EXTENDED REPORT

Infectious CNS disease as a differential diagnosis in systemic rheumatic diseases: three case reports and a review of the literature

K Warnatz, H H Peter, M Schumacher, L Wiese, A Prasse, F Petschner, P Vaith, B Volk, S M Weiner

Background: Immunosuppressive treatment of rheumatic diseases may be associated with several opportunistic infections of the brain. The differentiation between primary central nervous system (CNS) involvement and CNS infection may be difficult, leading to delayed diagnosis.

Objective: To differentiate between CNS involvement and CNS infection in systemic rheumatic diseases.

Methods and results: Three patients with either longstanding or suspected systemic rheumatic diseases (systemic lupus erythematosus, Wegener’s granulomatosis, and cerebral vasculitis) who presented with various neuropsychiatric symptoms are described. All three patients were pretreated with different immunosuppressive drugs (leflunomide, methotrexate, cyclophosphamide) in combination with corticosteroids. Magnetic resonance imaging of the brain was suggestive of infectious disease, which was confirmed by cerebrospinal fluid analysis or stereotactic brain biopsy (progressive multifocal leukoencephalopathy [PML] in two and nocardiosis in one patient).

Discussion: More than 20 cases of PML or cerebral nocardiosis in patients receiving corticosteroids and cytotoxic drugs for rheumatic disease have been reported. The clinical aspects of opportunistic CNS infections and the role of brain imaging, cerebrospinal fluid analysis and stereotactic brain biopsy in the differential diagnosis are reviewed.

Central nervous system (CNS) involvement may become a severe complication of several autoimmune disorders. In systemic lupus erythematosus (SLE) between 18 and 67% of patients have CNS involvement.1-5 Symptoms include psychosis, mood disorders, seizures, acute confusional states, stroke, migraine, chorea, aseptic meningitis, transverse myelopathy, as well as subtle cognitive impairment.7 There are no specific laboratory or magnetic resonance imaging (MRI) findings, making a proper diagnosis often difficult. MRI may be negative despite overt neuropsychiatric symptoms.8-10 Cerebrospinal fluid (CSF) analyses may show mild lymphocytic pleocytosis, raised protein levels, and IgG indices as well as oligoclonal bands in 25–50% of cases.11 In up to 81% of the cases CNS lupus may occur without systemic SLE activity,12 often leading to a delayed diagnosis.

In Wegener’s granulomatosis (WG) cerebral involvement is rare (2–8%).13-15 Forty years ago a review by Drachman described three patterns of CNS involvement in WG: (a) vasculitic changes; (b) meningeal involvement due to adjacent granulomatous disease; and (c) isolated granulomatous meningocerebral lesions. In recent MRI studies Murphy et al reported about 30% meningeal thickening and contrast enhancement, 36% meningeal involvement by extracerebral granulomatous disease, 20% cerebral infarcts, and about 50% non-specific white matter lesions.16 The main clinical manifestations are headache, seizure, or loss of function owing to focal lesion and stroke. Usually the diagnosis of WG is well established before CNS involvement occurs.17 There are no specific findings in WG suggesting CNS involvement. CSF analysis often shows only a slight rise in protein and a mild lymphocytic pleocytosis. The diagnosis is made by clinical findings, MRI of the CNS, and histology.18

Primary angiitis of the CNS is a very rare disorder.19 The diagnosis is based on the presence of neurological dysfunction, angiographic features of vasculitis (vessel irregularity, aneurysms, stenosis) and, if possible, stereotactic biopsy.20-22 MRI findings include infarction and white matter lesions, but both are non-specific.23,24 The diagnosis of this entity remains problematic and requires the exclusion of more common conditions.25

Other autoimmune diseases with common CNS involvement include polyarteritis nodosa, antiphospholipid antibody syndrome, and Behçet’s disease.25-27

Chronic dysregulation of the immune system in conjunction with immunosuppressive treatment predisposes patients with systemic autoimmune diseases to infections. Therefore the new onset of CNS symptoms always leads to the suspicion of underlying infections in the differential diagnosis of these patients.

