Metabolomics for rheumatic diseases: has the time come?

Luca Semerano,1,2,3 Paul-Henri Roméo,4 Marie-Christophe Boissier1,2,3

The last decade of biomedical research has been impacted by the appearance of systems biology, an analytical approach to complex biological systems that makes use of the ‘-omics’, that is, high-throughput technologies that allow the comprehensive characterisation of genes (genomics), RNA transcripts (transcriptomics) or proteins (proteomics). Since their appearance, the ‘omics’ held promise of a radical change in the research paradigms. The classical research strategy adopts a ‘bottom-up’ approach, that is, a scientific hypothesis is tested in an isolated system (a cell type, a tissue) or in an animal model, and then extrapolated to the systemic level. Conversely, systems biology uses a ‘top-down’ approach in which data are gathered at systemic level and scientific hypotheses are subsequently derived and tested in a controlled, limited setting.1 The latter approach has been defined as data-driven or hypothesis-forming. Although genomics, transcriptomics and proteomics have generated a large amount of data, translation of these data into useful information, and in turn into beneficial outcomes for the patient, is only starting.2

The newcomer in the ‘-omics’ family is metabolomics. The metabolome is defined as the complete set of metabolites present in a given biological system. Complementary to transcriptomics or proteomics, the metabolome analysis describes both qualitatively and quantitatively the final products of cellular regulatory pathways. As such, the metabolome reflects the ultimate response of a biological system to genetic and/or environmental changes.

Metabolites that are characterised in metabolomics studies are small molecules with size ranging from 100 to 1000 Da.3 Although the concept that metabolites in biological fluids may reflect health status dates back to the 1940s, the term ‘metabolic profile’ was only introduced in the early 1970s when metabolite characterisation entered into the era of gas chromatography coupled to mass spectrometry technology.4 Extensive metabolomic analyses have become possible only recently thanks to the use of mass spectrometry or nuclear MR coupled to multivariate analyses of the generated data. The actual version of the human metabolome database 3.6 refers to 41 808 metabolic entries,5 and ongoing developments in microfluidic and interfaced integrations will boost the human metabolome project in the next few years.6

Metabolomics might be well suited to study rheumatic conditions for several reasons. First, metabolites can be considered as ‘final’ responders to environmental cues and thus metabolomics will indicate biological responses of patients suffering from rheumatic conditions to environment, including nutrition, disease states, infection, exposition to xenobiotics, pharmacological treatments and others.8 Second, pathways that regulate metabolic homeostasis and immune response display an interplay that is evolutionary conserved.9 Metabolic diseases like obesity and type II diabetes are associated with chronic inflammation, and the high burden of cardiovascular disease in rheumatic conditions shows that immunoinflammatory diseases are associated with altered metabolic homeostasis. Indeed, proliferating cells involved in inflammation and immune response act in an environment of low-oxygen tension and rely on anaerobic glucose metabolism for their nutritional supply. Moreover, their proliferative and effector activity affects the metabolism of the involved tissues. This can reflect into modifications of the metabolic profile of the patient that will be depicted by metabolomics.10

But, rather than depicting metabolic repercussion of inflammation, the real challenge of metabolomics in rheumatology will be to identify metabolite profiles that are specific to a given rheumatic condition. For example, in a Swedish study, plasma metabolomics discriminated patients with rheumatoid arthritis (RA) or psoriatic arthritis (PsA) from controls with 90% sensitivity and 94% specificity.11

In another work, the urinary metabolic footprint of patients with RA or PsA predicted clinical response to anti-tumour necrosis factor agents with 88.9% sensitivity and 85.7% specificity. Moreover, response to treatment with either infliximab or etanercept reflected into a different metabolite profile.12

These results indicate that metabolomics may provide relevant biomarkers to improve diagnostic accuracy, define prognosis and predict and monitor treatment efficacy.13 Moreover, disease-specific metabolic pathways could indicate new potential therapeutic targets as acting on these pathways may allow to modulate the disease process by directly altering the metabolism of the involved cellular actors. Additionally, some metabolites may be endowed themselves with a pathological action, as is the case of arachidonic acid metabolites that are produced in response to proinflammatory cues and that further potentiate and amplify the inflammatory process. For example, synovial cells from patients with RA have been reported to preferentially convert oestrogens to 16OH α-metabolites, which may facilitate synovial inflammation,14 while the cholesterol metabolite 27-hydroxysterol promotes vessel wall inflammation and atherosclerosis.15

Finally, metabolomics comes at a time when the other omics techniques are well developed and advanced, which would allow to implement and integrate the data obtained from metabolic fingerprinting with those coming from the other three omics, forming in the ‘-omics’ world the group of the ‘big four’.2 This has been claimed as the inevitable way to validate metabolomics results, together with the replication of the results in independent cohorts.

Nevertheless, the systems biology approach does not explain the reason for the abundance of a given metabolite in a biological system, nor establish whether the metabolite level is associated with the cause or the consequence of the disease.16

For example, in a model of experimental colitis, serum glutamine prevalence over other amino acids correlated with the degree of colonic inflammation. Of note, glutamine supplementation ameliorated colonic lesions.17 This demonstrated that glutamine increased as a result of a compensatory and not of a pathogenic mechanism, and that, for therapeutic purposes, this increase needs to be enhanced and not antagonised.

Hence, we strongly support the idea that classical ‘bottom-up’ research can be usefully integrated with systems biology to dissect a metabolic pathway responsible
for the increased or reduced presence of a given metabolite in a sample. Guma et al.\(^{18}\) used a similar approach. To study the therapeutic potential of choline kinase-α (ChoKα) pathways inhibition in RA, the authors performed targeted \(^1\)H magnetic resonance spectroscopy (HMRS) on both synovium samples and cultured fibroblast-like synoviocytes (FLS) from patients with RA and showed the presence of high levels of choline-containing metabolites. Moreover, the ratio of glycerophosphocholine to phosphocholine in the samples suggested that these metabolites resulted from ChoKα activity. Then, they confirmed the hypothesis of ChoKα activation by repeating HMRS profiling of FLS in the presence or absence of ChoKα inhibitors. Finally, the authors confirmed the therapeutic relevance of ChoKα pathways by showing that the ChoKα inhibitor MN58b ameliorated arthritis and resulted in reduced synovial transcription of interleukin (IL)-1, IL-6 and matrix metalloproteinases in the K/BxN mouse model. In this work, the metabolites of interest had been chosen previously, hence a proper analysis of the study was not performed.\(^{19}\)

Contributors LS and M-CB, drafted the article. LS, P-HR and M-CB revised the article for important intellectual content. LS, P-HR and M-CB approved the final version.

Competing interests None.

Provenance and peer review Commissioned; externally peer reviewed.

REFERENCES


CrossMark

Received 5 November 2014
Revised 22 December 2014
Accepted 6 January 2015
Published Online First 21 January 2015

http://dx.doi.org/10.1136/annrheumdis-2014-205696

doi:10.1136/annrheumdis-2014-206618

REFERENCES