Leptin consumption in the inflamed joints of patients with rheumatoid arthritis

M Bokarewa, D Bokarew, O Hultgren, A Tarkowski

Background: Leptin has been shown to participate in bone remodelling and leptin substitution reported to have a protective effect in experimental septic arthritis.

Objective: To assess leptin levels in inflamed joints and plasma of patients with RA.

Material and methods: Leptin concentrations were assessed in matched blood and synovial fluid samples from 76 patients with RA. Blood samples from 34 healthy subjects acted as additional controls. Results were analysed and correlated with duration and activity of RA, x-ray changes, and treatment at time of sampling.

Results: In patients with RA, leptin levels were significantly higher in plasma than in synovial fluid samples obtained simultaneously and higher than in control samples. Plasma and synovial fluid leptin levels correlated strongly. Locally in the joint, leptin levels were related to WBC count. Such a relation was not seen in the bloodstream. Leptin levels were not related to sex, age, or disease duration. Difference between leptin levels in plasma and synovial fluid was greater in non-erosive arthritis (5.1 (SEM 1.2) v 3.7 (0.9) ng/ml, p=0.006), than in patients with erosive joint disease (6.2 (1.0) v 5.4 (0.8) ng/ml NS). Methotrexate treatment was associated with relatively high plasma leptin levels, while treatment with other DMARDs was associated with lower leptin levels than in patients receiving no DMARD treatment (p=0.0005).

Conclusions: Leptin production was significantly increased in patients with RA compared with healthy controls. Synovial fluid leptin levels were significantly lower than in matched plasma samples, suggesting an in situ consumption of this molecule.

Leptin is a cytokine-like 16 kDa peptide produced mostly by adipose tissue and regulating food intake, basal metabolism, and the β-oxidation of fatty acids. When binding to its receptor(s) located in hypothalamic nuclei occurs, leptin is an important trigger of adaptive mechanisms during starvation leading to down regulation of thyroid and reproductive functions and stimulation of hypothalamic. In healthy subjects, leptin levels in blood are proportional to the body fat mass. Leptin has recently been recognised as a modulator of inflammatory and immune responses by virtue of its interaction with leptin receptors expressed by peripheral blood mononuclear cells, vascular endothelial cells, smooth muscle cells, and osteoblasts. Interacting with its receptor in various tissues, leptin acts as a growth factor. Indeed, leptin facilitates proliferation of bone forming osteoblastic cells and osteosarcoma cells. Leptin facilitates proliferation of human endothelial cells supporting angiogenesis and neovascularisation. Leptin has a dual role in inflammation. On the one hand, it activates macrophages and potentiates production of the proinflammatory cytokines, tumour necrosis factor α (TNFα), interleukin (IL)6, and direct T cell differentiation to Th1 phenotype, expressing interferon γ and IL2. On the other hand, it expresses certain anti-inflammatory properties by releasing IL1 receptor antagonist. We have previously shown that supplementation with leptin alleviated the course of staphylococcal sepsis. In addition, subjects with higher plasma leptin levels had a better prognosis for survival of sepsis and septic shock.

In rheumatoid arthritis (RA) it has been suggested that hypothalamic-pituitary dysregulation is an important pathogenic mechanism. An increase in corticotrophin releasing hormone production and low levels of secreted cortisol seems to be crucial for the onset of RA. A regulatory loop exists between the hypothalamus-pituitary axis and levels of circulating leptin. Indeed, leptin administration during starvation reduces hypercortisolaemia, while treatment with glucocorticoids up regulates leptin levels. In addition, stimulation of hypothalamic centres by intracerebral injection of leptin diminishes response to proinflammatory stimuli exerted by lipopolysaccharide and TNFα. The role of leptin for the development of arthritis has been assessed in two experimental studies. Leptin deficient ob/ob mice were partly protected from antigen induced arthritis, because they developed less synovial tissue proliferation and less humoral response to the injected antigen. In contrast, leptin supplementation to outbred mice reduced the severity of joint destruction in septic arthritis.

