Suppression of inflammation reduces endothelial microparticles in active systemic lupus erythematosus

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ABSTRACT

Background In a prospective observational study, we investigated whether patients with active systemic lupus erythematosus (SLE) had higher indices of endothelial damage and dysfunction than healthy controls and whether improved disease control was associated with improvement in these indices.

Methods Twenty-seven patients with active SLE (four or more American College of Rheumatology (ACR) criteria) and 22 age-matched controls were assessed. Endothelial microparticles (EMPs: CD31+/annexin V+/CD42b−) were quantified using flow cytometry. Brachial artery flow-mediated dilatation (FMD) was measured using automated edge-tracking software. Twenty-two patients had a second assessment at a median (IQR) of 20 (16, 22) weeks after initiating new immunosuppressive therapy.

Results SLE patients had a median (IQR) baseline global British Isles Lupus Assessment Group Disease Activity Index (BILAG-2004) score of 14 (12, 22). CD31+/annexin V+/CD42b− EMPs were higher (157 548/ml (59 906, 272 643) vs 41 025 (30 179, 98 082); p=0.003) and endothelial-dependent FMD was lower (1.63% (0.98, 5.09); p=0.1) at the second visit.

Conclusions Active SLE is associated with evidence of increased endothelial damage and endothelial dysfunction, which improved with suppression of inflammation. Better control of active inflammatory disease may contribute to improved cardiovascular risk in patients with SLE.

INTRODUCTION

Women with systemic lupus erythematosus (SLE) have a greater than fivefold increased risk of coronary heart disease (CHD) events and, notably, there is a 50-fold increased risk in younger patients.1 SLE patients also have an increased burden of subclinical atherosclerosis, as measured by coronary calcium score, carotid plaque, and arterial stiffness.2-5 Although classic Framingham risk factors are more prevalent in SLE,6 they do not fully explain this excess CHD risk,2 and SLE is therefore considered an independent risk factor for cardiovascular disease (CVD).

The increased CVD risk is multifactorial,8 and the vascular endothelium may be a key interface between inflammation and atherogenesis in SLE.9 Endothelial dysfunction represents the earliest clinically detectable stage in the development of atherosclerosis, and reduced flow-mediated dilatation (FMD) of conduit arteries is associated with traditional CHD risk factors10-12 and prevalent CVD13 and predicts future coronary events.14 We and others have demonstrated impaired FMD in patients with SLE, and, although FMD improves with statins15 and omega-3 fatty acids,16 the contribution of inflammation to endothelial dysfunction has yet to be assessed in an SLE population.

Endothelial microparticles (EMPs) are membrane-bound subcellular microparticles (MPs) produced by endothelial cells in response to a variety of triggers, and may act as a biomarker for endothelial damage. EMPs are increased in acute inflammatory disease activity would result from the prothrombotic environment in antiphospholipid syndrome22 and contribute to the immune-pathogenesis of SLE.26 To date, there are no studies examining whether EMPs act as biomarkers of endothelial damage and cardiovascular risk in SLE.

In the present study, we investigated whether active SLE is associated with increased endothelial damage (EMPs) and dysfunction (FMD), compared with age- and gender-matched controls. We subsequently examined the hypothesis that improvements in inflammatory disease activity would result in improvements in these measures in a prospective longitudinal observational study, and assessed the relationship between endothelial damage, endothelial dysfunction and disease activity.

METHODS

Patients Patients with SLE (four or more 1997 American College of Rheumatology (ACR) revised criteria27) were recruited from The Kellgren Centre for Rheumatology, Central Manchester NHS Foundation Trust (CMFT) and East Lancashire Hospitals NHS Trust. Patients were included if they had active SLE sufficient for their treating physician to initiate a change in their immunosuppressive therapy. Specifically, we included patients starting treatment with any of azathioprine, mycophenolate.

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mofetil, cyclophosphamide or rituximab. Healthy, age-matched controls were recruited from friends of patients or from staff at the University of Manchester and CMFT. We excluded subjects with a recent acute infection (≤1 month), recent cardiovascular event (≤3 months), any chronic infection, pregnant/lactating patients, and patients with chronic kidney disease (estimated glomerular filtration rate ≤20 ml/min). All subjects gave written informed consent, and ethics approval was obtained from Oldham Research Ethics Committee.

