162	TCZ 8 mg/kg IV q4w, n=162
Lp(a) (mg/dL) change from baseline to week 8			
ACR20 responders			
n		79	104
Mean (SD)		-2.5 (18.0)	-8.1 (11.9)
min, max		-102.5, 86.4	-60.0, 4.8
ACR20 non-responders			
n		78	52
Mean (SD)		0.3 (11.4)	-6.5 (12.1)
min, max		-22.0, 55.2	-53.6, 21.9
ACR50 responders			
n		45	76
Mean (SD)		-3.6 (7.2)	-9.0 (13.0)
min, max		-32.6, 6.3	-60.0, 4.8
ACR50 non-responders			
n		112	80
Mean (SD)		-0.1 (17.2)	-6.2 (10.8)
min, max		-102.5, 86.4	-53.6, 21.9
EULAR good/moderate responders			
n		88	125
Mean (SD)		-2.0 (17.8)	-8.6 (12.6)
min, max		-102.5, 86.4	-60.0, 4.8
EULAR non-responders			
n		69	31
Mean (SD)		-0.0 (10.8)	-3.5 (7.8)
min, max		-22.0, 55.2	-17.9, 21.9

*Includes only patients with both baseline and week 8 values. ACR, American College of Rheumatology; ADA, adalimumab; EULAR, European League against Rheumatism; IV, intravenous; Lp(a), lipoprotein (a); q2w, every 2 weeks; q4w, every 4 weeks; SC, subcutaneous; TCZ, tocilizumab.

non-responders, the numbers were too small to make a statistical comparison. In contrast, results from a recent cohort study of patients with RA treated with TNF- α inhibitors, including adalimumab, showed that HDL-SAA was significantly decreased and that ApoAII was increased in EULAR good responders but not in non-responders, suggesting that the changes were due to the anti-inflammatory effect of TNF- α inhibition.³⁶ Furthermore, no change in Lp(a) levels was observed in patients with RA treated with adalimumab for 3 months, and adalimumab did not have a direct effect on Lp(a) promoter activity in human hepatocytes, as was observed with TCZ.³⁴ The small number of patients in the current analysis preclude statistical comparison between responders and non-responders in each treatment group, and larger studies would be required to investigate mechanisms of IL-6 and TNF- α effects on HDL-SAA, sPLA₂ IIA and Lp(a).

Limitations of this data analysis from the ADOPTA study are that it was conducted post-hoc and that the treatment duration analysed was relatively short. Longer-term effects on lipid changes were not investigated, though these same effects seen at week 8 have been seen previously with TCZ treatment out to 24 weeks;¹⁶ nevertheless, the effect on CV risk cannot be determined. It is unknown whether the changes in lipid profiles and the reductions in Lp(a), HDL-SAA and sPLA₂ IIA levels observed with TCZ treatment and the differences observed in

treatment with adalimumab will have any implications on CV risk in patients with RA. Results from meta-analyses of mostly observational studies suggest that aTNF therapy for RA is likely associated with a decreased CV risk despite increases in lipid levels.^{37, 38} Although the risk of CV events with TCZ has not been determined, the number of CV events in the placebo-controlled periods of the core TCZ Phase III trials was low. In addition, long-term data available to date have shown a stable rate of CV events over time with TCZ exposure.²⁵ Furthermore, risk of on-treatment major adverse CV events was found to be associated with control of disease activity, but not lipid changes, in a retrospective post-hoc analysis of pooled data from trials, though admittedly the number of adjudicated events was modest.³⁹ Clearly, studies designed specifically to compare the effects of biologics on lipid changes and CV outcomes are still required.

In conclusion, despite the overall increased LDL-C levels observed with TCZ compared with adalimumab, TCZ may potentially exert beneficial actions on HDL-SAA, sPLA₂ IIA and Lp(a), suggesting that IL-6 blockade may work in different ways to influence CVD risk.

