EXTENDED REPORT

Preclinical characterisation of the GM-CSF receptor as a therapeutic target in rheumatoid arthritis


ABSTRACT

Objective Previous work has suggested that the granulocyte macrophage colony stimulating factor (GM-CSF)–GM-CSF receptor α axis (GM-CSFRα) may provide a new therapeutic target for the treatment of rheumatoid arthritis (RA). Therefore, we investigated the cellular expression of GM-CSFRα in RA synovial tissue and investigated the effects of anti-GM-CSFRα antibody treatment in vitro and in vivo in a preclinical model of RA.

Methods We compared GM-CSFRα expression on macrophages positive for CD68 or CD163 on synovial biopsy samples from patients with RA or psoriatic arthritis (PsA) to disease controls. In addition, we studied the effects of CAM-3003, an anti-GM-CSFR antibody in a collagen induced arthritis model of RA in DBA/1 mice. The pharmacokinetic profile of CAM-3003 was studied in naïve CD11c(+)CR mice (see online supplement) and used to interpret the results of the pharmacodynamic studies in BALB/c mice.

Results GM-CSFRα was expressed by CD68 positive and CD163 positive macrophages in the synovium, and there was a significant increase in GM-CSFRα positive cells in patients with RA as well as patients with PsA compared with patients with osteoarthritis and healthy controls. In the collagen induced arthritis model there was a dose dependent reduction of clinical arthritis scores and the number of F4/80 positive macrophages in the inflamed synovium after CAM-3003 treatment. In BALB/c mice CAM-3003 inhibited recombinant GM-CSF mediated margination of peripheral blood monocytes and neutrophils.

Conclusions The findings support the ongoing development of therapies aimed at interfering with GM-CSF or its receptor in various forms of arthritis, such as RA and PsA.

INTRODUCTION

Rheumatoid arthritis (RA) is a chronic inflammatory disease of the synovial joints and is associated with considerable morbidity, disability and mortality. It affects approximately 1% of the population. Although its aetiology is unknown, our understanding of the cellular and molecular mechanisms that contribute to the pathogenesis in RA has allowed the development of new successful treatments for RA. However, there is an ongoing need for new therapeutics targeting different molecules, because a substantial number of patients remain with active disease.

Several lines of data suggest that granulocyte macrophage colony stimulating factor (GM-CSF) and macrophages strongly influence the development and pathogenesis of RA as reviewed recently.1–4 GM-CSF is a cytokine initially defined by its ability to promote the formation of myeloid cells from bone marrow precursor cells. In addition, GM-CSF mediates the functional activation of mature neutrophils, eosinophils and macrophages—and therefore is associated with a number of inflammatory disorders such as RA.5 Clinical support for a link between GM-CSF and inflammatory arthritis has been suggested through a number of case reports that describe exacerbation of established inflammatory disorders including RA following treatment with exogenous GM-CSF.6–8

GM-CSF functions through a high-affinity heterodimeric receptor composed of a GM-CSF receptor specific α-subunit (GM-CSFRα) and a signal-transducing subunit, the common β-chain (βc) that is shared with the receptors for interleukin 3 (IL-3) and IL-5.9–5 GM-CSF and its receptor are found in the plasma, synovial fluid and synovial tissue (ST) of patients with RA.10–12 Early phase clinical trials have suggested beneficial effects of GM-CSF or GM-CSFRα inhibition in patients with RA.13–15 Taken together, these data support the involvement of GM-CSF and its receptor in the pathogenesis of RA. To further explore the potential of targeting this axis as a novel therapy for RA we carefully investigated the cellular expression of GM-CSFRα on macrophages in RA ST compared with disease controls and studied the effects of GM-CSFRα inhibition in vitro as well as in an animal model of RA. The results strongly support the development of anti-GM-CSFRα antibody treatment as a new therapeutic strategy in RA.

METHODS

Patients

We obtained ST from 26 patients with RA classified according to the 1987 American College of Rheumatology criteria16 and from 24 patients with psoriatic arthritis (PsA) according to the Classification Criteria for Psoriatic Arthritis17 by mini-arthroscopy as described previously.18 All patients with RA and PsA had active disease defined by a disease activity score evaluated in 28 joints >3.2. We included ST of 9 healthy controls (HCs) and 10 patients with osteoarthritis (OA) undergoing exploratory arthroscopic surgery of a knee or
ankle as controls. We acquired written informed consent of all participants prior to study participation as approved by the Medical Ethical Committee of the Academic Medical Center/University of Amsterdam.