Clinical and laboratory assessment

Patients with SLE were assessed before the change in therapy and again approximately 4–5 months later. The referring rheumatologist dictated all changes in therapy. Control subjects were assessed once. Subjects fasted for 12 h before the study, and were asked to abstain from smoking tobacco on the morning of the study. All subjects underwent a full history and physical examination at each visit, with CVD history and drug exposures documented. Patients underwent detailed assessment of their current and past SLE features and therapeutic history. Two composite scores of disease activity were recorded at each visit: the British Isles Lupus Assessment Group Disease Activity Index (BILAG-2004)28 and the SLE Disease Activity Index 2000 (SLEDAI-2K).29 We report the original BILAG-2004 and the newer global BILAG-2004 score.30 Cumulative damage was recorded using the Systemic Lupus International Collaborating Clinics/ACR-Damage Index.31 A fasting blood sample was drawn for assessment of blood glucose, lipid profile, erythrocyte sedimentation rate, complement levels and autoantibody titres by CMFT laboratories. Adiponectin, vascular cell adhesion molecule-1 (VCAM-1) and vascular endothelial growth factor (VEGF) were measured using DuoSet ELISA development kits (R&D Systems, Abingdon, UK). Plasma soluble endothelial protein C receptor (ePCR) was measured by ELISA (Adipobiotech, Beijing, China). Fasting serum insulin was measured by ELISA (ultrasensitive solid phase ELISA; DRG, Marburg, Germany), and the homoeostatic model assessment of insulin resistance (HOMA-IR) was calculated using the HOMA2 formula.

Assessment of endothelial damage and dysfunction

The methodologies used to assess endothelial damage and dysfunction are described in full in online supplementary file 1. In brief, EMPs were quantified in platelet-poor plasma, generated using a two-step centrifugation regimen. These platelet-poor plasma samples were stored at −80°C and analysed in batches. EMPs were enumerated (n/ml) by flow cytometry, after the addition of counting beads, and defined as CD31-positive, annexin V-positive and CD42b-negative events. FMD of the brachial artery was performed using B-mode ultrasound and automated edge-tracking software as per guidelines.33 The brachial artery was imaged longitudinally proximal to the antecubital fossa, and the probe fixed in place. A blood pressure (BP) cuff was placed around the mid-forearm. After the baseline diameter had been recorded, the BP cuff was inflated to 50 mm Hg above systolic BP, to at least 200 mm Hg, for 5 min and then deflated. Reactive hyperaemia was confirmed using Doppler ultrasound. The brachial artery diameter was recorded continuously, and endothelial-dependent brachial artery dilatation (EDD) was recorded 60 s after cuff deflation. Peak diameter was also recorded if this did not occur at 60 s. Sublingual glyceryl trinitrate was then used to assess endothelial-independent brachial artery dilatation.

Statistical analysis

A sample size calculation was performed for a co-primary outcome (change in FMD (%) over time) using data from preliminary validation studies.33 On the basis of a mean (SD) FMD (%) of 7.01% (1.77) on serial FMD measurements within one individual, a sample size of 13 patients would give 80% power at 5% significance to detect a 2% change in FMD over time. A recruitment target of 30 cases was set to allow for a 20% dropout rate. Fifteen healthy controls would provide 80% power to detect a 2% difference in FMD (%) between cases and controls. Significance of between-group differences was assessed using a two-sided t test or Mann–Whitney U test for continuous variables and χ² test for categorical variables. Correlation between measures was assessed using Spearman’s correlation coefficient. Linear regression was used to assess the association between SLE and endothelial damage/dysfunction. Data analysis was performed using the Stata V10 software package.

RESULTS

Twenty-seven patients with active SLE and 22 healthy controls were assessed. Baseline characteristics of cases and controls are described in tables 1 and 2. Fourteen (51.9%) patients were starting standard immunosuppression, six of whom were naïve to immunosuppressant therapy. Thirteen (48.1%) patients were starting rituximab, all of whom had either received (and had failed to respond to) standard immunosuppressant therapy (n=10) or were receiving a further cycle of therapy with biological agent (n=3). The most common indications for changing therapy were active lupus arthritis (n=8) and active lupus nephritis (n=9).

