EXTENDED REPORT

HRES-1/Rab4-mediated depletion of Drp1 impairs mitochondrial homeostasis and represents a target for treatment in SLE

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ABSTRACT

Objective Accumulation of mitochondria underlies T-cell dysfunction in systemic lupus erythematosus (SLE). Mitochondrial turnover involves endosomal traffic regulated by HRES-1/Rab4, a small GTPase that is overexpressed in lupus T cells. Therefore, we investigated whether (1) HRES-1/Rab4 impacts mitochondrial homeostasis and (2) Rab geranylgeranyl transferase inhibitor 3-PEHPC blocks mitochondrial accumulation in T cells, autoimmunity and disease development in lupus-prone mice.

Methods Mitochondria were evaluated in peripheral blood lymphocytes (PBL) of 38 SLE patients and 21 healthy controls and mouse models by flow cytometry, microscopy and western blot. MRL/pr mice were treated with 125 μg/kg 3-PEHPC or 1 mg/kg rapamycin for 10 weeks, from 4 weeks of age. Disease was monitored by antinuclear antibody (ANA) production, proteinuria, and renal histology.

Results Overexpression of HRES-1/Rab4 increased the mitochondrial mass of PBL (1.4-fold; p=0.019) and Jurkat cells (2-fold; p=0.000016) and depleted the mitophagy initiator protein Drp1 both in human (−49%; p=0.01) and mouse lymphocytes (−41%; p=0.03). Drp1 protein levels were profoundly diminished in PBL of SLE patients (−86±3%; p=0.012). T cells of 4-week-old MRL/pr mice exhibited 4.7-fold over-expression of Rab4A (p=0.0002), the murine homologue of HRES-1/Rab4, and depletion of Drp1 that preceded the accumulation of mitochondria, ANA production and nephritis. 3-PEHPC increased Drp1 (p=0.03) and reduced mitochondrial mass in T cells (p=0.02) and diminished ANA production (p=0.021), proteinuria (p=0.00004), and nephritis scores of lupus-prone mice (p<0.001).

Conclusions These data reveal a pathogenic role for HRES-1/Rab4-mediated Drp1 depletion and identify endocytic control of mitophagy as a treatment target in SLE.

INTRODUCTION

Mitochondrial dysfunction, characterised by the persistent elevation of the mitochondrial transmembrane potential (∆Ψm) or mitochondrial hyperpolarisation (MHP), oxidative stress and depletion of intracellular adenosine triphosphate (ATP) and glutathione, plays fundamental roles in abnormal T-cell activation in patients with SLE.1–3 The mammalian target of rapamycin (mTOR) serves as a sensor of mitochondrial homeostasis in T cells4 and is activated in SLE patients.5 Blockade of mTOR with rapamycin improved disease activity in murine lupus6 and in SLE patients.7 N-acetylcysteine (NAC), which serves as both a precursor of glutathione and antioxidant in and of itself, also diminished disease activity in SLE patients, through inhibiting mTOR.8 However, mTOR blockade with either rapamycin7 or NAC failed to reverse MHP and the accumulation of mitochondria,9 indicating that mitochondrial dysfunction is located upstream of mTOR activation in SLE.

The efficacy of mTOR blockade is related to the reversal of enhanced Ca2+ signalling through the T-cell receptor,7 which is attributed to the lysosomal degradation of CD4 and CD3ζ and their substitution by FcεRI7 which, in turn, are regulated by the small GTPase HRES-1/Rab4 through endosomal traffic.5 mTOR has been localised to endosomes,10 including those carrying HRES-1/Rab4.5 Polymorphic alleles of HRES-1 locus are associated with SLE11,12 and influence expression of HRES-1/Rab4,13 which may represent the gene product that confers susceptibility to lupus at 1q42.14 Although mTOR has been widely implicated in the suppression of autophagy,15,16 the opposing effects of rapamycin on the degradation of proteins, such as CD4 and CD3ζ, and organelles, such as mitochondria, both of which occur in lysosomes, appear paradoxical. During autophagy, proteins and organelles alike are carried to the lysosome as endosomal cargo.17 Therefore, we examined the role of HRES-1/Rab4, which is over-expressed in T cells of SLE patients,15 as a selective regulator of autophagy pathways, that is, facilitator of protein microautophagy and inhibitor of organellar macroautophagy.