In this study we show that leptin levels are increased in patients with RA compared with healthy controls. Moreover, leptin levels in synovial fluid are reduced as compared with matched plasma samples, suggesting local consumption of this molecule in the joint cavity.

PATIENTS AND METHODS

Patients

Plasma and synovial fluid samples were collected from 76 patients who attended the rheumatology clinics, at Sahlgrenska University Hospital in Göteborg, for acute joint effusion. RA was diagnosed according to the American College of Rheumatology criteria. At the time of synovial fluid and

Abbreviations: BSA-PBS, bovine serum albumin-phosphate buffered saline; CRP, C reactive protein; DMARDs, disease modifying antirheumatic drugs; ESR, erythrocyte sedimentation rate; GABA, γaminobutyric acid; IL, interleukin; MTX, methotrexate; RA, rheumatoid arthritis; TNFα, tumour necrosis factor α; WBC, white blood cell
blood sampling all the patients were receiving non-steroidal anti-inflammatory drugs. Disease modifying antirheumatic drugs (DMARDs) were used by 45 patients, of whom 28 used methotrexate (MTX) alone or in combination with other DMARDs (two with sulfasalazine, three with cyclosporin, five with TNFα inhibitors); five used sulfasalazine (one in combination with cyclosporin); six used parenteral or oral gold salt compounds; three used cyclosporin A (one in combination with etanercept); and the remaining patient received hydroxychloroquine. Six of the 45 patients were treated with azathioprine (one in combination with etanercept); and the remaining 31 patients had no DMARD treatment at the time of blood and synovial fluid sampling. Twenty four patients used oral glucocorticoids (2.5–10 mg/day, mean 6.5 mg), of whom 11 were also treated with MTX and six with other DMARDs. Seven patients received monotherapy with oral glucocorticosteroids.

Recent radiographs of the hands and feet were obtained for all the patients. The presence of bone erosions, defined as the loss of cortical definition at the joint, was recorded in proximal interphalangeal, metacarpophalangeal, carpus, wrist, and metatarsophalangeal joints. The presence of one erosion was sufficient to fulfill the requirement of an erosive disease. The presence of rheumatoid factor of any of the immunoglobulin isotypes was considered as positive.

Collection and preparation of samples
Synovial fluid was obtained by arthrocentesis, aseptically aspirated, and transmitted into sodium citrate (0.129 mol/l; pH 7.4) containing tubes. In most cases synovial fluid was obtained from knee joints. Three samples were from elbows and six samples were obtained from shoulder joints. Blood samples were simultaneously obtained from the cubital vein and directly transferred into sodium citrate medium. Blood samples from 34 healthy subjects (aged 18–67, mean 42 (SEM 7) years) were used as the control group. Collected blood and synovial fluid samples were centrifuged at 800 g for 15 minutes, aliquoted, frozen, and stored at −20°C until use.

Laboratory measures of disease activity
Serum levels of C reactive protein (CRP) were measured by standard nephelometry, with established normal range 0–5 mg/l. The erythrocyte sedimentation rate (ESR) was measured by the Westergren method having normal range of 0–20 mm/1st h. White blood cell (WBC) counts in blood and synovial fluid were obtained with a microcell counter F300 (Sysmex, Toa, Japan). Synovial fluid samples were treated with hyaluronidase before the cell count. The level of leptin in the blood and synovial fluid samples was determined by a sandwich enzyme linked immunosorbent assay (ELISA) using matched antibody pairs (R&D systems, Stockholm, Sweden). Briefly, 96 well polystyrene dishes (Nunc, Denmark) were coated with capture antibodies and left overnight at room temperature. After washing, the plates were blocked with bovine serum albumin-phosphate buffered saline (BSA-PBS) containing 5% sucrose. Matched samples of plasma and synovial fluid were introduced into the parallel strips, at a dilution of 1:10 in BSA-PBS. Horseradish peroxidase labelled detection antibodies and corresponding substrate were used for colour development. Double wave length reading at 450 and 570 nm was used, and the difference of absorbances was calculated. The absorbance values obtained were compared with serial dilution of recombinant leptin and presented as ng/ml.