Several viruses, bacteria, and fungi can imitate the clinical and the radiological picture of CNS involvement of the primary autoimmune diseases. Therefore careful evaluation of the CNS manifestation is essential because the impact on the therapeutic decision is great. However, the diagnostic procedure is limited by time because in all patients with severe CNS involvement prompt treatment should be instituted in order to preserve the maximum of cerebral function. In this paper we report the cases of three patients with infectious CNS involvement imitating autoimmune disorders. We also review the literature and suggest a standard diagnostic procedure in

Abbreviations: ANA, antinuclear antibodies; ANCA, antineutrophil cytoplasmic antibodies; CMV, cytomegalovirus; CNS, central nervous system; CRP, C reactive protein; CSF, cerebrospinal fluid; EBV, Epstein-Barr virus; HIV, human immunodeficiency virus; HSV, herpes simplex virus; IV, intravenous; MRT, magnetic resonance imaging; PCR, polymerase chain reaction; PML, progressive multifocal leukoencephalopathy; SLE, systemic lupus erythematosus; VZV, varicella zoster virus; WG, Wegener’s granulomatosis
intensity on T-weighted imaging. CSF analysis showed a normal cell count, an increased total protein content (652 mg/l), an IgG index of 0.5, and positive oligoclonal bands. A stereotactic brain biopsy was performed that showed abnormal oligodendroglial cells, demyelination, and swollen astrocytes associated with inflammatory cell infiltrates, suggesting the diagnosis of progressive multifocal leukoencephalopathy (PML). The diagnosis was confirmed by electron microscopy (fig 1B) and the detection of JC virus DNA in the brain tissue (nested polymerase chain reaction (PCR)) and in CSF samples (quantitative PCR). Tests for human immunodeficiency virus (HIV), cytomegalovirus (CMV) antigen, CMV-RNA, and antitoxoplasma IgM antibodies were negative. Leflunomide was stopped and the drug elimination was hastened by a two week treatment with cholestyramine.

Within three weeks the neurological status worsened, leading to a progressive motor weakness of the right side, a central paresis of the facial nerve, and a central impairment of bladder function. An MRI follow up disclosed progressive brain lesions. Antiviral treatment was started with 5 mg/kg cidofovir every two weeks. The course of the disease was fluctuating with phases of progression followed by phases of clear improvement of the neurological deficits.

**Patient 2**

Four years before admission a 52 year old white man had been diagnosed with WG. After a new onset of haemoptysis, proteinuria, scleritis, and arthralgias, diagnosis was histologically proved by a necrotising, granulomatous vasculitis in a bronchial biopsy specimen and by a positive cANCA (titre 1/100, anti-PR3 positive). With oral cyclophosphamide (2 mg/kg body weight) the clinical course stabilised, except for a residual proteinuria of 0.5 g/day. After 29 months of cyclophosphamide treatment (total dose 95 g), treatment was changed to methotrexate. Two months before admission, a slight increase in arthralgias and fever had developed. Because of suspected WG relapse the treating family doctor increased the daily steroid dose to 50 mg prednisolone a day. The patient was then admitted to our hospital. Because physical examination, a computed tomography scan of the lungs, and laboratory investigations showed no signs of WG activity or of infection, the patient was discharged and tapering of prednisolone was recommended.

Three weeks later he was readmitted because of cough, fever, night sweats, and weight loss of 8 kg. Now, the C reactive protein (CRP) was raised (24 g/l) and the differential blood count showed a lymphocytopenia, with CD4+ T cell counts reduced to 0.2×10⁹/l. An initial chest x-ray examination, bronchoscopy, and blood cultures failed to identify any pathogens. Subsequently, the carbon monoxide diffusion capacity was decreased and a new chest x-ray examination led to a suspicion of interstitial pneumonia. A bronchoalveolar lavage was performed and showed a chronic inflammation with no signs of pulmonary haemorrhage. All bacterial cultures, PCR for tuberculosis, legionella, and chlamydia, and PCR for Herpes viruses (herpes simplex virus (HSV), CMV, Epstein-Barr virus (EBV)) were negative. Treatment with methotrexate (15 mg weekly) was stopped and a third class cephalosporin and clarithromycin were started. Because the pulmonary function deteriorated, clarithromycin was changed to intravenous co-trimoxazole for suspected *Pneumocystis carinii* pneumonia. Because blood cultures were positive with *Staphylococcus epidermidis*, cephalosporins were replaced by imipenem. With this regimen dyspnoea and CRP levels decreased slightly and improved further when three days later treatment was started with 50 mg of intravenous (IV) prednisolone/day. On the 6th day of this regimen the patient suddenly presented with a generalised seizure. An MRI scan showed numerous small hyperintense lesions disseminated all over the brain (fig 2A) with a homogeneously high signal intensity on diffusion weighted imaging.
weighted images (fig 2B). As serological tests for toxoplasma IgM, aspergillus, borrelia, varicella zoster virus (VZV) IgA, CMV IgM, HSV, HIV were still negative, a stereotactic brain biopsy was performed. Pathological examination showed non-specific inflammation, but microscopic evaluation suggested coryne-like bacteria; the culture confirmed *Nocardia farcinica*. Intravenous co-trimoxazole, the preferred treatment, was continued for 40 days and then switched to oral maintenance treatment. A control MRI scan of the brain showed a complete remission six months later and clinically there was no WG activity detectable despite discontinuation of methotrexate.