Immunohistochemistry

We fixed and stained frozen serial sections with mouse monoclonal antibodies to GM-CSFRα (clone 4H1; Lifespan BioSciences, Seattle, Washington, USA) and CD68 (clone EBM11; Dako, Glostrup, Denmark) to detect macrophages. GM-CSFRα was stained using a 2-step immunoperoxidase method and CD68 was stained using a 3-step immunoperoxidase method as described previously.19 Control sections were stained with isotype control matched primary antibodies. Stained sections were analysed by computer-assisted image analysis for all markers using the QWin image analysis system (Leica, Cambridge, UK) as previously described.20 The number of positive cells was measured as stained cell count/mm².

Immunofluorescence

To investigate GM-CSFRα expression by macrophages we incubated synovial biopsies of 5 patients with RA overnight at 4°C using primary antibodies specific for mouse antihuman GM-CSFRα (clone 4H1; Lifespan BioSciences), followed by Alexa Fluor 488 goat antimouse IgG2b antibody (Invitrogen, Carlsbad, California, USA). Subsequently, mouse monoclonal antibodies specific for macrophages (CD68; clone Y1/82A and CD163; clone GH/61; Biolegend, San Diego, California, USA) were incubated for 60 min at room temperature and were washed with flow assisted cell sorting (FACS) buffer (2% v/v). Monoclonal antibody (MAb) (BD Pharmingen, San Jose, California, USA) was added at 0.5 µg/well as a Fc block and incubated at 4°C for 30 min and 0.125 µg of antimouse-Ly6G (GR-1) fluorescein isothiocyanate (FITC) (eBioscience) were incubated with cells at 4°C for 1 h. Cells were washed in FACS buffer and fixed in 2% v/v formaldehyde in phosphate buffered saline (PBS) and analysed using FACS Canto II flow cytometer (BD Biosciences, San Jose, California, USA). Data were processed using FlowJo software (Tree Star, Ashland, Oregon, USA) and expressed as geometric mean.

In vivo margination assay

On Day 0 female BALB/c mice (n=7–8/group) were injected intraperitoneally with a dose response of CAM-3003 (10 mg/kg, 1 mg/kg and 0.1 mg/kg), CAT004 (isotype control 10 mg/kg) or vehicle alone. Twenty-four hours post dosing mice were injected subcutaneously with mouse GM-CSF (0.25 µg; ProSpec Tany TechnoGene Limited, Rehovot, Israel) and peripheral blood collected by cardiac puncture following terminal anaesthesia at 15 min post administration of recombinant GM-CSF. Differential blood cell counts were analysed on an ADVIA120 (Bayer, Tarrytown, New York, USA).

The effects of GM-CSFRα inhibition in the collagen induced arthritis model of RA

Male DBA/1 mice (Jackson Laboratory, Bar Harbor, Maine, USA) were dosed with 100 µg bovine type II collagen in Freund’s Complete Adjuvant at the base of the tail (two intradermal sites, 50 µL/site) followed by a subcutaneous injection of buprenorphine (0.1 mL/mouse) to induce arthritis. Onset of arthritis was determined as the 1st day that the clinical paw swelling score (range 0–4 per paw) was ≥1, typically 28±7 days post injection of collagen; each animal was then allocated alternately into the following treatment groups: vehicle; isotype control (CAT-004); CAM-3003 either 1 mg/kg of 10 mg/kg; or prednisolone (n=14/15 per group). Animals were dosed daily for 14 days and the clinical score was recorded daily.23

Mice were terminated at study end point and histological analyses of the rear ankle joints were performed to detect murine macrophages (clone Cl:A3-1, AbD Serotec, Kidlington, UK) (see online supplementary material for methods). All immunohistochemistry staining steps were carried out using a Dako AutoStainer (Glostrup, Denmark). Digital images were generated with an Aperio Scanscope XT scanning system (Vista, California, USA). The numbers of clearly distinguishable macrophages present within the annotated synovial membrane of the tibia-calcaneum were manually counted, up to an upper limit of 200 cells/mm². The number of macrophages present was then recorded as total macrophage count.