Statistical analysis
Leptin levels were determined by a sandwich enzyme linked immunosorbent assay (ELISA) using matched antibody pairs (R&D systems, Stockholm, Sweden). Briefly, 96 well polystyrene dishes (Nunc, Denmark) were coated with capture antibodies and left overnight at room temperature. After washing, the plates were blocked with bovine serum albumin-phosphate buffered saline (BSA-PBS) containing 5% sucrose. Matched samples of plasma and synovial fluid were introduced into the parallel strips, at a dilution of 1:10 in BSA-PBS. Horseradish peroxidase labelled detection antibodies and corresponding substrate were used for colour development. Double wave length reading at 450 and 570 nm was used, and the difference of absorbances was calculated. The absorbance values obtained were compared with serial dilution of recombinant leptin and presented as ng/ml.

RESULTS
Clinical and demographic data of the patients at presentation
Table 1 presents the clinical and demographic data of the patient and control groups. Matched samples of synovial fluid and blood were collected from 76 patients with RA—49 patients had destructive joint disease (erosive RA), and the remaining 27 patients had no changes of the joints as judged by the recent radiological examination of the hands and feet (non-erosive RA). The patients with erosive RA were, as
Counts (Leptin levels in synovial fluid correlated with synovial WBC, the age or the sex of patients with RA and control subjects. p=0.0001). Leptin levels in plasma were not related either to

Expected, significantly more often positive for rheumatoid factor in blood (45/49 v 7/27, p<0.0001), had longer duration of joint disease (p=0.002), and were more often receiving MTX treatment (23/49 v 5/27, p=0.013) than the group with non-erosive RA. No difference in glucocorticoid treatment between the groups was seen. Although the group with erosive RA tended to be older, the patient groups and the controls did not differ significantly in age or sex.

**Increased levels of leptin in blood of patients with RA**

Plasma of patients with RA contained significantly higher leptin levels than that of the control group (5.8 (SEM 0.7) ng/ml v 2.0 (0.6) ng/ml, p=0.037) (table 2). There was a significant decrease of leptin levels in joint fluid as compared with plasma in the patient group studied (5.8 (0.7) ng/ml v 4.8 (0.5) ng/ml, p=0.019). Leptin levels in plasma and synovial fluid of patients with RA showed close correlation (r=0.86, p=0.0001). Leptin levels in plasma were not related either to the age or the sex of patients with RA and control subjects. Leptin levels in synovial fluid correlated with synovial WBC counts (r=0.47, p=0.021), whereas such a correlation was not found for the leptin levels in plasma. Leptin levels in plasma were not related to the levels of CRP.

**Correlation between leptin levels and clinical characteristics of patients with RA**

A comparison of leptin levels was made in patients with RA stratified according to the erosivity of joint disease (table 2). Leptin levels in the plasma of patients with erosive and non-erosive joint disease did not differ significantly (6.2 (1.0) v 5.1 (1.2), NS), whereas in synovial fluid the leptin levels were higher in patients with erosive RA than in those with non-erosive RA (5.4 (0.8) v 3.7 (0.9), respectively, p=0.047). Patients with non-erosive RA displayed a more pronounced difference between the levels of leptin in plasma and its relative reduction in synovial fluid samples (p=0.006), while in the group of patients with erosive RA the difference between leptin levels in plasma and synovial fluid was negligible.