Unfortunately, one year later the patient presented with proteinuria (6 g/day) and histological signs of segmental glomerulonephritis. The patient was treated with mycophenolate (1000 mg/day) without recurrence of the nocardia infection during the three month follow up.

**Patient 3**

A 55 year old white man complained of difficulties in writing four months before admission. His previous medical history was unremarkable. As the following neurological examinations, including EEG and MRI of the brain, were without clearcut pathological findings, the initial stage of Parkinson’s disease was suspected. MRI of the brain was repeated six months later and clinically there was no recurrence of the nocardia infection during the three month follow up.

On admission an MRI scan of the brain showed multiple confluent non-enhancing lesions of the periventricular, subcortical, and deep white matter with discrete involvement of the cortex (fig 3A). The diffusion weighted MRI scan showed increased diffusion of the brain lesions (fig 3B). Further examinations showed no underlying autoimmune process (negative results for rheumatoid factor, antinuclear antibodies (ANA), antineutrophil cytoplasmic antibodies (ANCA), antiphospholipid antibodies) or embolic disease. Laboratory findings, including CRP values, were unremarkable with the exception of a steroid induced leucocytosis of 23.5×10^9/l (13% lymphocytes) and a mild hypogammaglobulinaemia (IgG 5.6 g/l, normal range 7–16; IgA 0.6 g/l, normal range 0.7–4). Because of the impression of an accelerated deterioration of the underlying disease cyclophosphamide pulse therapy was started. The patient felt some improvement and was discharged one week later to rehabilitation.

One month later, after the patient had received a second bolus of cyclophosphamide, he became febrile, somnolent, and tetraparetic. On readmission, an MRI scan of the brain showed progressive confluent non-enhancing subcortical white matter lesions. A spinal tab was repeated and the CSF was positive for JC viral DNA. Before treatment could be applied he died with signs of septic shock.
### Table 1 Clinical presentation of primary cerebral involvement of rheumatic diseases and opportunistic CNS infections

<table>
<thead>
<tr>
<th>Clinical presentation</th>
<th>NPSLE</th>
<th>Primary CNS vasculitis</th>
<th>Wegener’s granulomatosis</th>
<th>Neuro-Behçet</th>
<th>Neurosarcoidosis</th>
<th>PML</th>
<th>HSV encephalitis</th>
<th>Bacterial meningitis</th>
<th>Brain abscess</th>
</tr>
</thead>
<tbody>
<tr>
<td>Onset of CNS disease</td>
<td>Acute, subacute, chronic</td>
<td>Acute, subacute, chronic</td>
<td>Acute, subacute, chronic</td>
<td>Acute, subacute, chronic</td>
<td>Subacute (weeks)</td>
<td>Acute, subacute (days-weeks)</td>
<td>Acute (days)</td>
<td>Subacute (days-weeks)</td>
<td></td>
</tr>
<tr>
<td>Seizures</td>
<td>13–35%</td>
<td>15–20%</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>5–10%</td>
<td>10%</td>
<td>5%</td>
</tr>
<tr>
<td>Headache</td>
<td>34–57%</td>
<td>30–64%</td>
<td>Frequent</td>
<td>95%</td>
<td>30%</td>
<td>5%</td>
<td>Frequent</td>
<td>90%</td>
<td>70–90%</td>
</tr>
<tr>
<td>Motor weakness/paresis</td>
<td>20%</td>
<td>50%</td>
<td>?</td>
<td>10–21% Paresis, 24% pyramidal signs</td>
<td>5–10%</td>
<td>33%</td>
<td>Frequent</td>
<td>10–15%</td>
<td>20–50%</td>
</tr>
<tr>
<td>Impaired consciousness</td>
<td>5–10%</td>
<td>29%</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>1%</td>
<td>Frequent</td>
<td>30–50%</td>
</tr>
<tr>
<td>Cognitive disorders</td>
<td>12–86%</td>
<td>40–50%</td>
<td>Rare</td>
<td>88%</td>
<td>10%</td>
<td>36%</td>
<td>Frequent</td>
<td>Frequent</td>
<td>?</td>
</tr>
<tr>
<td>Psychosis</td>
<td>4–6%</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Frequent</td>
<td>Rare</td>
<td>Rare</td>
</tr>
<tr>
<td>Mood disorder</td>
<td>10–30%</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Reactive</td>
<td>Rare</td>
<td>Rare</td>
</tr>
<tr>
<td>Visual deficits</td>
<td>5% (vascular, neuropathy, amaurosis fugax)</td>
<td>10–15%</td>
<td>Frequent (cranial neuropathy, ocular motor deficits)</td>
<td>1% Optic neuropathy</td>
<td>5–38% Optic neuritis</td>
<td>35% e.g. homonymous hemianopia</td>
<td>Frequent</td>
<td>Rare</td>
<td>Hemianopia</td>
</tr>
<tr>
<td>Sensory deficits</td>
<td>6–20%</td>
<td>15–20%</td>
<td>?</td>
<td>Up to 27%</td>
<td>5–10%</td>
<td>17%</td>
<td>Frequent</td>
<td>Rare</td>
<td>30%</td>
</tr>
<tr>
<td>Cerebellar disorders</td>
<td>Rare (infection)</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>10 (N. III, VI, VII)</td>
<td>Rare</td>
<td></td>
</tr>
<tr>
<td>Cranial nerve involvement</td>
<td>5–35%</td>
<td>Frequent</td>
<td>25% Ophthalmoplegia, 10–15% bulbar paresis</td>
<td>50–72% Cranial nerve palsy</td>
<td>Rare</td>
<td>Rare</td>
<td>10 (N. III, VI, VII)</td>
<td>Rare</td>
<td></td>
</tr>
<tr>
<td>CRP</td>
<td>Normal</td>
<td>Normal</td>
<td>Frequently</td>
<td>Frequently</td>
<td>Normal</td>
<td>Normal</td>
<td>normal</td>
<td>normal</td>
<td>↑</td>
</tr>
<tr>
<td>Other</td>
<td>–</td>
<td>1% Transverse myelitis, 2–3% chorea</td>
<td>30% Aphasias</td>
<td>Chronic meningitis, myelopathy</td>
<td>8% Meningitis, encephalitis, 6% movement disorders</td>
<td>S-12% Meningitis, 10–28% myelitis, 10% hypothalamic and pituitary dysfunction</td>
<td>Impaired speech</td>
<td>Wernicke aphasia, dysphasia</td>
<td>Meningism, fever, arena, 50% Fever, 25–30% meningitis</td>
</tr>
</tbody>
</table>