### Table 1 Demographic, clinical and laboratory variables of SLE cases and age- and gender-matched healthy controls

<table>
<thead>
<tr>
<th>Variable</th>
<th>SLE patients (n=27)</th>
<th>Controls (n=22)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years), mean (SD)</td>
<td>41.5 (14.1)</td>
<td>38.5 (9.3)</td>
<td>0.56</td>
</tr>
<tr>
<td>Female</td>
<td>26 (96.3)</td>
<td>19 (86.3)</td>
<td>0.96</td>
</tr>
<tr>
<td>Caucasian</td>
<td>17 (63.0)</td>
<td>20 (91.0)</td>
<td>0.20</td>
</tr>
<tr>
<td>BP systolic (mm Hg)</td>
<td>131 (103, 144)</td>
<td>119 (114, 127)</td>
<td>0.78</td>
</tr>
<tr>
<td>BP diastolic (mm Hg)</td>
<td>76 (66, 86)</td>
<td>77 (69, 80)</td>
<td>0.84</td>
</tr>
<tr>
<td>AHT therapy</td>
<td>12 (44.0)</td>
<td>0 (0)</td>
<td>0.002</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>25.7 (21.5, 28.5)</td>
<td>25.1 (23.1, 30.8)</td>
<td>0.42</td>
</tr>
<tr>
<td>WC (cm)</td>
<td>80 (74, 91.6)</td>
<td>80 (72, 93.5)</td>
<td>0.86</td>
</tr>
<tr>
<td>Total cholesterol (mmol/l)</td>
<td>5.2 (4.6, 6.7)</td>
<td>5.54 (4.83, 6.87)</td>
<td>0.54</td>
</tr>
<tr>
<td>HDL-cholesterol (mmol/l)</td>
<td>1.33 (1.15, 1.61)</td>
<td>1.66 (1.46, 1.78)</td>
<td>0.06</td>
</tr>
<tr>
<td>LDL-cholesterol (mmol/l)</td>
<td>2.71 (1.92, 3.6)</td>
<td>3.01 (2.70, 3.58)</td>
<td>0.35</td>
</tr>
<tr>
<td>Triglycerides (mmol/l)</td>
<td>1.36 (0.90, 1.87)</td>
<td>0.88 (0.64, 1.00)</td>
<td>0.01</td>
</tr>
<tr>
<td>Lipid-lowering therapy</td>
<td>3 (11)</td>
<td>0 (0)</td>
<td>0.18</td>
</tr>
<tr>
<td>Glucose (mmol/l)</td>
<td>4.40 (4.1, 5.0)</td>
<td>4.9 (4.8, 5.1)</td>
<td>0.05</td>
</tr>
<tr>
<td>Diabetes</td>
<td>2 (7.4)</td>
<td>0 (0)</td>
<td>0.08</td>
</tr>
<tr>
<td>Family history of CVD</td>
<td>10 (43.5)</td>
<td>4 (18.2)</td>
<td>0.09</td>
</tr>
<tr>
<td>MetS</td>
<td>11 (40.7)</td>
<td>1/15 (6.7)</td>
<td>0.03</td>
</tr>
<tr>
<td>Fasting insulin (mU/l)</td>
<td>16.3 (12.4, 22.9)</td>
<td>14.0 (12.2, 20.6)</td>
<td>0.55</td>
</tr>
<tr>
<td>HOMA2-IR</td>
<td>2.3 (1.9, 3.0)</td>
<td>2.0 (1.8, 3.5)</td>
<td>0.50</td>
</tr>
<tr>
<td>Adiponectin (mg/l)</td>
<td>3.57 (2.46, 5.90)</td>
<td>2.93 (2.42, 3.60)</td>
<td>0.12</td>
</tr>
<tr>
<td>hsCRP (mg/l)</td>
<td>2.4 (0.5, 5.1)</td>
<td>0.51 (0.26, 2.83)</td>
<td>0.09</td>
</tr>
</tbody>
</table>

Unless otherwise stated, values are n (%) or median (IQR).