All mice were housed in specific pathogen-free conditions at MedImmune, Cambridge, in compliance with Home Office Regulations (UK). All animal experiments were approved by the Animal Welfare and Ethics Committee (Babraham Institute).

Data analysis

Patient data: Continuous data were tested for normality and described as mean±SD or median and IQR as appropriate. A Mann–Whitney U test was performed to compare differences in GM-CSFRα positive cell counts between RA and PsA versus non-inflammatory controls (OA and HC).

Mouse model data: one way analysis of variance with Dunn’s multiple comparison test was used to analyse joint score and histopathology counts from the collagen induced arthritis (CIA) model. The same statistical analyses were also used for the blood cell margination pharmacology model.

In all analyses a p value of <0.05 was considered statistically significant. All statistical analyses for in vitro and in vivo studies were performed using GraphPad PRISM 5 (GraphPad Software, La Jolla, California, USA).

RESULTS

GM-CSFRα overexpression in synovial tissue of patients with RA and PsA

Clinical characteristics of study patients are described in Table 1. The number of GM-CSFRα positive cells was significantly higher in ST from patients with active RA and PsA when compared with OA and HC (non-inflammatory controls) (Figure 1A). There was no statistically significant difference in the number of GM-CSFRα positive cells in RA compared with PsA (p=0.79; data not shown). Immunofluorescent labelling demonstrated colocalisation with the macrophage markers CD68 and CD163 (Figure 1B). No significant correlation was observed with disease activity parameters (disease activity score in 28 joints or C reactive protein) (data not shown).

Table 1  Patient characteristics

<table>
<thead>
<tr>
<th>Age (median (IQR))</th>
<th>RA (N=26)</th>
<th>PsA (N=24)</th>
<th>OA (N=10)</th>
<th>HC (N=9)</th>
</tr>
</thead>
<tbody>
<tr>
<td>54 (44 – 62)</td>
<td>55 (42 – 61)</td>
<td>74 (67 – 76)</td>
<td>51 (33 – 61)</td>
<td></td>
</tr>
<tr>
<td>Sex, Female (%)</td>
<td>50%</td>
<td>38%</td>
<td>91%</td>
<td>11%</td>
</tr>
<tr>
<td>DAS28 (mean ±SD)</td>
<td>5.4±1.0</td>
<td>4.4±1.6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ESR, mm/h (median (IQR))</td>
<td>27 (17–40)</td>
<td>17 (9–36)</td>
<td></td>
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<tr>
<td>IgM RF positives (%)</td>
<td>58%</td>
<td>4%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anti-CCP positives (%)</td>
<td>54%</td>
<td>0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>NSAID only (%)</td>
<td>27%</td>
<td>50%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>DMARD (%)</td>
<td>62%</td>
<td>38%</td>
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<tr>
<td>Biological (%)</td>
<td>12%</td>
<td>13%</td>
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</tbody>
</table>

Categorical variables: n (%)  Continuous variables (data not normally distributed): median (IQR)  Continuous variables (data normally distributed): mean (±SD).

Anti-CCP, anticyclic citrullinated peptide antibodies; CRP, C reactive protein; DAS28, disease activity score in 28 joints; DMARD, disease modifying antirheumatic drug; ESR, erythrocyte sedimentation rate; HC, healthy controls; IgM RF, IgM rheumatoid factor; NSAID, non-steroidal anti-inflammatory drug; OA, osteoarthritis; PsA, psoriatic arthritis; RA, rheumatoid arthritis.

Figure 1  (A) The quantification of granulocyte macrophage colony stimulating factor (GM-CSFRα) positive cells in ST from patients with rheumatoid arthritis (RA) and psoriatic arthritis (PsA) (inflammatory arthritis; n=50) compared with ST from patients with osteoarthritis (OA) and healthy controls (HCs) (non-inflammatory; n=19). Median (IQRs) are shown. *p Value <0.05 (Mann–Whitney U test). (B) Representative picture of an immunofluorescence staining for GM-CSFRα (green). GM-CSFRα shows colocalisation with either CD68 (red) or CD163 (red) (arrows).

