Susceptibility to ankylosing spondylitis: no evidence for the involvement of transforming growth factor β1 (TGFB1) gene polymorphisms


Background: Genetic factors are thought to be crucial in the pathogenesis of ankylosing spondylitis. Transforming growth factor β1 (TGFB1) is a multifunctional cytokine that plays a key role in inflammation. Two functional single nucleotide polymorphisms (SNPs) in the TGFB1 gene have been described: TGFB1 T869C and TGFB1 G915C.

Objective: To determine whether these SNPs contribute to ankylosing spondylitis susceptibility or its disease characteristics.

Methods: Genomic DNA was isolated from the peripheral blood of 134 patients with ankylosing spondylitis and 194 healthy blood donors. All subjects were unrelated and of white Dutch ethnicity. The diagnosis of ankylosing spondylitis was made according to the modified New York criteria. The TGFB1 T869C and TGFB1 G915C SNPs were genotyped by a polymerase chain reaction–single strand conformation polymorphism haplotyping method.

Results: No significant differences were found between patients and controls in genotype, allele, and haplotype frequencies or in the carrier rate of the rare alleles of the TGFB1 T869C and TGFB1 G915C SNPs.

Conclusions: TGFB1 T869C and TGFB1 G915C SNPs are not major factors in the susceptibility to ankylosing spondylitis or its disease characteristics.

Ankylosing spondylitis (AS) is a common familial rheumatic disorder. The disease is characterised by chronic inflammation of the sacroiliac joints and the vertebral column. The inflammatory process leads gradually to ossification and calcification of the spine and sacroiliac joints. Ultimately, ossification, interosseous bridging, and ankylosis can occur. Spinal osteoporosis is often observed in patients with active disease and is probably induced by regulatory cytokines. 

The precise aetiology of the disorder remains unclear, though heritability plays a major role. Twin studies suggest that up to 97% of the susceptibility to AS can be attributed to genetic factors. For over 30 years the main known genetic component has been human leucocyte antigen (HLA) B27. More than 95% of white patients with primary AS and at least 50% of those with AS associated with psoriasis or inflammatory bowel disease are HLA-B27 positive, whereas HLA-B27 is present in only 8% of the general Dutch population.

However, family studies have shown that the major histocompatibility complex (MHC), including HLA-B27, contributes less than 40% to the recurrence risk ratio in AS. Apart from the MHC, Laval et al identified six other loci—on chromosomes 1p, 2q, 9q, 10q, 16q, and 19q—that are linked to AS. The peak of linkage on chromosome 19 was observed with the marker D19S420 (LOD score 3.58). The TGFB1 gene encoding the human transforming growth factor β1 (TGFB1) is located 1.8 Mb from marker D19S420 on chromosome 19q13. TGFB1 is a multifunctional cytokine involved in the regulation and proliferation of cells. This cytokine promotes differentiation of leucocytes, but has inhibitory effects on proliferation of T lymphocytes and activation of macrophages, suggesting a regulatory role in inflammatory states. TGFB1 is known to be able to control the production of many components of the extracellular matrix and to be a potent regulator of osteoblast proliferation and differentiation. In general, TGFB1 stimulates cartilage and bone formation, but the precise effects are complex, depending on the concentration of TGFB, whether the bone is growing or damaged, the state of activation of the cells, and the presence of other factors, including hormones. Moreover, TGFB1 is required to switch B cells into production of IgA, which is often increased in AS patients. Therefore, TGFB1 is a good candidate for a key cytokine in an inflammatory disease such as AS, in which new bone formation and ultimately ankylosis of the sacroiliac joints and the vertebral column are major symptoms and fibrosis is a related phenomenon.

Recently, the single nucleotide polymorphisms (SNPs) at positions +869 (T→C) and +915 (G→C) in the signal protein sequence of the TGFB1 gene, which change codon 10 (Leu10Pro) and codon 25 (Arg25Pro), respectively, have been reported to be related to variations in the production of TGFB1, both in vitro and at the serum level. In the present study we investigated whether the TGFB1 T869C and TGFB1 G915C SNPs or their haplotypes are associated with susceptibility to or disease characteristics of AS.

METHODS

Subjects

After informed consent, 134 AS patients were recruited from the outpatient department of rheumatology of the Jan van Groenesteynziekenhuis. A total of 194 healthy Caucasian blood donors were recruited from the population. Subjects and controls were unrelated and of white Dutch ethnicity.

Abbreviations: HLA, human leucocyte antigen; HWE, Hardy–Weinberg equilibrium; MHC, major histocompatibility complex; SNP, single nucleotide polymorphism; TGFB1, transforming growth factor β1
Bremmen Institute. They all fulfilled the diagnostic requirements of AS according to the modified New York criteria. Controls were 194 randomly selected healthy blood donors from the Amsterdam region. All subjects were unrelated and of white Dutch ethnicity.

