Conspicuous synovial lymphatic capillaries in juvenile idiopathic arthritis synovitis with rice bodies

E Rovenska, S Stvrtina, O Greguska, L Pravda, J Rovensky


The paper of Mohr discussing the development of rice bodies with apatite crystals in fibrinous debris synovitis in rheumatoid arthritis1 prompted us to describe our recent morphological findings.

During synovectomy in a 33 year old woman with a longlasting systemic form of juvenile idiopathic arthritis (JIA) a large number of rice bodies and numerous synovial villi connected to the synovial membrane (SM) by very thin stalks were visible in the joint space (JS). Light microscopy of paraffin sections showed that the villi contained fibrin and had degenerated. In the SM, mononuclear infiltration, neoangiogenesis, intimal layer hyperplasia, and fibrin at the synovial surface were found. Light microscopy of serial semi-thin resin sections enabled a distinction to be made between lymphatic capillaries (LC). Prominent LC were found under villous fibrin (fig 1A). In connective tissue around the LC, macrophages were seen. Cells and debris were rarely seen inside the lumina of the LC (fig 1B). LC were also seen in areas of the SM not covered with fibrin (fig 1C). These LC were situated in the subintimal connective tissue and were often surrounded by numerous mononuclear cells. In some of these LC, cells (mostly lymphocytes) were found.

Kuhns presented a detailed morphological study of lymphatic drainage of synovial joints in rabbits.2 He discovered that inflammation in the synovial tissue decreased the ability of LC to absorb material larger than that of molecular size and presumed that persistent inflammation was, to a certain extent, dependent on the non-functioning of the lymphatic vessels. Later, Pullinger and Florey proved that LC proliferated in acute inflammation and repair.3 They demonstrated that LC proliferated also in chronic inflammation induced in the skin of mice, and emphasised that debris was removed from the damaged areas by the LC, either directly or by phagocytic cells. Recently, in rheumatoid arthritic synovium, debris and cells were seen inside the lumina of LC and, moreover, endothelial microvalves were visualised in the walls of the LC by transmission electron microscopy.4

Endothelial microvalves of LC probably have an important role in drainage of excessive tissue fluid, allowing cells and debris to be removed from SM connective tissue spaces into the lymph. The LC are an integral part of connective tissue, in which prelymphatic tissue channels have been described.5 6 In the patient reported in this paper, fibrin deposition and mononuclear infiltration may have blocked part of the prelymphatic tissue channels in the SM, thus reducing the drainage of the JS. This might have contributed to the formation of rice bodies in synovial fluid (SF).

Rice bodies in rheumatoid SF contain mononuclear cells, mostly macrophagic in appearance.7 Accumulation of rice bodies in the JS may contribute to increased cytokine levels in the SF. It is known that SF cytokines can modulate the level of vascular endothelial growth factor (VEGF) secretion.8 VEGF-C and D were shown to stimulate lymphangiogenesis.9 Recently, mature VEGF-C was found in rheumatoid arthritis synovial tissue.10

Figure 1

Synovial LC (asterisks) in semi-thin resin sections stained with toluidine blue. (A) LC in the SM area covered with fibrinous material (f). Walls of the LC are composed of endothelial cells only. (B) Mononuclear phagocytes (arrows) in the vicinity of the LC. One phagocyte (large arrow) and debris (arrowheads) are visualised inside the lumina of the LC. (C) Large LC with an irregular shaped lumen surrounded by numerous mononuclear cells, situated in the sublining connective tissue of the SM under the intimal layer showing hyperplasia. Some blood capillaries (arrows) and a venule (arrowhead) are also visualised. JS, joint space. Scale bars = 100 μm.
Our observation of conspicuous LC suggests that lymphangiogenesis may occur in JIA synovitis. In chronic synovitis, neogenesis of LC seems to be aimed at improving drainage and thus promoting homeostasis in the JS.

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REFERENCES

Periostitis as the initial manifestation of systemic vasculitis
P M Aries, M Reuter, P Lamprecht, W L Gross

The greatest challenge in diagnosing vasculitis is the diversity of its clinical presentation. Awareness of the heterogeneity of uncommon manifestations can be decisive for the course of the disease.

CASE REPORT
We report on a patient presenting with periostitis as the initial manifestation of systemic vasculitis. A 38 year old female patient complained about progressively painful swelling and reddening of the distal right lower leg for several weeks. The patient had been healthy until then and had no history of arterial or venous insufficiency. She presented at hospital with reduced pulses and a severe compartment syndrome of the lower extremities on admission. The patient had been healthy until then and had no history of arterial or venous insufficiency. She presented at hospital with reduced pulses and a severe compartment syndrome of the lower extremities on admission.