Figure 2  Characterisation of the cell surface expression of CD11b on mouse granulocytes stimulated with mouse granulocyte macrophage colony stimulating factor (GM-CSF). Mouse granulocytes were stimulated with a dose response of mouse GM-CSF for 1 hr and cell surface expression of CD11b quantified by flow cytometry using the mean fluorescent intensity (MFI). GM-CSF was able to dose dependently increase CD11b expression. (B) Mouse granulocytes were stimulated with mouse GM-CSF corresponding to the EC80 (2.5 ng/mL) and incubated with increasing concentrations of CAM-3003 for 1.5 h. Cells were then labelled for CD11b expression and the MFI quantified by flow cytometry. CAM-3003 dose-dependently inhibited GM-CSF induced CD11b upregulation.
GM-CSFRα inhibition reduces disease severity in the CIA model of RA

To investigate the effect of GM-CSFRα inhibition, mice that were subjected to CIA were treated with CAM-3003 for 14 days after the first signs of arthritis. A statistically significant inhibition of clinical score was observed with maximal effect at 10 mg/kg intraperitoneally when compared with vehicle (figure 4A, B). This reduction in clinical score was equivalent to anti-GM-CSF and anti-tumour necrosis factor (TNF) inhibition in this model (see online supplementary figure S3). In addition, mice treated with CAM-3003 had less evidence of cellular infiltration, cartilage damage and bone erosion (figure 4C) and a significant reduction in the number of F4/80 positive macrophages when compared with isotype control (figure 4D). Splenomegaly was observed in arthritic animals and this was significantly reduced in mice treated with prednisolone and mice treated with 10 mg/kg of CAM-3003 (see online supplementary figure S4A). Thoracic and inguinal lymph node weights were pooled and weighed. Anti-GM-CSF treated animals showed no reduction in lymph node weights in contrast to prednisolone treated animals (see online supplementary figure S2) when compared with vehicle using a one way analysis of variance. However, in arthritic mice the absolute neutrophil count was significantly lower in mice treated with CAM-3003 10 mg/kg when compared with isotype treated mice (see online supplementary figure S3) using a t test. There was no statistically significant reduction in anti-type II collagen antibody levels between treatment groups (see online supplementary Figure S6).

DISCUSSION

The results presented here show an increased number of GM-CSFRα expressing cells in ST of patients with either RA or...
PsA when compared with non-inflammatory controls. GM-CSFRα expression in ST in RA and OA has been described before, but this is the first study to ascertain that GM-CSFRα positive cell numbers are higher in inflammatory arthritis versus non-inflammatory arthritis. In addition GM-CSFRα expression in PsA ST has not been described before. In addition, we have shown expression of GM-CSFRα by CD68 positive and CD163 positive macrophages. Of importance, the number of CD68 positive macrophages, one of the target cells for GM-CSF, correlates with disease activity in RA and the number of these cells reduce in the inflamed synovium following successful treatment. Together with the notion that the infiltration of the synovium by macrophages is similar between patients with RA and PsA, our results suggest that targeting GM-CSFRα could be effective in patients with RA and PsA.

Cell trafficking and retention of inflammatory cells play a key role in synovial inflammation. We found that mouse granulocytes incubated with CAM-3003, a GM-CSFR antagonist, could concentration-dependently inhibit GM-CSF induced upregulation of the integrin CD11b, which could indicate that blockade...
of GM-CSFRα can reduce synovial inflammation via blocking the actions of its ligand. Moreover, we showed with our mouse model that GM-CSFR blockade with CAM-3003 prevented the marginalization response of recombinant GM-CSF of white blood cells, consistent with observations in man and cynomolgus monkey. As we (data not shown) and others have shown that GM-CSF is elevated within the SF of patients with RA it appears likely that peripheral white blood cells coated in antibodies to the receptor would not respond to locally high concentrations of GM-CSF and therefore reduced margination and ultimately migration, diapedesis and retention within the joint would be expected to occur.