*Leptin levels correlated with the duration of the joint disease (r=0.47, p=0.017). Both in plasma and synovial fluid samples, leptin levels gradually increased with the duration of RA, being highest in the patients with disease lasting over 20 years (plasma 8.4 (1.6) ng/ml, synovial fluid 6.6 (1.2) ng/ml). This increase of leptin levels was, however, not related to the age of patients (the mean age of the group of patients with RA of over 20 years’ duration was 59.9 (3.0) years).

**Influence of treatment on leptin levels**

For further analysis, patients were stratified according to their treatment at the time of sampling (table 3). Patients treated with DMARDs other than MTX had significantly lower leptin levels in plasma than those having no immune modulating treatment (3.4 (0.3) v 6.8 (1.2), p=0.0005). The difference between leptin levels in plasma and synovial fluid of patients receiving no DMARD at the time of sampling was significant (p=0.024), while leptin levels in plasma and synovial fluid of DMARD treated patients were similar. Interestingly, patients receiving MTX treatment had significantly higher plasma and synovial fluid leptin levels than the patients receiving other DMARDs (p=0.002). The clinical characteristics of the patients treated with MTX and with other DMARDs, such as the incidence of erosive disease and the levels of inflammation assessed as CRP level and WBC count, were similar and could not be the reason for the difference in leptin levels between the groups. To assess whether the level of folic acid might be responsible for the increase of leptin levels in patients treated with MTX, plasma folate levels in patients with RA and controls were compared (table 3). The folate levels were similar in patients with RA and healthy controls (patients with RA 22.6 (3.0) nmol/l, controls 23.2 (4.0) nmol/l). However, when patients with RA were separated into those treated and those not treated with DMARDs, the non-treated group had significantly lower plasma folate levels (14.2 (3.1) v 30.0 (4.3) nmol/l, p=0.003). The MTX treated group did not differ from patients treated with other DMARDs (28.5 (4.9) v 33.5 (9.5) nmol/l, NS).

### Table 2

<table>
<thead>
<tr>
<th></th>
<th>RA, erosive (n=49)</th>
<th>RA, non-erosive (n=27)</th>
<th>Healthy controls (n=34)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Leptin level (ng/ml)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Plasma</td>
<td>6.2 (1.0)</td>
<td>5.1 (1.2)</td>
<td>2.0 (0.6)*</td>
</tr>
<tr>
<td>Synovial fluid</td>
<td>5.4 (0.8)</td>
<td>3.7 (0.9)†</td>
<td></td>
</tr>
<tr>
<td>White blood cell count (x10³/ml)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Blood</td>
<td>8.1 (0.4)</td>
<td>7.5 (0.5)</td>
<td></td>
</tr>
<tr>
<td>Synovial fluid</td>
<td>13.3 (2.5)</td>
<td>8.4 (2.0)</td>
<td></td>
</tr>
<tr>
<td>C reactive protein (mg/ml)</td>
<td></td>
<td>45.4 (6.7)</td>
<td>35.9 (9.0)</td>
</tr>
</tbody>
</table>

*p=0.00019, patients with RA versus healthy controls; †p=0.006 plasma versus synovial fluid, non-erosive RA; N/A, not assessed.

### Table 3

<table>
<thead>
<tr>
<th></th>
<th>Treated with MTX (n=28)</th>
<th>Treated with other DMARD (n=17)</th>
<th>Non-treated (n=31)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Disease duration (years)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Erosive disease [No (%)]</td>
<td>13.9 (1.7)</td>
<td>8.8 (1.6)</td>
<td>11.5 (1.8)</td>
</tr>
<tr>
<td>Oral glucocorticoids [No (%)]</td>
<td>23 (82)</td>
<td>11 (65)</td>
<td>15 (48)</td>
</tr>
<tr>
<td>C reactive protein (mg/l)</td>
<td>46.3 (6.9)</td>
<td>49.8 (16.8)</td>
<td>36.0 (8.9)</td>
</tr>
<tr>
<td>Folic acid (nmol/l)</td>
<td>28.5 (4.9)</td>
<td>33.5 (9.5)*</td>
<td>14.2 (3.1)</td>
</tr>
<tr>
<td>Leptin levels (ng/ml)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Plasma</td>
<td>5.9 (1.2)*</td>
<td>3.4 (0.3)</td>
<td>6.8 (1.2)‡</td>
</tr>
<tr>
<td>Synovial fluid</td>
<td>5.1 (1.0)</td>
<td>3.5 (1.3)</td>
<td>5.1 (0.9)</td>
</tr>
</tbody>
</table>