AHT, anti-hypertensive; BMI, body mass index; BP, blood pressure; CVD, cardiovascular disease; HDL, high-density lipoprotein; HOMA2-IR, homoeostatic model assessment of insulin resistance; hsCRP, high-sensitivity C-reactive protein; LDL, low-density lipoprotein; MetS, metabolic syndrome; SLE, systemic lupus erythematosus; WC, waist circumference.
### Table 2  Clinical and immunological features of SLE patients at entry into study (n=27)

<table>
<thead>
<tr>
<th>Feature (n=27)</th>
<th>n (%) or median (IQR)*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Disease duration (years)</td>
<td>7.0 (3.5, 12)*</td>
</tr>
<tr>
<td>ANA-positive ever</td>
<td>27 (100)</td>
</tr>
<tr>
<td>ANA-positive at baseline visit</td>
<td>23 (85.2)</td>
</tr>
<tr>
<td>Elevated anti-dsDNA antibody†</td>
<td>7 (25.9)</td>
</tr>
<tr>
<td>Low C3†</td>
<td>3 (11.1)</td>
</tr>
<tr>
<td>Low C4†</td>
<td>10 (37.0)</td>
</tr>
<tr>
<td>Anti-cardiolipin antibody-positivete</td>
<td>8 (29.6)</td>
</tr>
<tr>
<td>Total annexin V+ MPs (n/ml)</td>
<td>2 (3.7)</td>
</tr>
<tr>
<td>Total IL-6 (pg/ml)</td>
<td>14 (12, 22)*</td>
</tr>
<tr>
<td>SLEDAI-2K</td>
<td>6 (5,13)*</td>
</tr>
<tr>
<td>SLICC/ACR-DI</td>
<td>1 (1.2)*</td>
</tr>
<tr>
<td>Oral corticosteroids</td>
<td>24 (88.9)</td>
</tr>
<tr>
<td>Average daily corticosteroid dose (mg)</td>
<td>12.5 (10, 17.5)*</td>
</tr>
<tr>
<td>Current immunosuppressant use</td>
<td>12 (44.4)</td>
</tr>
<tr>
<td>Current antimalarial use</td>
<td>20 (74.1)</td>
</tr>
</tbody>
</table>
| *Defined as positive/present at study entry unless stated. †As per laboratory reference range. ACR, American College of Rheumatology; ANA, antinuclear antibodies; BILAG, British Isles Lupus Assessment Group Disease Activity Index; DI, Damage Index; dsDNA, double-stranded DNA; SLE, systemic lupus erythematosus; SLEDAI-2K, SLE Disease Activity Index 2000; SLICC, Systemic Lupus International Collaborating Clinics.

### Endothelial damage and dysfunction in SLE

Patients with active SLE had significantly elevated CD31+/annexin V+ EMP counts and reduced flow-mediated dilatation (FMD) compared with controls (1146 Parker B, Figure 1). Reduced EMP count and FMD were similar (table 3). There was a moderate correlation between EMP count and FMD (%): $r^2=-0.40; p=0.006$ in all subjects, which was similar in SLE patients only ($r^2=-0.42; p=0.008$). There was also a significant correlation between VCAM-1 and CD31+/annexin V+/CD42b− EMP count ($r^2=0.28; p=0.03$). EMP count did not correlate with global BILAG-2004 score ($r^2=0.19, p=0.20$) or SLEDAI-2K ($r^2=0.16, p=0.27$). In a multiple regression model including SLE, age, BP, total cholesterol, glucose and estimated glomerular filtration rate, SLE was independently associated with CD31+/annexin V+/CD42b− EMP levels (B coefficient 145 (29, 260); $p=0.02$).

### Figure 1  (A) Reduced flow-mediated dilatation (FMD) and (B) elevated CD31+/annexin V+/CD42b− endothelial microparticles (EMPs) in patients with systemic lupus erythematosus (SLE) versus controls.