In line with the notion that anti-GM-CSFRα antibody treatment could have a beneficial effect on arthritis, we found in the murine CIA model of RA that CAM-3003 treatment dose-dependently inhibited clinical signs and symptoms of arthritis, with reduced synovial inflammation and joint destruction. This effect is probably due to inhibition of trafficking and retention of inflammatory cells, and secondary to other effects such as reduced survival of fully differentiated macrophages within the joint. These observations are supported by previous work demonstrating a reduction in inflammatory cell numbers in the joint and a reduction in Ly-6Chigh and Ly-6Clow monocytes in joint homogenates after GM-CSF blockade. A 14.7% reduction in spleen weight was also observed consistent with reduced inflammation in these animals. Splenomegaly has previously been described in a subset of patients with RA. The significance of this reduction has yet to be determined. There were no statistically significant changes in peripheral cell counts or body weight in mice with arthritis treated with the GM-CSFRα antibody throughout the study, however, whereas neutropenia was not observed, a slight reduction in neutrophils was observed in arthritic mice after CAM-3003 administration, suggesting that close monitoring of these cells in clinical studies with a GM-CSFRα inhibitor is warranted.

Taken together, these results further validate the GM-CSF–GM-CSFRα pathway as a new therapeutic target for the treatment of RA and perhaps PsA. Recent phase 1 and phase 2 clinical trials targeting either GM-CSF or GM-CSFRα support the development of new treatments interfering with this pathway.

Competing interests PPT and DMG became employees of GlaxoSmithKline after study completion. GSK has not been involved in this study, but does have a proprietary interest in MOR103, an anti-GM-CSF antibody, by virtue of an alliance with Morphosys. ESC, ND, AL, SH, DC, AD, EJ and MAS are employees of MedImmune, a wholly owned subsidiary of AstraZeneca. IKA, AN, JC, JW and MMC were at the time of completing this work full-time employees of MedImmune, a wholly owned subsidiary of AstraZeneca. IPW received grants from CSL during the conduct of the study; grants from CSL, from null, outside the submitted work. MedImmune and AstraZeneca has an anti human GM-CSFRα chain antibody, mavrilimumab, in clinical development in rheumatoid arthritis.

Ethics approval Ethics committee of the AMC.

Provenance and peer review Not commissioned; externally peer reviewed.

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Supplementary files

**Preclinical characterisation of the GM-CSF receptor as a therapeutic target in rheumatoid arthritis**

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**Supplementary methods**

**Pharmacokinetic analysis of a mouse anti-GM-CSFR\(\alpha\) antibody (CAM-3003)**

We generated an anti-mouse GM-CSFR neutralising antibody (CAM-3003) by immunising rats and subcloning the resultant neutralising anti mouse GM-CSFR rat IgG2a into a mouse IgG1 backbone to match the mechanism of action of mavrilimumab and to allow chronic dosing in vivo with a reduced potential for immunogenicity. The potency of the engineered mouse antibody, CAM-3003, was 130pM (95% CI: 99 to 171pM) [32]. The pharmacokinetic profile of the antibody was determined in a single ascending dose study in female CD1(ICR) mice (supplemental table 1). Female CD1(ICR) mice (Charles River, Margate, UK) (n=4/time-point) were given single intravenous (i.v.) doses of CAM-3003 at 1, 10 or 100mg/kg. Single serum samples were obtained from individual animals at intervals up to 30 days post-dose. Samples obtained on sacrifice at each time-point were assayed separately for CAM-3003 concentration. Results for each scheduled time-point and dose were then pooled for pharmacokinetic analysis of the geometric means (supplemental figure 1). Time-course pharmacokinetic data-sets were analysed using WinNonlin Pro (v4.1; Pharsight Corporation, St. Louis, MO).
Supplementary Table 1. Pharmacokinetic parameters of the anti GM-CSFRα antibody (CAM-3003) in female CD1(ICR) mice.

<table>
<thead>
<tr>
<th>Route of Administration (i.v.)</th>
<th>Dose (mg/kg)</th>
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<tbody>
<tr>
<td>Parameter</td>
<td>Units</td>
</tr>
<tr>
<td>Tmax</td>
<td>Days</td>
</tr>
<tr>
<td>Cmax</td>
<td>µg/ml</td>
</tr>
<tr>
<td>Cmax / Dose</td>
<td>kg/L</td>
</tr>
<tr>
<td>AUC last</td>
<td>µg.d/ml</td>
</tr>
<tr>
<td>AUC last / Dose</td>
<td>kg.d/L</td>
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<tr>
<td>T½</td>
<td>Days</td>
</tr>
<tr>
<td>AUC inf</td>
<td>µg.d/ml</td>
</tr>
<tr>
<td>AUC inf / Dose</td>
<td>kg.d/L</td>
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Tmax, time to maximum plasma concentration; Cmax, peak plasma concentration; Cmax/dose, maximum plasma concentration per dose; AUC last, area under the plasma-concentration versus time curve from zero time until the last measurable concentration; T½, half-life; AUC inf, Area under the plasma concentration versus time curve from zero to infinity.
Immunohistochemistry for synovial tissue macrophages in a collagen induced arthritis (CIA) model for RA upon GM-CSF inhibition