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REFERENCES

- Solomon DH, Goodson NJ, Katz JN, *et al.* Patterns of cardiovascular risk in rheumatoid arthritis. *Ann Rheum Dis* 2006;65:1608–12.
- Aviña-Zubieta JA, Choi HK, Sadatsafavi M, *et al.* Risk of cardiovascular mortality in patients with rheumatoid arthritis: a meta-analysis of observational studies. *Arthritis Rheum* 2008;59:1690–7.
- del Rincón I, Williams K, Stern MP, *et al.* High incidence of cardiovascular events in a rheumatoid arthritis cohort not explained by traditional cardiac risk factors. *Arthritis Rheum* 2001;44:2737–45.
- Sattar N, McCarey DW, Capell H, *et al.* Explaining how "high-grade" systemic inflammation accelerates vascular risk in rheumatoid arthritis. *Circulation* 2003;108:2957–63.
- Myasoedova E, Crowson CS, Kremers HM, *et al.* Lipid paradox in rheumatoid arthritis: the impact of serum lipid measures and systemic inflammation on the risk of cardiovascular disease. *Ann Rheum Dis* 2011;70:482–7.
- Kugiyama K, Ota Y, Takazoe K, *et al.* Circulating levels of secretory type II phospholipase A(2) predict coronary events in patients with coronary artery disease. *Circulation* 1999;100:1280–4.