Angiography. According to the nomenclature of the Chapel Hill Consensus Conference, the patient’s disease was diagnosed as polyarteritis nodosa. Treatment was switched to cyclophosphamide (the so-called “NIH standard”: cyclophosphamide 2.0 mg/kg body weight per day with daily prednisolone po). After induction of remission, treatment was switched to azathioprine. Follow up bone radiography disclosed a moderate reduction of the new periostal bone formation and clinical remission was maintained at a 3 year follow up.

DISCUSSION
The patient presented initially with an unusual manifestation of systemic vasculitis. Vasculitis restricted to the local vascular region may be the initial manifestation of systemic vasculitis. In this particular case, vasculitis of the periosteum might have induced local hypoxia of the bone, with subsequent release of bone derived growth factors and manifestation of periostitis. Periostitis is seen in many other conditions but is not common in necrotising vasculitis. It was described for the first time by Lovell and Scott in 1956. Until now only a few cases of periostitis in patients with polyarteritis nodosa have been reported; remarkably, the lower extremities were affected in all cases. However, periostitis has also been reported in other forms of systemic vasculitis. Most cases responded well to glucocorticoids. In refractory cases other cytotoxic treatment like methotrexate, azathioprine, or cyclophosphamide may be useful.

Thus, as demonstrated by this case, in patients with painful swelling of the lower limb, clinicians should consider periostitis as an unusual manifestation of systemic vasculitis.

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www.annrheumdis.com
Presence of rheumatoid factor and antibodies to citrullinated peptides in systemic lupus erythematosus


Rheumatoid factor (RF) is found commonly in patients with systemic lupus erythematosus (SLE), and has been associated with a more benign disease course.1 2 Anti-citrullinated peptide antibodies (ACPA) are more specific for rheumatoid arthritis (RA).3–5 Several assays for ACPA detection have been developed: among others, an enzyme linked immunosorbent assay (ELISA) for anti-cyclic citrullinated peptide (anti-CCP) antibodies3 and a line immunoassay (LIA) for antibodies to peptide A (pepA) and peptide B (pepB), two synthetic citrullinated peptides.4 Few reports exist about the presence of ACPA in SLE. Although patients with SLE are often part of the control group when determining the specificity of ACPA for RA, SLE alone is seldom studied. Mediwake et al found that 3/66 patients with SLE were positive for anti-CCP1 antibodies; two of them had erosive arthritis.6 We investigated the presence of RF and three different ACPA (anti-CCP, anti-pepA, and anti-pepB antibodies) in SLE.

Two hundred and thirty five patients with SLE, meeting American College of Rheumatology (ACR) revised criteria for classification of SLE,7 were prospectively included in four European centres. The study investigated associations between symptoms and specific antinuclear reactivities and has been reported elsewhere.8 Serum was available for further analysis in 201 patients. The male to female ratio was 25:176. The mean age was 40 years. The study was...
Letters 331

Table 1 Characteristics of ACPA positive patients with SLE

<table>
<thead>
<tr>
<th>Patient No</th>
<th>RF</th>
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<th>Anti-PepA</th>
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<th>Rx</th>
<th>RA crit</th>
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<td>Ro60</td>
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</table>

RF titres and anti-CCP2 concentrations (U/ml; cut off point 42 U/ml) are given. Anti-pepA and anti-pepB antibodies were scored –, 1+, 2+, or 3+. Fine antinuclear reactivities were noted. Shared epitope (SE) status is recorded as the presence of 0, 1, or 2 copies (0, 1, 2). Radiographic data (Rx) are listed as the presence (+) or absence (−) of erosions. ACR classification criteria for RA (RA crit) were noted as fulfilled (+) or not (−). Clinical symptoms being part of the ACR criteria for SLE are listed. NA = not available.

approved by the local ethics committees. Informed consent was obtained from all patients.

Fine antinuclear reactivities were determined with INNO-LIA-ANA Update (Innogenetics, Ghent, Belgium) and by indirect immunofluorescence on Cricetidae luciae. RF was detected using the latex fixation method (Becton Dickinson, Sparks, Maryland, USA). Titres >160 were considered positive, which corresponds to a specificity for RA of 95.9% in an independent control cohort, consisting of 146 patients with rheumatic complaints but no RA (data not shown). Anti-CCP2 antibodies were detected by ELISA (ImmunoScreen RA, mark 2, Eurodagnostica, Arnhem, Netherlands). A cut off value of 42 U/ml was used. Anti-pepA and anti-pepB antibodies were detected by a research LIA (Innogenetics). During each run, a strip was developed using a control serum, providing a cut off intensity for each antigen line. In the control population mentioned earlier, all three ACPA had a specificity of at least 98.5%. The RA associated HLA-DR shared epitope (SE) was determined with INNO-LiPA (Innogenetics).