Autophagy is a fundamental stress-induced catabolic process that moves cargo to the lysosome through three main mechanisms: (1) microautophagy, transferring cytoplasmic materials into the lysosome through endosome fusion, (2) chaperone-mediated autophagy that requires binding of a chaperone first to its cytotoxic protein target and second to a receptor on the lysosomal membrane followed by translocation of the cytotoxic protein...
into the lysosome and (3) macroautophagy, characterised by the formation of a cytosolic double-membrane vesicle primarily involving organelles such as mitochondria.18 Macroautophagy of mitochondria or mitophagy is initiated by the translocation of dynamin-related protein 1 (Drp1) from the cytosol to mitochondria, which causes the fission, fragmentation and lysosomal degradation of mitochondria.17 19 Here, we reveal that HRES-1/Rab4 promotes the lysosomal degradation of Drp1 in Jurkat human T cells. The overexpression of HRES-1/Rab4 increased mitochondrial mass in Jurkat cells and PBL. Drp1 was reduced in lupus T cells and was unaffected by mTOR blockade, similar to the persistence of mitochondrial accumulation in rapamycin-treated patients. Moreover, expression of the murine homologue of HRES-1/Rab4, Rab4A, was found to be increased, while Drp1 was found to be decreased in the spleen and thymus of lupus-prone NZB/W F1 and MRL/lpr mice relative to C57BL/6 controls at 4 weeks of age well before the onset of ANA production, proteinuria and glomerulonephritis (GN). Treatment with Rab geranylgeranyl transferase inhibitor 3-PEHPC, which inactivated HRES-1/Rab4, reversed the depletion of Drp1 as well as the accumulation of mitochondria, and prevented ANA production and nephritis in MRL/lpr mice.

**MATERIALS AND METHODS**

**Human subjects**

Thirty-eight patients with SLE were investigated; each patient satisfied the criteria for a definitive diagnosis.20 21 As controls, 21 healthy subjects were matched for gender, ethnicity and age within 10 years of SLE patients for each blood donation and studied in parallel. Additional details are provided in the online supplementary methods section.

**Separation and culture of human PBL**

Peripheral blood mononuclear cells (PBMC) were isolated from heparinised venous blood on Ficoll–Hypaque gradient. Peripheral blood lymphocytes (PBL) were separated from monocytes by adherence to autologous serum-coated Petri dishes.22 T cells (>95% CD3) were negatively isolated from PBMC with Dynal magnetic beads conjugated to IgG antibodies for CD14, CD16 human leucocyte antigen (HLA) class II DR/DP/CDS6, CD235a; Invitrogen Cat No 113-11D). CD4 T cells (>98% CD4) were negatively isolated with magnetic beads conjugated to IgG antibodies for CD8, CD14, HLA class II DR/DP/CDS6, CDw123 and CD235a (Invitrogen Cat No 113-39D). The resultant cell population was resuspended at 10⁶ cells/mL in RPMI 1640 medium, supplemented with 10% fetal calf serum (FCS), 2 mM L-glutamine, 100 μg/mL penicillin, and 100 μg/mL streptomycin, 10 μg/mL amphotericin B, 1% MEM non-essential amino acid solution (10 mM, Invitrogen), and 1 mM sodium pyruvate.23

**Flow cytometric analysis of ΔΨ_m mitochondrial mass and nitric oxide**

Flow cytometry was performed on freshly isolated cells, as described in the online supplementary methods section.

**Microarray analysis of gene expression**

Biotinylated cRNA was produced by in vitro transcription and hybridised to Affymetrix HG-U133 Plus-2 chips, as described in the online supplementary methods section.

**Western blot analysis**

Protein lysates were separated by SDS-PAGE, transferred to nitrocellulose, probed with antibodies, and analysed, as described in the online supplementary methods section.