The rear ankle joints (male DBA/1 mice) were removed and fixed for up to 48 hours in 10% neutral buffered formalin. Joints were transferred to 50mM EDTA (pH 6.0) for up to 5 weeks to undertake decalcification. Following decalcification, joints were processed and embedded into paraffin wax. The joints were orientated so that a complete cross-section of the joint including tibia, talus, calcaneum and synovial membranes were clearly evident. 3µm sections were cut and stained with haematoxylin and eosin. Sections were dewaxed, rehydrated and treated with 3% hydrogen peroxide in methanol to block endogenous peroxidases. Following rinsing, the sections were incubated with 20µg/ml Proteinase K for 20 minutes for antigen retrieval. After rinsing the sections were incubated in 2.5% bovine serum albumin (BSA) for 20 minutes followed by a 60 minute incubation in monoclonal rat anti-F4/80 antibody (1µg/ml) to detect murine macrophages (clone Cl:A3-1, AbD Serotec, Kidlington, UK). Parallel control sections were incubated with a matched isotype antibody. The sections were then rinsed and incubated with goat anti-rat IgG (Stratech, Suffolk, UK) at 2µg/ml for 30 minutes, followed by Goat ImmPress HRP polymer (Vector Laboratories, Peterborough, UK) for 30 minutes. Staining was developed incubating with diaminobenzidine (Dako, Glastrup, Denmark). Sections were counterstained with GILLS I haematoxylin, dehydrated and mounted using xylene-based mounting media (DePeX).
Supplementary Table 2. Description of the effect of GMCSFR inhibition on peripheral blood cell counts in established arthritis.

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</thead>
<tbody>
<tr>
<td>Naïve</td>
<td>-</td>
<td>2.49 ± 0.51</td>
<td>5.45 ± 2.37*</td>
<td>0.74 ± 0.16</td>
<td>1.59 ± 0.35</td>
<td>0.07 ± 0.012</td>
<td>0.06 ± 0.009</td>
<td>0.00 ± 0</td>
<td>512.60 ± 203.3***</td>
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<tr>
<td>Vehicle</td>
<td>-</td>
<td>3.38 ± 0.366</td>
<td>10.18 ± 0.76</td>
<td>1.25 ± 0.19</td>
<td>1.78 ± 0.23</td>
<td>0.17 ± 0.076</td>
<td>0.07 ± 0.009</td>
<td>0.03 ± 0.019</td>
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<tr>
<td>Prednisolone</td>
<td>3</td>
<td>1.67 ± 0.306*</td>
<td>10.23 ± 1.25</td>
<td>1.09 ± 0.16</td>
<td>0.54 ± 0.07**</td>
<td>0.05 ± 0.01</td>
<td>0.05 ± 0.008</td>
<td>0.03 ± 0.017</td>
<td>872.88 ± 120.04*</td>
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<tr>
<td>CAM3003</td>
<td>1</td>
<td>3.04 ± 0.385</td>
<td>9.71 ± 0.95</td>
<td>1.09 ± 0.15</td>
<td>1.64 ± 0.26</td>
<td>0.09 ± 0.014</td>
<td>0.06 ± 0.011</td>
<td>0.02 ± 0.013</td>
<td>1020.54 ± 102.67</td>
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<tr>
<td>CAM3003</td>
<td>10</td>
<td>2.87 ± 0.407</td>
<td>9.57 ± 1.01</td>
<td>0.85 ± 0.18</td>
<td>1.62 ± 0.28</td>
<td>0.08 ± 0.01</td>
<td>0.06 ± 0.01</td>
<td>0.01 ± 0.002</td>
<td>860.15 ± 117.81</td>
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<tr>
<td>Isotype Control</td>
<td>10</td>
<td>3.71 ± 0.311</td>
<td>10.91 ± 0.61</td>
<td>1.35 ± 0.17</td>
<td>1.90 ± 0.24</td>
<td>0.18 ± 0.08</td>
<td>0.08 ± 0.017</td>
<td>0.03 ± 0.014</td>
<td>1105.17 ± 70.44</td>
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</table>