**PCR-SSCP analysis**

A polymerase chain reaction–single strand conformation polymorphism (PCR-SSCP) method was optimised for the simultaneous detection of the SNPs at positions +869 and +915 in the TGFB1 gene and their haplotypes. The method has been described previously by García-González et al. In that study in a Dutch population, the alleles were shown to form three haplotypes, denoted as haplotype 1 (TGFB1+869T–TGFB1+915G), haplotype 2 (TGFB1+869C–TGFB1+915G), and haplotype 3 (TGFB1+869C–TGFB1+915C). In brief, the region containing the SNP at position +869 (NCBI SNP CLUSTER ID: rs1982073) and the SNP at position +915 of the TGFB1 gene (NCBI SNP CLUSTER ID: rs1800471) was amplified by PCR in a Perkin-Elmer 9600 thermal cycler (Perkin-Elmer, Norwalk, Connecticut, USA) according to the following parameters: 97°C for 90 seconds, 61°C for 90 seconds, and 72°C for 60 seconds for three cycles, followed by 32 cycles of 97°C for 30 seconds, 61°C for 60 seconds, and 72°C for 60 seconds, with a final elongation at 72°C for 10 minutes.

PCR products diluted twofold in 99% formamide were heated at 95°C for three minutes and placed on ice. Electrophoresis on a pre-cast non-denaturing 20% polyacrylamide PhastGel at 20°C and silver staining were done semi-automatically on the PhastSystem™ (Amersham Pharmacia, LKB Biotechnology AB, Uppsala, Sweden).

**Statistical analysis**

Allele and genotype frequencies were tested for Hardy–Weinberg equilibrium (HWE) by the $\chi^2$ test. To compare frequencies $\chi^2$ testing was used or, when appropriate, Fisher’s exact test. A two sided $p$ value of $<0.05$ was considered significant. The magnitude of association was expressed as the odds ratio (OR) with 95% confidence intervals (CI). The statistical analyses were undertaken using SPSS 10.0 for Windows.

**RESULTS**

Characteristics of the 134 AS patients are summarised in table 1. Genotype and distribution of allele frequencies in AS patients and controls for the TGFB1 T869C and TGFB1 G915C polymorphism are shown in table 2.

The data for the controls were published previously by Schrijver et al. The genotype frequencies in the control group did not deviate from HWE equilibrium. No significant differences were observed between AS patients and controls in the frequencies of the carrihership of the allele TGFB1 869C (OR = 0.96 (95% CI, 0.69 to 1.33), $p = 0.81$) and the allele TGFB1 915C (OR = 1.62 (0.93 to 2.84), $p = 0.09$) (table 2); neither were significant differences observed between AS patients and controls in the frequencies of the carrihership of the three TGFB1 haplotypes (table 3).

No significant associations were found between carriage of either of the alleles TGFB1 869C and TGFB1 915C, or their haplotypes, and sex, past or present peripheral arthritis or acute anterior uveitis, age at first complaints, years between these first complaints and the diagnosis of AS, and the number of patients with at least one first degree family member with AS (data not shown).

**DISCUSSION**

In this study the TGFB1 T869C and TGFB1 G915C SNPs or their haplotypes were not significantly associated with susceptibility to AS or its manifestations in a white Dutch population. Genotype and allele frequencies of the TGFB1 T869C and TGFB1 G915C SNPs in our control population were similar to those reported in some European studies undertaken in healthy controls from Northern Ireland and France and from the United Kingdom, as summarised by García-González et al.

Eight polymorphisms have been identified in TGFB1: three in the upstream region of the gene at positions –988, –800, and –509 from the transcription start site; one in the non-translated region at position +72; two non-synonymous coding SNPs in the signal peptide sequence: TGFB1 T869C which changes codon 10 (Leu\textsuperscript{10}→Pro), and TGFB1 G915C which changes codon 25 (Arg\textsuperscript{25}→Pro); one SNP in the region of the gene coding for the precursor part of the protein is not present in the active form: TGFB1 C1632T which changes codon 263 (Thr\textsuperscript{263}→Ile); and a one base pair deletion in intron 4 (713-8delC). The TGFB1 promoter polymorphisms and the T869C variant account for most of the diversity at this locus and a relatively small increase in diversity is

### Table 1: Demographic and clinical characteristics of patients with ankylosing spondylitis (n = 134)

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Frequency</th>
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<tbody>
<tr>
<td>Age (years)*</td>
<td>50.9 (12.9) (19 to 79)</td>
</tr>
<tr>
<td>Age at first complaints (years)†</td>
<td>23.0 (19.0 to 31.5)</td>
</tr>
<tr>
<td>Age at diagnosis (years)†</td>
<td>34.7 (10.6) (16 to 63)</td>
</tr>
<tr>
<td>Years between first complaints and diagnosis†</td>
<td>7.0 (2.0 to 12.5)</td>
</tr>
<tr>
<td>Women</td>
<td>18.7%</td>
</tr>
<tr>
<td>HLA-B27 positive</td>
<td>94%</td>
</tr>
<tr>
<td>Iridocyclitis</td>
<td>43%</td>
</tr>
<tr>
<td>Peripheral arthritis</td>
<td>41%</td>
</tr>
<tr>
<td>First degree relatives with AS</td>
<td>28%</td>
</tr>
</tbody>
</table>

*Median (interquartile range). †Mean (SD) (range).  