**Effect of improved disease control on endothelial damage and dysfunction in active SLE**

Twenty-two (81.5%) patients returned for follow-up after a median (IQR) interval of 20 (16, 22) weeks; their characteristics are described in table 4. Twelve patients received standard immunosuppressive therapies and 10 received rituximab. The characteristics of patients who failed to return did not differ from those followed-up (data not shown). Disease activity improved significantly over time. Median (IQR) change in global BILAG-2004 score was $-11 (-18, -3)$, and change in SLEDAI-2K was $-5 (-9, -2)$. No significant changes in traditional CHD risk factors, obesity indices, insulin metabolism or brachial artery diameter were observed over the study period. Although the median (IQR) daily prednisolone dose at follow-up remained stable at follow-up (12.5 (10, 17.5) mg; median change 0 (0–2.5) mg), dose changes were observed within individuals. The dose remained identical in nine patients, changed by ±2.5 mg in eight patients, and changed by ±5 mg in four patients. Oral prednisolone was stopped in just one individual. Total annexin V+ MPs declined over time (991 513 (492 455, 1 227 363) vs 55 655 (29 475, 188 659); $p=0.03$). One of these patients had lupus nephritis, and both received rituximab. Median (IQR) change in absolute CD31+/annexin V+/CD42b− EMP count
was −87 998/ml (−184 433, +4949), and percentage change was −63.8% (−79.6, 1.6). Median (IQR) FMD (%) at 60 s also improved over time (0.64% (−2.31, 4.47) vs 4.56% (1.71, 5.87); p=0.10), and the median (IQR) change in EDD was +3.34% (−1.61, 6.2). Although not our primary outcome, we also noted that patients treated with rituximab had a non-significant greater improvement in global BILAG-2004 score (median (IQR) change 13 (−25, −10) vs −5 (−16, 0); p=0.18), EMP count (median (IQR) change −107 549 (−184 433, 50 000) vs −81 886 (−189 393, −14 654); p=0.18), and FMD (+4.76% (+3.54, +6.3) vs +1.76% (−1.61, +3.89; p=0.28), compared with standard therapy. Overall, there was a moderate correlation between change in EMP count (%) and change in global BILAG-2004 score (r²=0.40 p=0.08), but change in EMP (%) did not correlate with change in FMD (%) (r²=0.11 p=0.71).

**DISCUSSION**

As far as we are aware, this is the first study to describe CD31+/-annexin V+/-CD42b− EMPs over time in an SLE cohort with active disease, and to examine their relationship with other markers of endothelial function. Patients with active SLE had significantly higher levels of circulating CD31+/-annexin V+/-CD42b− EMPs, independent of age and traditional CHD risk factors. Patients with active SLE also had significantly impaired FMD (%), as has been previously reported.³⁻⁴⁻⁵ We also noted a modest inverse correlation between these two measures, both in the whole cohort and in SLE patients alone. Although controls were well matched for age/gender, differences were noted at baseline between the two groups, which may have had an impact on baseline endothelial function/damage, such as an excess of CHD risk factors, a phenomenon well described in SLE.⁶ However, these risk factors remained stable between visits. Several studies have found a similar correlation between EMP levels and endothelial function in other disease states, such as obesity,²¹ renal failure²² and heart failure.³⁶ However, SLE is an ideal model in which to examine the interaction between inflammation, endothelial dysfunction and accelerated atherosclerosis, as the cohorts are characterised by younger female patients with a chronic inflammatory disease and an excess risk of premature cardiovascular events. While FMD remains the gold standard measure of endothelial function, it is observer-dependent and difficult to standardise in multicentre studies. CD31+/-annexin V+/-CD42b− EMPs have many potential advantages over FMD in this regard and may be a useful adjunctive measure of cardiovascular risk in larger-scale studies.

A greater proportion of patients in this study had low FMD (defined as <5%) than has been observed previously. For example, in our previous study, 54.8% had low FMD, compared with 75% in this study, despite the earlier study examining older patients (48 years vs 41.5 years). This may in part relate to the much lower disease activity and damage scores observed in our previous cohort. We also noted that the proportion of controls with low FMD at 60 s was also higher than expected (43%). Previous studies have noted that many (42%) healthy subjects have a peak arterial diameter outside the traditional 60 s time point, occurring earlier in younger subjects and later in older subjects.³⁷ When we used the peak FMD response, the proportion of controls with low FMD was 33% and remained lower in patients with SLE. This would suggest that the 60 s time point might not accurately capture true FMD response in all subjects, and recording both peak and 60 s brachial artery diameters as we have done is now recommended by methodological guidelines.³⁸