Mice were treated daily for 14 days post onset of arthritis with either anti-GM-CSFR antibody (CAM-3003) at 1 or 10 mg/kg, isotype control (CAT-004) at 10mg/kg and 3mg/kg Prednisolone by oral gavage. Peripheral blood was sampled at Day 14 by cardiac puncture under terminal anaesthesia for ADVIA (Manufacturer) and blood cell counting. Data was analysed using an ANOVA with Dunnet’s multiple comparison test comparing vehicle versus all groups. *p<0.05, **p<0.01 & ****p<0.0001
Supplementary Figure 1. The non-linear pharmacokinetic profile of a single ascending dose of CAM-3003, an anti-mouse GM-CSFR monoclonal antibody. Data are plotted as mean ± SEM (n=4 /group/time point).

Supplementary Figure 2 shows representative images of the areas of quantification typically observed in a non-inflamed paw (A) and an arthritic inflamed paw (B). The area of synovial tissue analysed was determined using the Aperio Scanscope XT digital image scanning. The numbers of clearly distinguishable macrophages present within the annotated synovial membrane of the tibia-calcaneum were counted (see article figure 4).
Supplementary Figure 3. Characterisation of the effect of either GM-CSF or TNF α inhibition in established arthritis. a) Mice were treated daily for 14 days post the onset of arthritis with either an anti-GM-CSF antibody (Clone 22E9) at 100mcg daily or CAT-004 (100μg) as an isotype control. Mean clinical score was plotted daily to map disease progression. b) Mice were treated daily for 14 days post the onset of arthritis with either etanercept at 0.5mg/kg daily or CAT-004 (10mg/kg) as an isotype control. Mean clinical score was plotted daily to map disease progression.

Supplementary Figure 4. Characterisation of the effect of GM-CSFR inhibition on lymphoid organ weights in established arthritis. Mice were treated daily for 14 days post onset of arthritis with either anti-GMCSFR antibody (CAM-3003) at 1 or 10 mg/kg, isotype control (CAT-004) at 10mg/kg and 3mg/kg Prednisolone by oral gavage. A) Spleen weights were measured at Day 14 post
onset of arthritis and B) Lymphnodes, 2 thoracic and 2 inguinal, were pooled and weighed at Day 14. Data shown are represented as mean weight ± SEM. Data were analysed using an ANOVA with Dunnet’s multiple comparison test comparing vehicle versus all groups. *p<0.05, **p<0.01 & ***p<0.001.

Supplementary Figure 5. Characterisation of the effect of GM-CSFR inhibition on peripheral blood neutrophil counts in established arthritis. Mice were treated daily for 14 days post onset of arthritis with either anti-GMCSFR antibody (CAM-3003) at 1 or 10 mg/kg, isotype control (CAT-004) at 10mg/kg and 3mg/kg Prednisolone by oral gavage. Peripheral blood was sampled at Day 14 by cardiac puncture under terminal anaesthesia for differential blood cell
count (ADVIA; Manufacturer) including neutrophil cell counting. Data was analysed using a Two-tailed unpaired T-Test comparing isotype control versus 10mg/kg CAM-3003 group. *p<0.05.

Supplementary Figure 6. Characterisation of the effect of GM-CSFR inhibition on anticollagen type II antibody responses. Mice were treated daily for 14 days post onset of arthritis with either anti-GM-CSFR antibody (CAM-3003) at 10 mg/kg, isotype control (CAT-004) at 10mg/kg and 3mg/kg Prednisolone by oral gavage. Sera was collected from peripheral blood and analysed for levels of anti-collagen II antibodies by ELISA as per manufactures protocol (Chondrex Inc, Redmond, WA). As the variances between groups were significantly different a non-parametric Mann-Whitney test was used between Naive and Vehicle groups. No significant difference was observed between Vehicle, prednisolone, CAM-3003 and isotype control groups. **p<0.01