χ² Tests were used to determine associations. Antibody frequencies were compared using the McNemar test.

Anti-CCP2 antibodies were found in 11/201 (5.5%) patients, anti-pepA antibodies 3 (1.5%) patients, and anti-pepB antibodies in 5 (2.5%) patients. Table 1 shows the characteristics of patients positive for ACPA. Anti-CCP2 antibodies were significantly more frequent then anti-pepA antibodies (p = 0.109). It is important to notice that in an independent control cohort all three ACPA obtained comparable specificities of at least 98.5%. Apparently, the different substrates behave differently in SLE. RF was found in 26 (12.9%) patients, which was significantly more frequent than anti-pepA (p<0.001), anti-pepB (p<0.001), and anti-CCP2 antibodies (p = 0.006). Although the diagnosis in the ACPA positive patients was SLE, and all fulfilled classification criteria for SLE, ACR criteria for RA were also fulfilled in 6/10 evaluable patients, with 3/10 carrying an SE allele; radiographic erosions were present in 3/7 evaluable patients.

Our data suggest that the presence of ACPA does not exclude a diagnosis of SLE. It remains to be evaluated whether ACPA in SLE predispose for a chronic RA-like arthritis in this case.

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INNO-LIA and LIA are trademarks of Innogenetics NV, Ghent, Belgium.

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REFERENCES

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Lack of efficacy of rituximab in Felty’s syndrome

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Felty’s syndrome (FS) is defined by the coexistence of rheumatoid arthritis (RA), neutropenia, and splenomegaly. The mechanisms underlying the neutropenia of FS may involve both cellular and humoral immunity, with a possible role of granulocyte-colony stimulating factor (G-CSF) antibodies. Various disease modifying antirheumatic drugs have been used to treat FS, but with varying success as this syndrome may arise in response to the excessive immune reaction found in RA. Interest has focused recently on a new biological tool in the treatment of RA, rituximab, a chimeric monoclonal antibody specific for human CD20 which targets B lymphocytes. Accordingly, we investigated here the safety and efficacy of rituximab in two patients presenting with active RA and severe and refractory FS.

METHODOLOGICAL RESULTS

Two men, were studied, aged 67 (patient 1) and 53 (patient 2) years, with a duration of RA of 6 and 11 years, respectively. FS had been diagnosed respectively 5 and 3 years ago, and RA remained active in both patients despite corticotherapy and respectively one (sulfasalazine) and two (sulfasalazine and methotrexate) previous disease modifying antirheumatic drugs. Anti-tumour necrosis factor treatment was not used because of neutropenia and the risk of severe infection. The absolute neutrophil count was persistently less than 0.8×10⁹/l and complicated with recurrent sino-pulmonary infections. There was no suggestion of congenital hypogammaglobulinaemia and, in particular, no sign of selective IgG2 immunodeficiency. Blood and bone marrow immunophenotyping did not disclose any features of myelodysplasia or lymphoproliferation, or any large granular lymphocytes. No other classical cause of neutropenia, such as toxicity, chronic infection, vitamin deficiency, or liver disease, was present. Anti-G-CSF (IgG) antibodies, which were determined by enzyme linked immunosorbent assay (ELISA), were detected in one patient without previous administration of haematopoietic factor (G-CSF).

Owing to the presence of refractory RA associated with severe FS, rituximab was administered as an intravenous infusion at a dose of 375 mg/m² once weekly for 4 weeks. Concomitant treatment consisted of prednisone (15–20 mg/day) for more than 12 months in both patients and methotrexate (20 mg/week) since March 2003 in patient 2. The duration of follow up was 6 months. Rituximab was well tolerated, with no signs of allergic or toxic reactions. A significant improvement of neutrophil count was observed in both patients. The neutrophil count increased from 22.6×10⁹/l at baseline to 61.9×10⁹/l at 1 month after the first rituximab infusion in patient 1 and from 29.6×10⁹/l to 42.1×10⁹/l in patient 2. In addition, CRP decreased in both patients from 11.1 mg/l to 2.8 mg/l in patient 1 and from 12.1 mg/l to 3.1 mg/l in patient 2. The duration of remission was 6 months. Rituximab was well tolerated, with no signs of allergic or toxic reactions. A significant improvement of neutrophil count was observed in both patients. The neutrophil count increased from 22.6×10⁹/l at baseline to 61.9×10⁹/l at 1 month after the first rituximab infusion in patient 1 and from 29.6×10⁹/l to 42.1×10⁹/l in patient 2. In addition, CRP decreased in both patients from 11.1 mg/l to 2.8 mg/l in patient 1 and from 12.1 mg/l to 3.1 mg/l in patient 2.