**Transfection of siRNA**

Knock-down of HRES-1/Rab4 via siRNA transfection is described in the online supplementary methods section.

**Transduction of HRES-1/Rab4 by adeno-associated virus**

HRES-1/Rab4 cDNA was cloned upstream of the internal ribosomal entry site (IRES) in pAAV-IRESGFP vector, as described in the online supplementary methods section.

**Confocal microscopy**

Lysosomal compartments were labelled LysoTracker Red (LTR). Mitochondria were visualised by staining with MitoTracker Deep Red (MTDR). Additional details are provided in the online supplementary methods section.

**In vitro prenylation assay**

Prenylation of HRES-1/Rab4 and Rab5 was evaluated by western blot of soluble and insoluble fractions from Triton×114-treated Jurkat cells, as described in the online supplementary methods section.

**Autophagy induction**

Autophagy was induced with 50 nM rapamycin in the presence and absence of lysosomal inhibitor, 200 nM bafilomycin A1, as described in the online supplementary methods section.

**In vitro prenylation assay**

Prenylation of HRES-1/Rab4 and Rab5 was evaluated by western blot of soluble and insoluble fractions from Triton×114-treated Jurkat cells, as described in the online supplementary methods section.

**Treatment of MRL/lpr mice with Rab geranylgeranyl transferase inhibitor 3-PEHPC and rapamycin**

4-week-old MRL/lpr mice were separated into four treatment groups receiving phosphate-buffered saline (PBS) (solvent control for 3-PEHPC; n=4), 125 μg/kg 3-PEHPC in PBS (n=8), 0.2% carboxymethylcellulose (CMC, solvent control for rapamycin, n=4), or 1 mg/kg rapamycin in CMC (n=8). Proteinuria was monitored every 2 weeks between 4–14 weeks of age. To assess treatment toxicity, mice were weighed weekly. Renal pathology was assessed by scoring for glomerulonephritis (GN), glomerulosclerosis (GS), and interstitial nephritis (IN) on a 0–4 scale, and percentage of sclerotic and crescentric glomeruli, as described in the online supplementary methods section.

**ANA and cytokine assays**

ANA, IFN-γ, IL-10 and IL-17A were measured by ELISA, as described in the online supplementary methods section.

Statistical analyses

Statistical analyses were performed using Statview V.5.0.1 (SAS Institute, Cary, North Carolina) and GraphPad Prism V.5.0 Software (San Diego, California). Additional details are provided in the online supplementary methods section.

Supplementary materials include supplemental methods and online supplementary figures S1–S7.

RESULTS

HRES-1/Rab4 promotes the lysosomal degradation of Drp1

To assess the impact of HRES-1/Rab4 on autophagy pathways, we evaluated global gene expression in Jurkat cells carrying doxycycline-inducible GFP-producing, HRES-1/Rab4 and GFP-producing, or dominant-negative HRES-1/Rab4S27N (HRES-1/Rab4-DN) and GFP-producing expression vector following incubation with or without doxycycline for 24 h. 116 genes exhibited doxycycline-induced reciprocal changes of expression in response to HRES-1/Rab4 relative to HRES-1/Rab4-DN at 99.99% CIs (data not shown). Pathway analysis on the basis of protein–protein interaction with Strings 9.0 software (http://string-db.org) indicated a predominant influence of HRES-1/Rab4 on genes involved in mitochondrial outer membrane organisation (false discovery rate p value with Bonferroni correction (FRD p=1.8×10^{-9}), membrane raft formation (p=7.5×10^{-9}), and mTOR activation (p=9.7×10^{-3}). The predictive protein interactome analysis implicated genes in addition to the ones revealed by the microarray studies, such as Drp1 and AKAP10. Drp1 initiates the fission and autophagy of mitochondria. In turn, Drp1 is phosphorylated by protein kinase A (PKA) that associates with the endosomal membrane via the PKA anchoring protein AKAP10. As mitochondrial turnover involves endosomal traffic regulated by HRES-1/Rab4, a small GTPase that is overexpressed in lupus T cells, it was important to evaluate the impact of HRES-1/Rab4 on Drp1 and AKAP10.