NPSLE, neuropsychiatric systemic lupus erythematosus; PML, progressive multifocal leucoencephalopathy; HSV, herpes simplex virus; ? no data from clinical trials available.

### Table 2 MRI findings in primary cerebral involvement of rheumatic diseases and opportunistic CNS infection

<table>
<thead>
<tr>
<th>MRI findings</th>
<th>NPSLE</th>
<th>Primary CNS vasculitis</th>
<th>Wegener’s granulomatosis</th>
<th>Neuro-Behçet</th>
<th>Neurosarcoidosis</th>
<th>PML</th>
<th>HSV encephalitis</th>
<th>Bacterial meningitis</th>
<th>Brain abscess</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>13–50%</td>
<td>0–50%</td>
<td>50%</td>
<td>30%</td>
<td>11%</td>
<td>Only in the early phase</td>
<td>Rare</td>
<td>?</td>
<td>Rare</td>
</tr>
<tr>
<td>Territorial infarction</td>
<td>15–30%</td>
<td>Frequently (e.g. middle cerebral artery)</td>
<td>15–20%</td>
<td>18%</td>
<td>Rare</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td></td>
</tr>
<tr>
<td>In T2 hyperintense lesions of the cortex</td>
<td>0–9% Reversible</td>
<td>Rare</td>
<td>Cerebral granulomas (homogeneous, ring enhancement)</td>
<td>36–66% Met. diencephalic, 26% isolated brain stem or basal ganglia</td>
<td>30% Multiple or solitary suprasphenoidal and/or infratentorial, rarely brain stem or cerebellum</td>
<td>56% Thalamus, 32% posterior fossa</td>
<td>Haemorrh. necrosis, temporal, insula, thalamus, hippoc., subfrontal</td>
<td>–</td>
<td>Frequent</td>
</tr>
<tr>
<td>White matter lesions (WML)</td>
<td>30–75% Subcortical &gt; deep white matter &gt; periventricular</td>
<td>Frequent infarcts often in the deep white matter</td>
<td>50% Periventricular, subcortical</td>
<td>16% Para- and periventricular</td>
<td>40% Peri- and periventricular, subcortical, confluent</td>
<td>100% Subcortical, 93% parieto-occipital, 92% bilateral, 94% confluent, no mass effect</td>
<td>–</td>
<td>–</td>
<td>Frequent, capsule with low intensity signal, indistinct margins between abscess and surrounding</td>
</tr>
<tr>
<td>Gd-DTPA enhancement of WML or cortical lesions</td>
<td>Only active lesions</td>
<td>Active lesions</td>
<td>Active lesions</td>
<td>Often but only active lesions</td>
<td>Often nodular or annular enhancement</td>
<td>&lt;10% Enhancement of the periphery</td>
<td>Often</td>
<td>–</td>
<td>Strong contrast enhancement</td>
</tr>
<tr>
<td>Atrophy</td>
<td>10–60%</td>
<td>Chronic stage</td>
<td>42%</td>
<td>20% Brain stem</td>
<td>38–57% Nodular or diffuse meningeal enhancement, 28% optic nerve enhancement</td>
<td>58% Infratentorial (e.g. brain stem), no perifocal oedema</td>
<td>Often brain oedema associated with the lesions</td>
<td>Meningeal enhancement, nodular lesions</td>
<td>Mass effect</td>
</tr>
<tr>
<td>Other</td>
<td>Rarely meningeal enhancement</td>
<td>2–30% Focal or diffuse meningeal/dural thickening</td>
<td>38–57% Nodular or diffuse meningeal/dural thickening</td>
<td>20% Brain stem</td>
<td>38–57% Nodular or diffuse meningeal/dural thickening, 28% optic nerve enhancement</td>
<td>58% Infratentorial (e.g. brain stem), no perifocal oedema</td>
<td>Often brain oedema associated with the lesions</td>
<td>Meningeal enhancement, nodular lesions</td>
<td>Mass effect</td>
</tr>
</tbody>
</table>