Table 1: Clinical and biological features of two patients with FS treated with rituximab

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<th>Normal range</th>
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<th>Neutrophil count</th>
<th>ESR</th>
<th>CRP</th>
<th>CD19+ cells</th>
<th>IgG</th>
<th>IgM</th>
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W0, biological data were obtained before first infusion of rituximab. DAS28, 28 joint count Disease Activity Score; ESR, erythrocyte sedimentation rate; CRP, C reactive protein; RF, rheumatoid factor; ND, not determined.
tolerated and efficiently controlled the clinical and biological activity of RA in patient 2, who fulfilled the American College of Rheumatology 50 response criteria and showed a marked decrease in serum levels of rheumatoid factor. However, results for FS were disappointing, because no increase in neutrophil count or modification of infection rates could be detected (table 1). In patient 1, a decrease in neutrophil count was observed at week 12, but without any clinical anomaly. Biological controls showed no modification of levels of anti-G-CSF antibodies, no appearance of anti-granulocyte antibodies, and no large granulocyte lymphocyte proliferation. 3

**DISCUSSION**

Several factors might account for the lack of efficacy of rituximab in the treatment of FS. Firstly, although different autoreactive B cells may be involved in the pathology of FS, the inability of rituximab to bind to plasma cells, which are CD20 negative, might prevent it from acting on FS. Nevertheless, the efficacy of rituximab in certain conditions associated with autoantibodies is not correlated with a reduction of these antibodies, which would suggest that in addition to autoantibody production, other roles of B cells (immunoglobulins, antigen presentation, T cell cooperation) are important in the pathogenesis of such diseases. 3

Secondly, a subpopulation of T lymphocytes having an antigranulocyte activity may exist independently of B cells in some forms of FS. 3

In conclusion, the lack of efficacy of rituximab in these two patients with FS raises some important questions about the mechanisms responsible for FS and the best therapeutic strategy to adopt.

**REFERENCES**


**Antinuclear and antiphospholipid autoantibodies in patients with peripheral arterial occlusive disease**

K Kroeger, H Mouradi, E Kreuzfelder, G Rudofsky, H Grosse-Wilde

According to the Chapel Hill Consensus Conference, large peripheral arteries are only affected by giant cell vasculitis and, in rare cases, by polyarteritis nodosa. 1-3 Vasculitis becomes apparent through involvement of typical organs (lung, kidney, skin) or raised C reactive protein (CRP) level or erythrocyte sedimentation rate (ESR). Thus, a specific diagnostic effort to exclude vasculitis as an underlying disease in patients with peripheral arterial occlusive disease (PAOD) may be unnecessary. On the other hand, there is increasing evidence that humoral immunity may have a role in the pathogenesis of atherosclerosis. 1,2 Antinuclear antibodies were reported in 70% of patients with severe coronary heart disease (CHD), compared with in only 17% in the control group. 3 Thus, we prospectively studied the importance of autoantibody determination in patients with symptomatic PAOD.

**METHODS AND RESULTS**

Six hundred and ninety eight patients (mean (SD) age 68 (10) years) referred for treatment of PAOD between 1998 and 1999 were included. In 121 patients with PAOD (aged 61 (12) years) with a low atherosclerotic risk profile, or with rarefied distal arteries without media calcification, or with raised ESR or CRP not due to a local infection, the following autoantibodies were determined: antinuclear antibodies (ANA) by an indirect immunofluorescence technique; antibodies against extractable nuclear antigens (Scl-70, RNP, SSA, SSB, Jo-1, SM) by western blot; double stranded DNA antibodies, antineutrophil cytoplasmic antibodies (c- and pANCA), and antiphospholipid antibodies (cardiolipin, phosphatidylserine (APSAs), and β2-glycoprotein) by enzyme linked immunoassay. To stratify the importance of autoantibody determination all patients with increased autoantibody concentration were clinically and sonographically followed up for 24 (6) months for evidence of vasculitides or collagen disease. A multivariate logistic regression analysis was performed to evaluate the importance of CRP and ESR in patients with autoantibody concentrations above the appropriate reference value.

Thirty eight of the 121 patients had increased autoantibody concentrations (table 1). ANA were the most common autoantibodies detected in 14 patients followed by APSA in 11, and β2-glycoprotein antibodies in 12. Patients with increased autoantibody concentration did not differ in their PAOD stages and affected segments, but in patients with increased autoantibody concentrations the ESR was higher (p = 0.0043). The ESR at 2 hours was associated with an odds ratio of 7.1 (95% confidence interval 1.5 to 33.8) in determination of increased autoantibody concentrations. During the follow up of 24 (6) months no vasculitides or collagen diseases could be detected by clinical examination or by nailfold capillary microscopy, pulmonary or gastrointestinal imaging in the 38 patients.