### Table 2: TGFB1 genotype and allele frequencies in patients with ankylosing spondylitis and controls

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Controls (n = 194)</th>
<th>Patients (n = 134)</th>
<th>OR (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>TGFB1 T869C</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>TT</td>
<td>80 (41.2)</td>
<td>57 (42.5)</td>
<td>64.6</td>
</tr>
<tr>
<td>TC</td>
<td>94 (48.5)</td>
<td>59 (44.0)</td>
<td>35.4</td>
</tr>
<tr>
<td>CC</td>
<td>20 (10.3)</td>
<td>18 (13.4)</td>
<td>0.96 (0.69 to 1.33) (p = 0.81)</td>
</tr>
<tr>
<td>TGFB1 G915C</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GG</td>
<td>168 (86.6)</td>
<td>107 (79.9)</td>
<td>89.6</td>
</tr>
<tr>
<td>GC</td>
<td>26 (13.4)</td>
<td>26 (19.4)</td>
<td>1.63 (0.93 to 2.84) (p = 0.09)</td>
</tr>
<tr>
<td>CC</td>
<td>0</td>
<td>1 (0.7)</td>
<td>10.4</td>
</tr>
</tbody>
</table>

AF, allele frequency; CI, confidence interval; OR, odds ratio.
attributable to the rare variants TGFB1 C1632T and TGFB1 G915C.

The TGFB1 polymorphism 713-8delC is more frequent in patients with osteoporosis than in normal controls and seems to be associated with very low bone mass in osteoporotic women. As bone mineral density is often decreased in AS patients, TGFB1 polymorphisms may play a crucial role in the pathogenesis of AS, although the association with transforming growth factor-beta1 production, fibrotic lung disease. Apical pulmonary fibrosis is rare but well recognised in patients with AS. No association between Crohn's disease or ulcerative colitis, which are clinically related to AS, and TGFB1 SNPs has been found in previous studies.

The genotype TGFB1 915GG has been related to higher serum concentrations of TGFB1 in a previous study. A combined study from Finland and the United Kingdom found a weak association between age of symptom onset of AS and the TGFB1 G915C SNP. Furthermore, this study noted a weak positive association of the rare allele TGFB1 1632T with AS and also with a younger age at symptom onset of AS. These investigators concluded from their study that the polymorphisms (G-800A, C-509T, T+869C, G+915C and C+1632T) in the TGFB1 gene play at most a small role in AS and that other genes on chromosome 19 are involved in the susceptibility to AS.

The results from our study do not show an association between the TGFB1 G915C SNP and AS, although the frequency of allele TGFB1 915G was greater in AS patients than in controls, but this difference did not reach significance (p = 0.09). In our group of AS patients no association between the TGFB1 G915C SNP and a younger age at symptom onset of AS could be found.

As TGFβ may play a crucial role in the pathogenesis of AS, especially in new bone formation, further research is necessary to elucidate the role of TGFβ1 in this disease, and possibly open the way towards new pharmacological approaches for preventing the most disabling phenomenon of this disorder, ankylosis.

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Table 3 Genotype, phenotype, and haplotype frequencies of the TGFB1 polymorphisms in patients with ankylosing spondylitis and controls

<table>
<thead>
<tr>
<th></th>
<th>Controls (n = 194)</th>
<th>Patients (n = 134)</th>
</tr>
</thead>
<tbody>
<tr>
<td>TGFB1 haplotype</td>
<td>n (%)</td>
<td>PF, n (%)</td>
</tr>
<tr>
<td>1.1</td>
<td>80 (41.2)</td>
<td>174 (89.7)</td>
</tr>
<tr>
<td>1.2</td>
<td>76 (39.2)</td>
<td></td>
</tr>
<tr>
<td>1.3</td>
<td>18 (9.3)</td>
<td></td>
</tr>
<tr>
<td>2.2</td>
<td>12 (6.2)</td>
<td>96 (49.5)</td>
</tr>
<tr>
<td>2.3</td>
<td>8 (4.1)</td>
<td></td>
</tr>
<tr>
<td>3.3</td>
<td>0</td>
<td>26 (13.4)</td>
</tr>
</tbody>
</table>

*Overall odds ratios for carriership of haplotypes 1–3.

Haplotype 1: TGFB1 B869T–TGFB1 B915G; haplotype 2: TGFB1 B869C–TGFB1 B915C; haplotype 3: TGFB1 B869C–TGFB1 B915C.

REFERENCES


24 Langdahl BL, Knudsen JY, Jensen HK, Gregersen N, Eriksen EF. A sequence variation: 713-8delC in the transforming growth factor-beta 1 gene has higher prevalence in osteoporotic women than in normal women and is associated with very low bone mass in osteoporotic women and increased bone turnover in both osteoporotic and normal women. Bone 1997;20:289–94.