The functional consequences of HRES-1/Rab4 overexpression on Drp1 were initially evaluated in Jurkat human T cells. As shown in figure 1A, the over-expression of HRES-1/Rab4 reduced Drp1 protein levels by 49% (p=0.01). The phosphorylation of Drp1 at Ser 637 was also reduced by HRES-1/Rab4 (66% decrease, p=0.005, figure 1A). As Drp1 is phosphorylated at Ser 637 by PKA, which is anchored to endosomes via AKAP10, we also evaluated the effect of HRES-1/Rab4 on AKAP10. As shown in figure 1A, AKAP10 protein levels were reduced by HRES-1/Rab4-DN. Both the HRES-1/Rab4-induced depletion of Drp1 and the HRES-1/Rab4-DN-induced depletion of AKAP10 were strikingly reversed in bafilomycin-treated Jurkat cells (figure 1A), suggesting a role for increased lysosomal degradation. Along these lines, the knockdown of HRES-1/Rab4 markedly upregulated Drp1 protein levels in HeLa cells which were amenable to transfection by siRNA without loss of viability (figure 1B). As expected, the deletion of Drp1 increased mitochondrial mass in MEFs (figure 1C) which was consistent with a role for Drp1 in promoting mitophagy. In the absence

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**Figure 1** Depletion of Drp1 by HRES-1/Rab4. (A) Effect of HRES-1/Rab4 on Drp1 protein levels, phosphorylation of Drp1 at Ser 637, and levels of AKAP10, which regulates phosphorylation of Drp1, were evaluated in Jurkat human T cells carrying doxycycline-inducible vectors that express wild-type HRES-1/Rab4 or dominant-negative HRES-1/Rab4S27N (HRES-1/Rab4-DN). Bafilomycin reversed the depletion of Drp1 by HRES-1/Rab4 and depletion of AKAP10 by HRES-1/Rab4-DN. Left panel, representative western blots; right panel, mean±SEM of 4–6 experiments. (B) Upregulation of Drp1 protein levels by siRNA-mediated knock-down of HRES-1/Rab4. Left panel, representative western blots; right panel, mean±SEM of 4–6 experiments. (C) Effect of Drp1 deletion in MEFs on Rab4 protein levels assessed by western blot and on mitochondrial mass measured by MTG fluorescence. Representative western blot (left panel) and bar chart reflect mean±SEM of 4 experiments (right panel).
of Drp1, the expression of Rab4A, the mouse homologue of HRES-1/Rab4, was diminished (figure 1C).

**HRES-1/Rab4 induced the accumulation of mitochondria by inhibiting mitophagy**

Next, we investigated if the HRES-1/Rab4-mediated depletion of Drp1 elicited changes in mitochondrial mass. Jurkat cells overexpressing HRES-1/Rab4 exhibited twofold greater mitochondrial mass by MitoTracker Deep Red (MTDR) staining ($p=1.6 \times 10^{-5}$; see online supplementary figure S1A and B). Accumulation of mitochondria was also indicated by 1.7-fold elevated voltage-dependent anion channel (VDAC) protein levels in Jurkat cells overexpressing HRES-1/Rab4 ($p=0.03$; data not shown). To investigate the role of autophagy in the effect of HRES-1/Rab4 on mitochondria, we examined their association with lysosomes. The colocalisation of mitochondria with lysosomes was diminished in cells overexpressing HRES-1/Rab4 (see online supplementary figure S1). Mitochondrial mass was also increased in PBL overexpressing HRES-1/Rab4 (1.4-fold; $p=0.019$; figure 2A). Consistent with its impact on mitochondrial homeostasis, HRES-1/Rab4 was enriched in mitochondria relative to the cytosol both in Jurkat cells (not shown) and PBL (figure 2B). The partitioning of HRES-1/Rab4 to mitochondria was markedly reduced in lupus T cells relative to matched healthy controls ($-52 \pm 12\%$, $p=0.005$; figure 2B). The diminished localisation of mitochondria to lysosomes upon HRES-1/Rab4 overexpression in Jurkat cells, the diminished association of HRES-1/Rab4 with mitochondria in lupus PBL, and the accumulation of mitochondrial mass in Jurkat cells and PBL overexpressing HRES-1/Rab4 together suggested an underlying mechanism of inhibited mitophagy.