**DISCUSSION**

The group of 121 patients with PAOD analysed is a group selected individually from all the patients, but represents those patients in whom possible vasculitis may be present.
Raised concentrations of autoantibodies were common in the patients investigated. Increased autoantibody concentrations significantly correlated with a raised ESR.

As far as we know, our data are the first to report the results of autoantibody determinations in a large group of patients with PAOD. In contrast with the high rate of ANA in patients with coronary atherosclerosis, the prevalence of ANA in our patients with PAOD was much lower. Whether this difference in the prevalence of ANA was due to different forms of atherosclerosis, or due to specific differences in coronary and peripheral manifestations can only be speculated.

In agreement with the coronary studies, no association of the determined autoantibodies with classical risk factors was found. The higher ESR in the patients with increased autoantibody concentrations might be associated with a higher degree of inflammatory activity of the atherosclerosis. Antiphospholipid and β2-glycoprotein antibodies, which are most relevant in association with atherosclerosis, did not seem to lead to a prognosis, but antibody determination in larger groups of non-selected patients is desirable.

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REFERENCES
Incidence of primary systemic vasculitides in Vilnius: a university hospital population based study

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As knowledge of the epidemiology of primary systemic vasculitides (PSV) is fragmentary, we attempted to investigate the incidence of temporal arteritis (TA), Takayasu’s arteritis (TAA), polyarteritis nodosa (PAN), Wegener’s granulomatosis (WG), Churg-Strauss syndrome (CSS), Henoch-Schönlein purpura (HSP), and hypersensitivity vasculitis (HSV) in Vilnius according to the American College of Rheumatology (ACR) 1990 criteria and to compare the data with the results from selected European studies. To be included in this study the patients had to (a) have been diagnosed with systemic vasculitides in the 10 year period from 1990 to 1999 and (b) have been resident in Vilnius at the time of diagnosis. Patients referred to Vilnius University Hospital rheumatology department were prospectively included in the study. Also, the patients’ registration books from tertiary nephrology, dermatology, and internal medicine departments were searched for a diagnosis of PSV retrospectively, but following the same inclusion criteria. Additionally, the data files of the centre of pathology available from 1995 onwards, including renal register, were searched.

We applied ACR 1990 criteria for the classification of PSV; however, for patients classified as HSV, the term cutaneous leucocytoclastic vasculitis (LCV) was equally used. Patients with microscopic polyangiitis (MPA) were included in the PAN group, and PAN criteria applied to both conditions. The group of PAN and HSV/LCV were reanalysed according the definition for MPA. The denominator population was the adult population over 16 years from Vilnius city, which comprised 468 504 people (53.7% female) in 1999.

Table 1 Number and annual incidence (per million inhabitants) of primary systemic vasculitides in Vilnius compared with the selected European studies conducted during the past decade

<table>
<thead>
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<tbody>
<tr>
<td>Giant (temporal arteritis) cell arteritis</td>
<td>11 (50-87) 2.3/10^6</td>
<td>24 31.9/10^6</td>
<td>– 45.6/10^6</td>
<td>110</td>
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<tr>
<td>Takayasu arteritis</td>
<td>6 (16-49) 1.3/10^6</td>
<td>0 0</td>
<td>0 0</td>
<td>0</td>
</tr>
<tr>
<td>Polyarteritis nodosa</td>
<td>36 (17-72) 7.7/10^6</td>
<td>5 6.7/10^6</td>
<td>33 8.0/10^6</td>
<td>13 6.9/10^6</td>
</tr>
<tr>
<td>Hypersensitivity vasculitis/cutaneous leucocytoclastic angiitis</td>
<td>122 (17-85) 26.0/10^6</td>
<td>45 2.7/10^6</td>
<td>2 17.8/10^6</td>
<td>37 29.7/10^6</td>
</tr>
<tr>
<td>Henoch-Schönlein purpura</td>
<td>14 (16-21) 3.0/10^6</td>
<td>3 6.7/10^6</td>
<td>3 1.2/10^6</td>
<td>3 14.0/10^6</td>
</tr>
<tr>
<td>Churg-Strauss syndrome</td>
<td>6 (27-58) 1.3/10^6</td>
<td>2 2.7/10^6</td>
<td>6 2.4/10^6</td>
<td>2 1.1/10^6</td>
</tr>
<tr>
<td>Wegener’s granulomatosis</td>
<td>10 (22-68) 2.1/10^6</td>
<td>5 6.7/10^6</td>
<td>21 8.5/10^6</td>
<td>9 4.8/10^6</td>
</tr>
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*7.2/million (95% CI 3.8 to 13.3) for the population aged 50 years and older; †Norway, 150 000 adult inhabitants in the Kristiansand study, 414 500 in the Norwich study, 250 000 in the Lugo study, 468 504 in Vilnius refer to the study period; ‡polyarteritis nodosa and hypersensitivity vasculitides in this study are reported for the period from 1988 to 1997" and from 1990 to 1994," respectively.
Bone mineral density in patients with rheumatoid arthritis treated with infliximab