We observed a significant reduction in disease activity in SLE patients who completed the study. This was associated with a
significant reduction in CD31+/annexin V+/CD42b− EMPs, with post-treatment levels comparable to those seen in healthy controls. The overall change in these indices did not correlate well with each other, in part related to the variability of the two measures in a small cohort, although the study was not powered to specifically address this issue. In our cohort, factors known to affect EMPs, such as dyslipidaemia, MetS, statins and prednisolone dose, did not change significantly over the study period. We therefore hypothesise that suppression of inflammation was the key factor mediating this change in EMPs. The overall change in disease activity (change in global BILAG-2004 score) correlated modestly with change in CD31+/annexin V+/CD42b− EMP levels ($r^2=-0.40$, $p=0.08$), although it should be noted that non-linear clinical disease activity indices are limited in their ability to be used in this way. Nevertheless, the trend to correlation and the stability of other key factors likely to affect EMPs supports this conclusion, as does the significant independent effect of active SLE on EMP levels at baseline. Active inflammatory disease is therefore likely to have a deleterious effect on the cardiovascular system, which appears to be modifiable over time. Our observation of a non-significant improvement in disease activity and endothelial function markers with rituximab therapy also suggests that better disease control may mitigate vascular risk in patients with SLE. However, controlled clinical trials are required to confirm this observation.

EMPs display paracrine and autocrine actions on vascular cells, and growing evidence suggests EMPs act as mediators in intracellular signalling because of their capacity to transfer a number of bioactive molecules to recipient cells. These bioactive molecules include growth factors, proteases, adhesion molecules, DNA and microRNAs. Functional proteins such as VEGF and endothelial nitric oxide synthase have also been identified in EMPs, and EMPs from patients with CVD have been shown to impair nitric oxide release from vascular cells. Cell culture-derived EMPs have also been shown to inhibit angiogenesis in mouse models of atherosclerosis, and EMPs may have vasculoprotective effects on the endothelium in acute vascular stress, such as septic shock. Therefore, rather than being inert markers of injury, EMPs may act as downstream delivery systems for proinflammatory products that are vasculoprotective in acute inflammatory conditions but which may perpetuate vascular dysfunction in chronic disease. Experiments are on-going in our laboratory to further examine their role in the vascular dysfunction observed in patients with SLE. There remains a lack of consensus on how best to identify and define MPs, with variation noted in the choice of cell-surface markers. In this study, we used a standard combination of markers to identify EMPs (annexin V+/CD31+/CD42b−), although we do acknowledge that other groups have used alternatives. EMPs are not specific to endothelial cells, and low-level expression is seen in platelets and monocytes. We did not include a B-cell marker in our flow-cytometry protocol and therefore cannot comment specifically on whether the marked fall in EMPs in patients receiving rituximab (an anti-CD20 therapy) was due to a fall in CD20+CD31+ B cells. However, monocots do not typically express CD20, and this scenario seems unlikely.

A key strength of this study is the prospective study design and recruitment of patients with active disease that permits exploration of change in disease activity over time and its influence on endothelial dysfunction. Cross-sectional studies of stable, older cohorts of SLE patients have limited ability to explore the interplay between inflammation and cardiovascular risk. Secondly, EMPs and FMD were measured in patients and controls simultaneously, permitting the validation of EMP levels as a biomarker of endothelial dysfunction in SLE. Finally, our cohort was younger than in comparable studies, minimising the influence of age on the outcomes. We also acknowledge several limitations. First, while the sample size was sufficient to examine the primary outcome, it did not allow full and detailed exploration of secondary outcomes, such as the effect of approach to treatment. Such a question will require adequately powered clinical trials. We did not assess an inactive SLE control group and therefore cannot comment on whether SLE itself is inherently associated with elevated EMPs, although we have previously shown that stable, inactive SLE is associated with impaired FMD. However, we have shown that EMP levels are similar to those observed in healthy controls when disease is controlled. The inherent variability in the methods used to
assess endothelial function and damage, despite optimal test conditions and techniques, also limits the exploration of secondary end points such as assessing correlation and agreement. Finally, not all patients returned for follow-up and we cannot exclude the possibility that this may have introduced some bias into our results.

We have therefore shown that CD31+/annexin V+/CD42b–EMP s are significantly elevated and FMD is significantly impaired in patients with active SLE. Improved control of inflammatory disease activity is associated with an improvement in these indices, especially EMP levels. Endothelial damage and dysfunction is therefore modifiable in patients with active SLE and raises the hypothesis that improved SLE disease control will contribute to reduced cardiovascular risk in this high-risk population.

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Contributors BP was involved in study conception and design, data acquisition, data analysis and interpretation, manuscript preparation and critical revision, and final approval of the published version. AA-H, PP and APT were involved in data acquisition and interpretation, drafting of the manuscript, and final review of the published version. PH, RG and LST were involved in data acquisition, drafting of the manuscript, and final review of the published version. MYA and INB were involved in study conception and design, data acquisition, data analysis and interpretation, manuscript preparation and critical revision, and final approval of the published version.