**Stimulation of microautophagy by HRES-1/Rab4**

Since early endosomes are involved in the formation of autophagosomes, we examined the effect of HRES-1/Rab4 on lipidation of the microtubule-associated light chain 3 (LC3) with phosphoethanolamine, an early event of autophagosome formation. While the lipidated LC3 (LC3-II) is associated with the membrane of autophagosomes, the unlipidated form of LC3 (LC3-I) resides in the cytosol. These studies were performed...
autophagy is required for T-cell activation, we investigated the reversed by rapamycin treatment (p=0.01, figure 3). Reduced in freshly isolated SLE PBL (p=8.8x10^{-5}), the expression of mitochondria in lupus T cells were associated with healthy controls (p=0.012). Diminished Drp1 levels were reduced by 86±3% in lupus PBL in comparison with matched controls (p=0.012). Diminished Drp1 levels were not corrected in SLE patients treated with rapamycin in vivo (−83±7%, p=0.017; figure 3), which was consistent with the enduring accumulation of mitochondria in lupus T cells treated with rapamycin (p<0.01, figure 3). Since Drp1 is a key player in the growth of mitochondria, we investigated the capacity of lupus T cells to upregulate the expression of LC3 II in response to CD3/CD28 costimulation. Indeed, the accumulation of LC3-II was 1.9-fold increased in lupus patients in comparison with healthy controls (p=8.8x10^{-5}). As expected, the LC3-II/LC3-I ratio was further enhanced by rapamycin (figure 3). In accordance with earlier findings, rapamycin treatment reduced mTOR activity but did not reverse the accumulation of mitochondria in T cells (data not shown). These findings suggested that the depletion of Drp1 (figure 3) along with the accumulation of mitochondria is independent of mTOR activation in SLE.

Drp1 expression is reduced and resistant to rapamycin treatment in SLE

Since HRES-1/Rab4 depleted the mitophagy initiator Drp1, we investigated if overexpression of HRES-1/Rab4 and accumulation of mitochondria in lupus T cells were associated with altered expression of Drp1. As shown in figure 3, Drp1 protein levels were reduced by 86±3% in lupus PBL in comparison with matched controls (p=0.012). Diminished Drp1 levels were not corrected in SLE patients treated with rapamycin in vivo (−83±7%, p=0.017; figure 3), which was consistent with the enduring accumulation of mitochondria in lupus T cells treated with rapamycin (p<0.01, figure 3). Since autophagy is required for T-cell activation, we investigated the capacity of lupus T cells to upregulate the expression of LC3 II in response to CD3/CD28 costimulation. Indeed, the accumulation of LC3-II was 1.9-fold increased in lupus patients in comparison with healthy controls (p=8.8x10^{-5}). As expected, the LC3-II/LC3-I ratio was further enhanced by rapamycin (figure 3). In accordance with earlier findings, rapamycin treatment reduced mTOR activity but did not reverse the accumulation of mitochondria in T cells (data not shown). These findings suggested that the depletion of Drp1 (figure 3) along with the accumulation of mitochondria is independent of mTOR activation in SLE.

Increased expression of Rab4A and depletion of Drp1 precede accumulation of mitochondria and disease development in lupus-prone mice

To assess the involvement of Rab4A in lupus pathogenesis, we examined its expression in lupus-prone mice relative to disease development. Disease-free mice were studied at 4 weeks of age, when we confirmed the absence of ANA, proteinuria, or nephritis. Rab4A expression was markedly increased (4.7-fold; p=0.0002; figure 4A) while Drp1 was diminished in T cells of 4-week-old female MRL/lpr mice (−54%, p=0.02; figure 4B). Expression of Rab4A was further increased at 8 weeks of age (36-fold; p=0.0005; figure 4C) when accumulation of mitochondria was evidenced by western blot detection of VDAC and MTG fluorescence in MRL/lpr mice (figure 4D). Rab4A and Drp1 protein levels were similar in naïve CD44lowCD62LhighCD4 T cells of B6/Lpr, MRL, and MRL/lpr mice relative to B6 controls (data not shown). Expression of Rab5, evaluated as a control Rab GTPase that mediates endocytic internalisation, was not altered in MRL/lpr, MRL, or B6/lpr relative to B6 mice (figure 4C).