M Vis, A E Voskuyl, G J Wolbink, B A C Dijkmans, W F Lems for the OSTAR study group


Methods and Results
This open cohort study consisted of consecutive patients with RA, who were treated with infliximab in the Sloetervaart Hospital and the VU University Medical Centre. All patients fulfilled the ACR 1987 criteria of RA and had active disease (defined by the modified 28 joint count Disease Activity Score (DAS28) of at least 3.2). Infliximab was given intravenously at 0, 2, 6, 14, weeks and from the fourth infusion every 8 weeks in a dose of 3 mg/kg. At each visit the DAS28 was calculated and changes in drug treatment were recorded. BMD measurements (g/cm²) of the hip (total hip) and lumbar spine (L1–4) were performed at baseline and after 1 year on a Hologic 4500.

Osteoporosis is a well known feature of rheumatoid arthritis (RA). Cross sectional studies have shown that patients with RA have a lower bone mineral density (BMD) than healthy controls. Disease activity, steroid use, and immobility are associated with loss of BMD in RA. It has been suggested that active treatment of patients with RA may prevent loss of BMD. The current most effective drugs in the treatment of RA are the tumour necrosis factor α blocking agents. The beneficial effects of short term treatment with infliximab on markers of bone metabolism in patients with active RA have recently been shown. From this we proposed the hypothesis that bone loss might be arrested in patients with RA during treatment with infliximab.
In total, 36 patients (29 (81%) female) were included into the study. Patients had a mean (SD) age of 53 (12) years, with a median (range) disease duration of 9.5 years (0–49). Methotrexate, prednisone, and bisphophonates were used by 100%, 50%, and 25% of the patients, respectively. The mean disease activity (DAS28) decreased from 5.6 at baseline to 3.8 at 6 weeks and stabilised around 3.6 for the rest of the studied period. In 36 patients dual x-ray absorptiometry (DXA) measurements of lumbar spine (L1–4) and in 30 patients DXA measurements of the hip were available at baseline and after 1 year. In four patients no DXA hip measurements were available because of bilateral hip replacement, and in two patients only one DXA of the hip was available owing to unknown causes.

Mean (SD) BMD at the lumbar spine increased non-significantly from 0.998 (0.205) to 1.001 (0.199) at 1 year (+1.1%, p = 0.117). BMD at the total hip decreased non-significantly from 0.857 (0.144) to 0.854 (0.132) at 1 year. (−0.3%, p = 0.683). In a linear regression model, changes in BMD at the hip or the spine were not associated with mean DAS28, prednisone use, or bisphosphonate use (data not shown).

**DISCUSSION**

This study indicates that BMD of the spine has a tendency to increase and that BMD of the hip slightly decreases during 1 year of treatment with infliximab. This is in contrast with previous longitudinal studies of patients with RA, in which a decrease of BMD was seen during conventional disease modifying antirheumatic drug treatment without tumour necrosis factor blockings agents (table 1).

In our view, these data suggest that treatment with infliximab can arrest generalised osteoporosis in patients with RA. This view is supported by the observation that markers of bone formation increased and markers of bone resorption decreased in the first 6 weeks of treatment with infliximab. We do realise that our observations are made in resorption decreased in the first 6 weeks of treatment with infliximab.7 We do realise that our observations are made in resorption decreased in the first 6 weeks of treatment with infliximab.

In summary, this study suggests that treatment with infliximab has a positive effect on BMD in patients with RA. Because patients with RA have an increased risk of bone loss and, subsequently, osteoporotic fractures, this might be an additional advantage of infliximab (above the well known favourable effect on disease activity and radiological damage), and warrants further study.

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We thank the members of the OOSTRA group: TK Kvien, G Haugeberg, T Uhlig, EA Haavardsholm (Oslo, Norway), and A Woolf (Truro, UK), for their valuable comments.