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Competing interests None.

Ethics approval Oldham Research Ethics Committee.

Patient consent Obtained.

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REFERENCES
Clinical and epidemiological research


Supplemental File 1: Endothelial damage and dysfunction methodology

Assessment of CD31+/AnnexinV+/CD42b- EMPs

Peripheral venous blood was collected from the study participant in a 5ml citrated vacutainer using a 21g needle. Platelet-poor plasma (PPP) was generated using a two-step centrifugation process. The venous sample was initially centrifuged at 1700g for 10 minutes at 4°C to generate plasma, which was harvested and centrifuged at 20,000g for a further 10 minutes at 4°C. The PPP was harvested and the final sample frozen in aliquots at -80°C to analyse in batches. We added 50µl of PPP and 50µl of 10µm diameter counting beads (Flow-Count™ Fluorospheres; Beckman Coulter, UK) to 900µl of calcium-rich buffer (Annexin V Apoptosis Detection Kit, eBioscience, UK). Simultaneous incubation for 10-minutes with fluorescent antibodies was performed using 5µl of phycoerythrin (PE)-conjugated antihuman CD31 (BD Pharminogen 55546), 5µl of allophycocyanin (APC)-conjugated anti-human CD42b (BD Pharminogen 551061), and 10µl efluor450 annexin-V marker (Annexin V Apoptosis Detection Kit eBioscience, UK). Flow cytometry was performed on prepared samples using the Beckman Coulter Cyan ADP flow-cytometer with Summit V4.3 software. Analysis was stopped once 1000 beads had been counted and gates were set to exclude artefact and beads. Events negative for CD42b (platelet-marker) were selected and events positive for CD31 (endothelial marker) and Annexin V (microparticle marker) were defined as CD31+/AnnexinV+/CD42b- EMPs. Absolute EMP counts per millilitre of plasma were then calculated.

Assessment of endothelial function (FMD (%))

All scans were performed on fasting subjects between 08:00 and 10:00, and vasoactive medications were withheld for 24 hours. Participants were examined in a temperature-controlled room (21-24°C), after a period of rest and equilibration. Endothelial-dependent FMD of the brachial artery was assessed using B mode ultrasound and a 12 MHz probe (Philips ATL HDI 5000) linked to a personal
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computer (PC) via a BNC cable (National Instruments). The PC was equipped with a frame grabber (National Instruments PCI 1405), vision deployment engine (National Instruments) and automated edge-tracking software (Vascular Image Analysis (VIA), MD Medics). The brachial artery was identified longitudinally, 5-10cm proximal to the ante-cubital fossa, and the probe was fixed in position using a stereotactic probe-holder. Depth and gain adjustments were made to obtain the best possible wall-lumen interface and a reference image was saved to disk, and referred to on the 2nd visit. A region-of-interest was selected and the walls of the brachial artery automatically tracked for the entirety of the test. B-mode ultrasound images were processed at 25 frames per second and the vessel diameter was displayed in real time. Baseline measures were recorded for at least 2 minutes and the BP cuff was then inflated to 50mmHg above the resting systolic blood pressure (to at least 200mmHg) for 5 minutes. Following cuff deflation, Doppler ultrasound was used to confirm reactive hyperaemia and recording continued until the arterial diameter had returned to baseline. Endothelial-independent dilatation of the brachial artery was then assessed following the administration of sublingual glycercyl trinitrate (GTN) 300mcg. Automated on-line analysis was performed immediately and average baseline brachial artery diameter (mm), average diameter at 60 seconds post-deflation (mm), and FMD (%) at 60 seconds were automatically calculated by VIA. Peak brachial artery diameter and peak FMD were also manually selected and recorded if different to the pre-set 60-second time point. Percentage FMD was calculated as follows:

% FMD = [(mean peak diameter-mean baseline diameter)/mean baseline diameter] x 100%.

In a preliminary study to validate the technique and protocol, the correlation coefficient for paired measures of baseline brachial artery diameter and FMD (%) in 8 healthy subjects at least 1 week apart was 0.95 (p = 0.0003) and 0.79 (p = 0.02), respectively.