

A4.9 HOW OSTEOBLAST REGULATES ENERGY METABOLISM AND SYSTEMIC INFLAMMATION DEPENDENT OF FRA-2 EXPRESSION

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Background and Objectives The transcription factor Fra-2 (Fosl2) is a member of the AP-1 complex and an important regulator of bone homeostasis. We have previously shown that Fra-2 controls bone development, osteoclast size [1] and osteoblast differentiation through direct regulation of Collagen 1a2 and Osteocalcin (Ocn) [2]. Recent studies have established that the skeleton functions as an endocrine organ affecting metabolism through Ocn [3], although only few transcription factors and only one osteoblast-derived hormone are known to affect the crosstalk between bone and metabolism.

Materials and Methods We have generated mice with specific deletion of Fra-2 (*Fosl2*) or ectopic expression of Fra-2 in osteoblast to study the role of Fra-2 beyond the bone e.g. in metabolism.

Results Here we show that mice with osteoblast specific deletion of Fra-2 (*Fosl2*) have despite a low bone mass, an increased body weight. In contrast, ectopic expression of Fra-2 in osteoblasts display increased bone mass and decreased body weight accompanied with reduced serum glucose and insulin levels, improved glucose tolerance and insulin sensitivity. In addition, these Fra-2 mutant mice are protected from metabolic impairment, when challenged with high fat diet (HFD). Surprisingly a systemic inflammation and macrophage infiltration in liver, spleen and lung was observed in Fra-2 osteoblast specific mice. Mechanistically, we showed that in osteoblasts Fra-2 transcriptionally represses an important adipocytokine Adiponectin (Adipoq), while it induces Ocn, both responsible for the glucose and insulin metabolism alteration. Whereas, the systemic inflammation was likely due to the transcriptional increased of Osteopontin (OPN) expression by Fra-2, which is known as a potent inducer of macrophage activation.

Conclusions Taking together these results show that Fra-2 expression in osteoblast transcriptionally modulates Adipoq, Ocn and OPN expression and secretion representing a novel mechanism for the endocrine function of the skeleton on systemic metabolism and inflammation.

References

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A4.10 HYDROGEN SULFIDE ATTENUATES STORE-OPERATED Ca²⁺ ENTRY IN ENDOTHELIAL AND SMOOTH MUSCLE CELLS

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Background and Objectives Endothelial cells are active participants in inflammatory processes. They are involved in diverse activities including the regulation of leucocyte extravasation, angiogenesis, cytokine production, protease and extracellular matrix synthesis, vasodilation, etc. The small gaseous molecule hydrogen sulphide (H₂S) is involved in a variety of physiological processes like vascular relaxation, angiogenesis, neurotransmission and inflammation. In the vascular system, ATP-sensitive K⁺-channels are a major target for H₂S but over the last few years evidence has accumulated that several Na⁺- and Ca²⁺-permeable channels are also sensitive to H₂S. In the present study we investigated the effect of H₂S on Ca²⁺ signalling in cultured endothelial and smooth muscle cells with special emphasis given to the role of H₂S in modulating store-operated Ca²⁺ channels.

Materials and Methods Experiments were performed with human microvascular endothelial cells (HMEC-1), endothelial cells isolated from porcine aorta, and smooth muscle cells isolated from rat aorta and rat trachea. Mobilisation of intracellular Ca²⁺ and Ca²⁺ entry was monitored by measuring the intracellular free Ca²⁺ concentration with FURA-2 in the absence and presence extracellular Ca²⁺, respectively. Activity of endothelial nitric oxide synthase (eNOS) in intact cells was determined as conversion of incorporated L-[³H]-arginine into L-[³H]-citrulline.

Results Incubation of human and porcine endothelial cells with the H₂S-donor NaHS (100 μM, 10–45 min) evoked a release of Ca²⁺ from intracellular stores that was not accompanied by Ca²⁺ influx from the extracellular space. In accordance with these data suggesting that H₂S may inhibit store-operated Ca²⁺ entry, incubation of cells with NaHS attenuated Ca²⁺ influx induced by depletion of Ca²⁺ stores with receptor agonists (ATP, histamine) or the endoplasmic reticulum ATPase inhibitor, thapsigargin. As a consequence, the stimulatory effect of these agonists on endothelial NO formation was strongly diminished, whereas the response to the Ca²⁺ ionophore A23187 was barely affected. Similar to the results obtained with endothelial cells, depletion of intracellular Ca²⁺ stores in smooth cells isolated from rat aorta or rat trachea also resulted in a pronounced Ca²⁺ entry that was completely blocked upon pre-treatment of cells with NaHS.

Conclusions H₂S inhibits the stimulatory effect of Ca²⁺ of mobilising agonists on endothelial NO formation by attenuating store-operated Ca²⁺ entry. Inhibition of store-operated Ca²⁺ channels by H₂S is not peculiarity of endothelial cells but also occurs in vascular and tracheal smooth muscle cells. These hitherto undescribed effects may be in part possible for the beneficial effects of H₂S in sulphur bath therapy.

A4.11 BASELINE ELEVATED SERUM LEVELS OF CALPROTECTIN AS INDEPENDENT MARKER FOR RADIOGRAPHIC SPINAL PROGRESSION IN ANKYLOSING SPONDYLITIS

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Background and Objective Syndesmophytes formation and complete fusion of the total spine are common characteristics leading to functional impairment and disability in ankylosing spondylitis (AS) patients. Predictors for progression of structural damage are smoking, elevated levels of acute phase reactants and the presence of syndesmophytes at baseline. These predictors identify increased risk for progression at group level but their specificity is not strong enough to be used as biomarkers in individual patients. We recently