**REFERENCES**

An open study of pulse pamidronate treatment in severe ankylosing spondylitis, and its effect on biochemical markers of bone turnover

A P Cairns, S A Wright, A J Taggart, S M Coward, G D Wright

Osteoporosis is a common feature of ankylosing spondylitis (AS), and vertebral fractures are an increasingly recognised complication. A cumulative fracture prevalence of between 9.5% and 18% has been reported, with a six- to eightfold relative increased risk of vertebral fracture. Osteoporosis and new bone formation (syndesmophytyosis) suggest that disordered bone turnover has a role in disease pathogenesis in AS. Bisphosphonates accumulate at sites of increased bone turnover, and inhibit bone resorption by inducing osteoclast apoptosis, thereby improving bone density and reducing fracture rates. Pulse pamidronate has recently been used with clinical efficacy in postmenopausal osteoporosis. The effect of bisphosphonate treatment on biochemical bone turnover markers has not previously been studied in AS. We aimed at studying the efficacy of pulse pamidronate treatment in severe AS, and at determining its effect on biochemical bone turnover markers.

METHODS AND RESULTS

Patients with severe AS were treated with 6 monthly intravenous pulses of pamidronate, receiving 30 mg for the first infusion and 60 mg subsequently. The Bath AS Disease Activity Index (BASDAI) and the Bath AS Metrology Index (BASMI) scores were recorded, and C reactive protein (CRP) and erythrocyte sedimentation rate (ESR) measured at each visit. Fifteen patients participated (13 male, mean age 44). Mean disease duration was 14.8 years. All patients were receiving non-steroidal anti-inflammatory drugs; two were also receiving sulfasalazine, one methotrexate, and one azathioprine. No patients were currently receiving an oral bisphosphonate. Four patients had a history of uveitis, two psoriasis, two inflammatory bowel disease, and eight peripheral arthritis.

Fasting blood samples were taken monthly for measurement of biochemical bone turnover markers, according to the manufacturers’ instructions. Degradation products of type I collagen C-terminal telopeptides were measured with the serum crosslaps enzyme linked immunosorbent assay (ELISA; Nordic Bioscience Diagnostics A/S, Herlev, Denmark). Serum bone GLA protein was measured using the N-MID osteocalcin ELISA kit (Nordic Bioscience Diagnostics A/S, Herlev, Denmark). Bone-specific alkaline phosphatase was measured with the Access Ostase assay (Beckman Coulter Inc, Fullerton, California, USA). Non-parametric analyses (Wilcoxon signed rank tests) on an intention to treat basis were used to analyse the data.

Three patients did not complete the study (arthralgia/headache, back pain/nausea, and myalgia, respectively). The mean total dose of pamidronate received was 277 mg. The initial median serum crosslaps was 1845.0 ng/ml. Mean (SD) population serum crosslaps concentrations are 506 (255) ng/l for postmenopausal women, 321 (155) ng/l for premenopausal women, and 332 (190) ng/l for men. The initial median ostase concentration was 12.4 μg/l. Mean (SD) population ostase concentrations are 12.3 (4.3) μg/l for men, 8.7 (2.9) μg/l for premenopausal women, and 13.2 (4.7) μg/l for postmenopausal women. The initial median osteocalcin concentration was 19.5 ng/ml. Mean (SD) population osteocalcin concentrations are 17.9 (6.5) ng/ml for premenopausal women, 28.4 (9.5) ng/ml for postmenopausal women, and 21.4 (9.1) ng/ml for men.

Median serum crosslaps fell from 1845.0 to 556.5 ng/l (Z = −3.29, p = 0.001) (fig 1). Median serum osteocalcin fell from 19.5 to 16.2 ng/ml (Z = −2.34, p = 0.02). Median serum ostase fell from 12.4 to 9.6 μg/l (Z = −3.11, p = 0.02). The median BASDAI improved from 6.80 to 3.75 (Z = −1.98, p = 0.048), but there was no significant improvement in the median BASMI (initial 8.00 v final 7.50, Z = −1.64, p = 0.10). There were non-significant trends of reduction in median CRP (initial 36.4 v final 26.6 mg/l, Z = −1.89, p = 0.06) and median ESR (initial 38 v final 29 mm/1st h, Z = −0.71, p = 0.48). There were no significant correlations between clinical measures and bone turnover markers.