At 4 weeks of age, we also observed a robust overexpression of Rab4A and depletion of Drp1 in splenocytes of female NZB/W F1 mice (figure 4E). In support of a mechanistic relationship between reciprocal changes in Rab4A and Drp1, overexpression of HRES-1/Rab4 reduced Drp1 protein levels in B6 splenocytes by −41±7.8% (p=0.034; figure 4F).

Rab4A expression was also increased in thymocytes of 4-week-old NZB/W F1 mice (see online supplementary figure S3A), showing no evidence of ANA production, proteinuria, or nephritis (data not shown). NZB/W F1 mice were also studied at 4 months and 6 months of age, when autoimmunity was detected by the production of ANA, and at 11 months of age when all mice developed nephritis of varying severity. At 4 months of age, Rab4A expression was further increased in splenocytes of NZB/W F1 mice, over 15-fold relative to age-matched B6 controls. To a lesser extent, Rab4A was overexpressed in splenocytes of 4-month-old B6.NZMSle1.Sle2.Sle3 mice (see online supplementary figure S3A). Rab5A was unchanged in splenocytes at 4 weeks of age, but it was overexpressed at 4 months of age in NZB/W F1 mice (p=0.02; see online supplementary figure S3B).

Since HRES-1/Rab4 targets CD3ζ and CD4 for lysosomal degradation, we examined this process in NZB/W F1 mice. CD3ζ expression was reduced, while CD4 expression was unchanged in T cells of 4-week-old NZB/W F1 mice in...
Figure 4  Increased expression of Rab4A and depletion of Drp1 in MRL/lpr and NZB/W F1 mice. (A) Western blot analysis of Rab4A and Rab5 expression in thymocytes from 4-week-old C57BL/6 (B6), MRL, C57BL/6/Lpr (B6/Lpr), and MRL/lpr mice. Representative blots are shown in the left panel, while cumulative analyses are shown in the right panel. p Values <0.05 are indicated. (B) Western blot analysis of Drp1 in negatively isolated T cells of 4-week-old mice. (C) Western blot analysis of Rab4A and Rab5 expression in splenocytes from 8-week-old mice. (D) Western blot detection of VDAC and flow cytometry of mitochondrial mass using MTG fluorescence in splenocytes of 8 week-old mice. For each parameter 4 or more mice were analysed; p values <0.05 are indicated. (E) Western blot analysis of Drp1 protein levels in B6 mouse splenocytes transduced with adeno-associated virus (AAV) expressing HRES-1/Rab4 (left panel). Data represent mean±SEM of five experiments; p values reflect comparison to cells infected with control AAV using paired two-tailed t test (right panel).
comparison with B6 controls (see online supplementary figure S3C). At 4 months of age, CD3ζ was further depleted and CD4 expression also became diminished (see online supplementary figure S3C). Beclin-1 expression was reduced in T cells from NZB/W F1 and NZW mice (see online supplementary figure S3D). Drp1 protein levels were also diminished in thymocytes of NZB/W F1 mice relative to B6 controls (see online supplementary figure S3D). Increased nitric oxide (NO) production was detectable in 4-week-old NZB/W F1 mice (see online supplementary figure S3E).