*p=0.003 DMARD treated versus non-treated; †p=0.002 MTX treated versus other DMARD treated; ‡p=0.024 plasma versus synovial fluid, non-treated group.
DISCUSSION

Our study showed (a) significantly increased leptin levels in the plasma of patients with RA compared with healthy subjects and concurrently, (b) a significant decrease of synovial fluid leptin levels compared with matched plasma samples. Leptin levels in synovial fluid correlated with the cellularity of the synovial fluid samples, implying local uptake/degradation of leptin in arthritis. Interestingly, the most clear cut difference in leptin levels between plasma and synovial fluid was found in patients with RA with the non erosive joint disease. A positive association between the consumption of leptin in the synovial cavity and the absence of bone destruction may be viewed as leptin mediated down regulation of the erosive process in the joints.

In favour of this suggestion is the knowledge that leptin induces IL1 receptor antagonist production.11 Treatment of patients with RA with IL1 receptor antagonist has been recently proved to stop the joint destructive process.12 Additionally, chondrocytes and fibroblasts are the two cell types sensitive to leptin stimulation and responding with increased proliferation.13,14 On the other hand, incubation of cell cultures in TNFα or IL1β enriched media suppresses leptin production.25 Increased levels of these cytokines locally in the synovial fluid of patients with RA may down regulate local production of leptin inside the joint, aggravating leptin deficiency.

This study shows that leptin levels in plasma are influenced by the use of DMARDs. Indeed, high leptin levels were found in patients treated with MTX, whereas patients receiving other DMARDs had significantly lower leptin levels (table 3). High leptin levels were not dependent on the age, disease duration, or activity of RA (judged as levels of CRP and ESR). Two mechanisms of MTX mediated increase of leptin levels may be proposed. MTX is a folic acid antagonist, being recognised as one of the most efficient drugs against chronic destructive arthritides, which can prevent the development of cartilage degradation and joint destruction. MTX competes with folic acid for transport into the cell using the reduced folate carrier protein and for binding to dehydrofolate reductase, resulting in depletion of products for biosynthesis of thymidylate and purines.25 MTX is known to block γ-aminobutyric acid (GABA) and to affect neuronal activity.26,27 In contrast, leptin stimulates activity in promelanocortin/GABA neurons.28 Interestingly, treatment with another GABA antagonist, valproic acid, indirectly induces IL1 receptor antagonist production.21


ACKNOWLEDGMENTS

The study was approved by the ethics committee of Sahlgrenska Hospital.

The work was supported by the Göteborg Medical Society, Swedish Association against Rheumatism, King Gustaf V’s Foundation, Swedish Medical Research Council, Nanna Svartz’s Foundation, Börje Dahlin’s Foundation, National Inflammation Network, Lundberg Foundation, and the University of Göteborg.

Authors’ affiliations

MBokarewa, DBokarewa, OHultgren, ATarkowski, Department of Rheumatology and Inflammation Research, Sahlgrenska University Hospital, Göteborg, Sweden

REFERENCES

1 Considine RV, Sinha MK, Heiman ML, Kriauciunas A, Stephens TW, Nyce MR, et al. Serum immunoreactive lep-
12 Hultgren OH, Tarkowski A. Leptin in rheumatoid arthritis 955

http://ard.bmj.com/ on 12 September 2003. Downloaded from


http://ard.bmj.com/ on 12 September 2003. Downloaded from