DISCUSSION

Pulse pamidronate treatment significantly reduced all three biochemical bone turnover markers. This was particularly marked for serum crosslaps, the bone resorptive marker, where there was a 69.8% relative reduction. Such marked reduction of the crosslaps concentration suggests that bisphosphonates have a role in the management of the osteoporosis of AS. Ultimately, studies assessing fracture rates in patients with AS receiving bisphosphonates would be of great interest. One possibility is that by reducing the rate of new bone formation, bisphosphonates may also reduce syndesmophyte formation. However, longer term controlled
Association between interleukin 6 gene polymorphisms and Behçet’s disease in Korean people

H K Chang, W C Jang, S B Park, S M Han, Y H Nam, S S Lee, J U Kim, H S Lee

Interleukin (IL) 6 is an important mediator of inflammatory and immune responses, and IL6 gene polymorphisms are known to play a part in chronic inflammatory and autoimmune disorders.1,2 Increased IL6 plasma levels and enhanced IL6 mRNA expression have been found in patients with active Behçet’s disease.3,4 Therefore, this study aimed at investigating the associations between Behçet’s disease in Korean people and two functional IL6 gene polymorphisms—namely, a single nucleotide polymorphism at −174 (G → C) in the IL6 gene promoter (IL6prom) and a variable number of tandem repeat polymorphisms in the 3′ flanking region of the IL6 gene (IL6vntr): the terms IL6prom and IL6vntr were designated as in a previous study.5

METHODS AND RESULTS

The study group included 89 Korean patients with Behçet’s disease (36 men, 53 women; mean (SD) age 39.1 (8.5)), fulfilling the International Study Group criteria,6 and 123 controls (47 men, 76 women; mean (SD) age 43.1 (13.4)). The cumulative history of severe manifestations was investigated during the disease course.7 Analyses of IL6prom and IL6vntr were carried out in all the subjects by polymerase chain reaction (PCR)-restriction fragment length polymorphism and PCR genotyping, respectively.1,8 Significance was evaluated using Fisher’s exact test or t test and defined as p < 0.05: p values with Bonferroni’s correction (pcorr) were calculated in certain cases. Haplotype and linkage disequilibrium (LD) analyses were assessed using the estimated haplotype (EH) programme.9

There was no evidence of genetic association conferred by the IL6prom polymorphism. In the case of the IL6vntr, four genotypes were identified with the following frequencies in the controls: AB, 2 (1.6%); BB, 117 (95.1%); BC, 3 (2.4%); CC, 1 (0.8%); and in the patients with Behçet’s disease: BB, 77 (86.5%); BC, 12 (13.5%). Because the vast majority (98.6%) of the subjects had one of the two common genotypes (BB and BC), comparisons between the groups were made with these major genotypes and alleles. There were significant differences in the frequencies of the IL6vntr alleles and alleles between patients with Behçet’s disease and controls (genotypes: p = 0.005, pcorr = 0.01; alleles: p = 0.022, pcorr = 0.044) (table 1). The odds ratio for Behçet’s disease associated with the C allele of IL6vntr (IL6vntrC) was 3.5 (95% confidence interval 1.2 to 10.0).

When the studied subjects were stratified according to the results of HLA-B51 testing, significant differences in the IL6vntr genotype and allele frequencies were found only in the HLA-B51 negative subjects (genotypes: p = 0.005, pcorr = 0.01; alleles: p = 0.02, pcorr = 0.04). Using the EH programme, the distribution of haplotypes between patients with Behçet’s disease and controls differed significantly only in those with the IL6promG/IL6vntrC haplotype (p = 0.001, pcorr = 0.004): the odds ratio for Behçet’s disease in the subjects with this haplotype was 7.3 (95% confidence interval 1.6 to 32.9). In addition, the EH programme revealed a D value of 0.08, suggesting the presence of LD at low level between the two polymorphic sites.
No significant associations were found between the genotypes of the two IL 6 polymorphisms (IL6prom and IL6vntr) and clinical variables, including disease duration, mean age at onset, clinical manifestations, severe manifestations, and HLA-B51 positivity, in patients with Behçet’s disease (all \( p > 0.05 \)). However, the distribution of the IL6vntr genotype differed significantly between male and female patients, and the frequency of the BC genotype was much higher in female patients with Behçet’s disease than in male patients (BB: male, 45.5% \( \times \) female, 54.5%; BC: male, 8.3% \( \times \) female, 91.7%; \( p = 0.024 \), \( p_{corr} = 0.048 \)).

**DISCUSSION**

Our data are consistent with previous investigations showing considerable interethnic variability in the distribution of the IL6prom and IL6vntr genotypes.\(^1\)\(^2\)\(^3\)\(^4\)\(^5\) There was no evidence for genetic association conferred by the IL6prom polymorphism, whereas significant differences in the IL6vntr genotype and allele frequencies were found between patients with Behçet’s disease and controls. These differences were particularly apparent in the HLA-B51 negative subjects or female patients. In addition, susceptibility to Behçet’s disease was increased significantly in subjects carrying the IL6vntr\(^C\) allele and the IL6prom\(^G\)/IL6vntr\(^C\) haplotype. To confirm these findings, further investigations are required in other ethnic populations.

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### REFERENCES