mTOR activity was not consistently elevated at 4 weeks of age but it was robustly increased in 4-month-old NZB/W F1 mice (see online supplementary figure S4). Disease progression was associated with the escalation of Rab4A overexpression, (4.3-fold at 4 weeks of age and 15.3-fold at 4 months of age) and activation of mTOR (0.8-fold at 4 weeks of age and 3.3-fold at 4 months of age; p=0.028 using 2-way analysis of variance) in the spleen of NZB/W F1 female mice with respect to age-matched B6 female controls (see online supplementary figure S4). A progressive decrease of mitochondrial mass was observed in B6 mice with aging (see online supplementary figure S5A). By contrast, NO production, MHP and accumulation of mitochondria, as detected by increased MTG fluorescence (see online supplementary figure S5B) and VDAC expression (see online supplementary figure S5C), were peaking in 11-month-old NZB/W F1 mice. Thus, lupus-prone mice exhibit overexpression of Rab4A, depletion of Drp1, accumulation of mitochondria, and mTOR activation, similar to patients with SLE.5, 16

Rab geranylgeranyl transferase blockade reverses Drp1 depletion, mitochondrial accumulation, ANA production, and nephritis in lupus-prone mice

Increased Rab4A and depleted Drp1 were thus observed in 4-week-old disease-free MRL MRL/lpr and NZB/W F1 mice, indicating that these changes may represent early events in lupus pathogenesis. The accumulation of mitochondria was noted in 8-week-old MRL/lpr and 11-month-old NZB/W F1 mice. This was consistent with earlier development of autoimmunity in MRL/lpr mice.28 To evaluate Rab4 and the pathway of endocytic recycling as target for treatment of SLE, we used a Rab geranylgeranyl transferase inhibitor 2-[3-pyridinyl]-1-hydroxyethylidene-1,1-phosphonocarboxylic acid (3-PEHPC)30 that promoted the partitioning of HRES-1/Rab4 and Rab5 from the membrane to the cytosol compartment (see online supplementary figure S6). MRL/lpr mice were treated from 4 to 14 weeks of age, three times weekly, with subcutaneous injection of 125 μg/kg 3-PEHPC, previously found to be effective in diminishing bone resorption.30 We compared the efficacy of 3-PEHPC to rapamycin, a potent inhibitor of lupus disease through mitophagy,17 19 the depletion of Drp1 by HRES-1/Rab4 leads to increased mitochondrial mass in Jurkat cells and primary human peripheral blood T cells. The paucity of Drp1 prevents mitochondrial fission and promotes unopposed mitochondrial fusion15 which is consistent with the formation of megamitochondria in lupus T cells.26 The importance of mitochondrial homeostasis in lupus pathogenesis has also been supported by the recent identification of an NZB/W F1-derived lupus susceptibility gene as oestrogen-related receptor gamma, which accounts for increased mitochondrial mass that is detectable by elevated VDAC protein levels in the spleen of Sle1c2 mice.16

Importantly, Rab4A, but not Rab5, was robustly overexpressed in the spleen and thymus of lupus-prone mice. The activation of Rab4A and depletion of Drp1 were detected at age of 4 weeks, well before the onset of ANA production and nephritis in NZB/W F1 or MRL/lpr mice. The overexpression of HRES-1/Rab4 led to the depletion of Drp1 both in human and murine lymphocytes, supporting a causal relationship between reciprocal changes in Rab4A and Drp1 both in lymphocytes of SLE patients and lupus-prone mice. The inactivation of Drp1 reduced Rab4A expression and increased mitochondrial mass in MEFs, which was compatible with the molecular order that overexpression of HRES-1/Rab4 and increased endocytic recycling were upstream of Drp1 depletion and mitochondrial accumulation in SLE.

Rab5 overexpression and CD4 depletion were detectable in the spleen of NZB/W F1 mice at 4 months of age, suggesting a step-wise activation of endosome recycling with progression of...
disease pathogenesis and mimicking the reciprocal overexpression of small GTPases HRES-1/Rab4 and Rab5 and depletion of CD4 and CD3ζ in T cells of SLE patients. We found that expression levels of Rab4A and Drp1 in naïve CD4 T cells of B6 mice were not different relative to those of B6/Lpr, MRL and MRL/lpr mice (data not shown). CD3/CD28 costimulation for 24 h reduced Drp1 levels in human PBL by 80.8±9.4% (p=0.007; data not shown). As earlier documented, CD3/CD28 costimulation increased HRES-1/Rab4 protein levels in human T cells. These findings indicate that Rab4A overexpression and Drp1 depletion may depend on T-cell activation in SLE.