- 7 Ridker PM, Hennekens CH, Buring JE, *et al.* C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women. *N Engl J Med* 2000;342:836–43.
- 8 Mallat Z, Benessiano J, Simon T, *et al.* Circulating secretory phospholipase A2 activity and risk of incident coronary events in healthy men and women: the EPIC-Norfolk study. *Arterioscler Thromb Vasc Biol* 2007;27:1177–83.
- 9 Robertson J, Peters MJ, McInnes IB, *et al.* Changes in lipid levels with inflammation and therapy in RA: a maturing paradigm. *Nat Rev Rheumatol* 2013;9:513–23.
- 10 Lee YH, Choi SJ, Ji JD, *et al.* Lipoprotein(a) and lipids in relation to inflammation in rheumatoid arthritis. *Clin Rheumatol* 2000;19:324–5.
- 11 Clarke R, Peden JF, Hopewell JC, *et al.* Genetic variants associated with Lp(a) lipoprotein level and coronary disease. *N Engl J Med* 2009;361:2518–28.
- 12 Kamstrup PR, Tybjaerg-Hansen A, Nordestgaard BG. Genetic evidence that lipoprotein(a) associates with atherosclerotic stenosis rather than venous thrombosis. *Arterioscler Thromb Vasc Biol* 2012;32:1732–41.
- 13 Kamstrup PR, Tybjaerg-Hansen A, Steffensen R, *et al.* Genetically elevated lipoprotein(a) and increased risk of myocardial infarction. *JAMA* 2009;301:2331–9.
- 14 Boffa MB, Koschinsky ML. Update on lipoprotein(a) as a cardiovascular risk factor and mediator. *Curr Atheroscler Rep* 2013;15:360.
- 15 Choy E, Sattar N. Interpreting lipid levels in the context of high-grade inflammatory states with a focus on rheumatoid arthritis: a challenge to conventional cardiovascular risk actions. *Ann Rheum Dis* 2009;68:460–9.
- 16 McInnes IB, Thompson L, Giles JT, *et al.* Effect of interleukin-6 receptor blockade on surrogates of vascular risk in rheumatoid arthritis: MEASURE, a randomised, placebo-controlled study. *Ann Rheum Dis* 2015;74:694–702.
- 17 Daien CI, Dony Y, Barnette T, *et al.* Effect of TNF inhibitors on lipid profile in rheumatoid arthritis: a systematic review with meta-analysis. *Ann Rheum Dis* 2012;71:862–8.
- 18 Popa C, Netea MG, Radstake T, *et al.* Influence of anti-tumour necrosis factor therapy on cardiovascular risk factors in patients with active rheumatoid arthritis. *Ann Rheum Dis* 2005;64:303–5.
- 19 Wijbrandts CA, van Leuven SI, Boom HD, *et al.* Sustained changes in lipid profile and macrophage migration inhibitory factor levels after anti-tumour necrosis factor therapy in rheumatoid arthritis. *Ann Rheum Dis* 2009;68:1316–21.
- 20 Cauza E, Cauza K, Hanusch-Enserer U, *et al.* Intravenous anti TNF-alpha antibody therapy leads to elevated triglyceride and reduced HDL-cholesterol levels in patients with rheumatoid and psoriatic arthritis. *Wien Klin Wochenschr* 2002;114:1004–7.
- 21 Seriollo B, Paolino S, Sulli A, *et al.* Effects of anti-TNF-alpha treatment on lipid profile in patients with active rheumatoid arthritis. *Ann N Y Acad Sci* 2006;1069:414–19.
- 22 Hjeltnes G, Hollan I, Førre O, *et al.* Serum levels of lipoprotein(a) and E-selectin are reduced in rheumatoid arthritis patients treated with methotrexate or methotrexate in combination with TNF-alpha-inhibitor. *Clin Exp Rheumatol* 2013;31:415–21.
- 23 Sattar N, Crompton P, Cherry L, *et al.* Effects of tumor necrosis factor blockade on cardiovascular risk factors in psoriatic arthritis: a double-blind, placebo-controlled study. *Arthritis Rheum* 2007;56:831–9.
- 24 Gabay C, Emery P, van Vollenhoven R, *et al.* Tocilizumab monotherapy versus adalimumab monotherapy for treatment of rheumatoid arthritis (ADACTA): a randomised, double-blind, controlled phase 4 trial. *Lancet* 2013;381:1541–50.
- 25 Schiff MH, Kremer JM, Jahreis A, *et al.* Integrated safety in tocilizumab clinical trials. *Arthritis Res Ther* 2011;13:R141.
- 26 Chiba H, Akizawa K, Fujisawa S, *et al.* A rapid and simple quantification of human apolipoprotein E-rich high-density lipoproteins in serum. *Biochem Med Metabolic Biol* 1992;47:31–7.
- 27 Nordestgaard BG, Chapman MJ, Ray K, *et al.* Lipoprotein(a) as a cardiovascular risk factor: current status. *Eur Heart J* 2010;31:2844–53.
- 28 Charles-Schoeman C, Fleischmann R, Davignon J, *et al.* Potential mechanisms leading to the abnormal lipid profile in patients with rheumatoid arthritis versus healthy volunteers and reversal by tofacitinib. *Arthritis Rheumatol* 2015;67:616–25.
- 29 Jahangiri A, de Beer MC, Noffsinger V, *et al.* HDL remodeling during the acute phase response. *Arterioscler Thromb Vasc Biol* 2009;29:261–7.
- 30 Rohrer L, Hersberger M, von Eckardstein A. High density lipoproteins in the intersection of diabetes mellitus, inflammation and cardiovascular disease. *Curr Opin Lipidol* 2004;15:269–78.
- 31 Khovidhunkit W, Kim MS, Memon RA, *et al.* Effects of infection and inflammation on lipid and lipoprotein metabolism: mechanisms and consequences to the host. *J Lipid Res* 2004;45:1169–96.
- 32 Chait A, Han CY, Oram JF, *et al.* Thematic review series: the immune system and atherogenesis. Lipoprotein-associated inflammatory proteins: markers or mediators of cardiovascular disease? *J Lipid Res* 2005;46:389–403.
- 33 Gabay C, Kushner I. Acute-phase proteins and other systemic responses to inflammation. *N Engl J Med* 1999;340:448–54.
- 34 Müller N, Schulte DM, Türk K, *et al.* IL-6 blockade by monoclonal antibodies inhibits apolipoprotein (a) expression and lipoprotein (a) synthesis in humans. *J Lipid Res* 2015;56:1034–42.
- 35 Smolen JS, Avila JC, Aletaha D. Tocilizumab inhibits progression of joint damage in rheumatoid arthritis irrespective of its anti-inflammatory effects: disassociation of the link between inflammation and destruction. *Ann Rheum Dis* 2012;71:687–93.
- 36 Jamnitski A, Levels JH, van den Oever IA, *et al.* High-density lipoprotein profiling changes in patients with rheumatoid arthritis treated with tumor necrosis factor inhibitors: a cohort study. *J Rheumatol* 2013;40:825–30.
- 37 Westlake SL, Colebatch AN, Baird J, *et al.* Tumor necrosis factor antagonists and the risk of cardiovascular disease in patients with rheumatoid arthritis: a systematic literature review. *Rheumatology (Oxford)* 2011;50:518–31.
- 38 Barnabe C, Martin BJ, Ghali WA. Systematic review and meta-analysis: anti-tumor necrosis factor alpha therapy and cardiovascular events in rheumatoid arthritis. *Arthritis Care Res* 2011;63:522–9.
- 39 Rao VU, Pavlov A, Klearman M, *et al.* An evaluation of risk factors for major adverse cardiovascular events during tocilizumab therapy. *Arthritis Rheumatol* 2015;67:372–80.