Increased production of NO during T-cell activation which induces the expression of HRES-1/Rab4 in vitro, may underlie the overexpression of Rab4A in 4-week-old NZB/W F1 mice. Lupus-associated polymorphic haplotypes within the long terminal repeat-enhancer also influence expression of HRES-1/Rab4 in human T cells.

Blocking of Rab geranylgeranyl transferase activity with 3-PEHPC prevented Drp1 depletion, accumulation of mitochondria in T and B cells, ANA production and nephritis, indicating that activation of the endocytic recycling machinery contributes to the pathogenesis of SLE. The reversal Drp1 depletion by 3-PEHPC may influence T-cell activation through positioning of mitochondria at the immunological synapse. Mitochondria serve as a buffer for the management of T-cell activation-induced Ca2+ fluxing, and Drp1-dependent fragmentation of the mitochondrial network may protect T cells from hyperactive Ca2+ signalling. Interestingly, rapamycin failed to reverse Drp1 depletion or to reduce mitochondrial mass, suggesting that the accumulation of mitochondria is mTOR-independent. These findings are consistent with the persistence of MHP and accumulation of mitochondria in T cells.

Figure 5 Effect of 3-PEHPC and rapamycin (Rapa) relative to solvent controls PBS and CMC, respectively, on autoimmunity and nephritis in MRL/lpr mice treated three times weekly, between ages of 4–14 weeks. (A) Body weight, ANA production, and proteinuria. (B) Renal pathology of 14-week-old mice. Upper and lower panels represent 200-fold and 400-fold original magnifications of identical sections; size bars represent 50 microns. PBS: GN is characterised by circumferential, predominantly cellular crescents with segmental sclerosis and hyalinosis. There are prominent interstitial lymphocytic infiltrates, tubular atrophy and proteinaceous casts. 3-PEHPC: glomeruli are free of crescents. There is minimal tubular atrophy and interstitial fibrosis. CMC: glomeruli show cellular or fibrocellular crescents. Glomerular tufts are hypercellular with lobular accentuation. There is tubular atrophy and interstitial lymphocytic infiltrates and fibrosis. Rapa: normocellular glomeruli; minimal tubular atrophy and interstitial fibrosis. (C) Renal disease scores in 14-week-old MRL/lpr mice. Left panel, for each kidney, the severity of GN, GS, and IN was graded in a 0–4 semiquantitative scale. Right panel, numbers of glomeruli with sclerosis and/or crescent formation were determined and expressed as a percentage of total glomeruli observed in the entire cortical field. p values <0.05 reflect two-way analysis of variance.
cells of SLE patients upon mTOR blockade with rapamycin or NAC. In addition to the distinct impact on mitochondrial homeostasis, the cytokine profile of MRL/lpr mice was also differentially influenced by 3-PEHPC and rapamycin. The production of IFN-γ and IL-10, but not of IL-10, which have been widely implicated in pathogenesis of human lupus. The diminished production of IL-10 may be related to the impact of 3-PEHPC on non-T cells, such as B cells and macrophages. Although 3-PEHPC failed to influence IFN-γ and IL17A, but reduced levels of IL-17A that have been widely implicated in pathogenesis of human lupus. The diminished production of IL-10 may be related to the impact of 3-PEHPC on non-T cells, such as B cells and macrophages. Although 3-PEHPC failed to influence IFN-γ and IL17A, but reduced levels of IL-10, which have been widely implicated in pathogenesis of human lupus.

In summary, our findings provide evidence for a role of HRES-1/Rab4-mediated Drp1 depletion in altered mitochondrial homeostasis in both SLE patients and lupus-prone mice. Rab geranylgeranyl transferase blockade with 3-PEHPC reversed the depletion of Drp1, accumulation of mitochondria, ANA production and nephritis in lupus-prone mice, identifying the regulation of mitochondrial homeostasis by HRES-1/Rab4 as a mechanism of pathogenesis and potential target for treatment in SLE.

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REFERENCES