Gd-DTPA, gadolinium-DTPA.
### Table 3 Cerebrospinal fluid (CSF) findings in primary cerebral involvement of rheumatic diseases and opportunistic CNS infections

<table>
<thead>
<tr>
<th>Condition</th>
<th>Normal findings</th>
<th>Pleocytosis</th>
<th>Raised protein content</th>
<th>CSF ACE</th>
<th>CSF IgG index</th>
<th>IgG index</th>
<th>Raised JCV PCR</th>
<th>Raised HSV PCR</th>
<th>Raised PML PCR</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wegener's granulomatosis</td>
<td>10–50%</td>
<td>25–50%</td>
<td>300–1000</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
</tr>
<tr>
<td>Neuro-Behçet</td>
<td>10–50%</td>
<td>25–50%</td>
<td>300–1000</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
</tr>
<tr>
<td>Neurosarcoiosis</td>
<td>10–50%</td>
<td>25–50%</td>
<td>300–1000</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
</tr>
<tr>
<td>PML</td>
<td>10–50%</td>
<td>25–50%</td>
<td>300–1000</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
</tr>
<tr>
<td>HSV encephalitis</td>
<td>10–50%</td>
<td>25–50%</td>
<td>300–1000</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
</tr>
<tr>
<td>Bacterial meningitis</td>
<td>10–50%</td>
<td>25–50%</td>
<td>300–1000</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
</tr>
<tr>
<td>Brain abscess</td>
<td>10–50%</td>
<td>25–50%</td>
<td>300–1000</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
<td>Rare</td>
</tr>
</tbody>
</table>

**DISCUSSION**

In recent years morbidity and mortality of autoimmune diseases—partly due to effective immunosuppressive treatment—are increasingly related to secondary infections. Cerebral involvement, especially, makes differential diagnosis difficult. Often the clinical presentation does not allow discernment between primary angiitis of the CNS, secondary CNS involvement of rheumatic diseases, and CNS infection, because the signs and symptoms are non-specific (table 1). Also, MR images (table 2) and laboratory findings, including CSF analysis (table 3), are rarely specific. Only a combination of several diagnostic procedures, additional specific serological tests, and, if possible, stereotactic brain biopsy permit a firm diagnosis.

Here we review the current literature on PML and necrotising in systemic rheumatic diseases and present a diagnostic algorithm for immunosuppressed patients with new onset or worsening of neuropsychiatric symptoms.

In the past 10 years JC virus has been recognised as an important pathogen in patients receiving immunosuppressive treatment. It is mainly associated with advanced HIV infection and causes PML, a fatal demyelinating JC virus induced disorder of the CNS. Reports on at least 20 cases of patients with PML receiving corticosteroids and cytotoxic drugs for rheumatic disease have been published: 11 patients with WG, three patients with rheumatoid arthritis, three patients with MS, two patients with inflammatory myositis, and one patient with mixed connective tissue disease.

Among healthy volunteers, 65–90% have antibodies against, and are carriers of, the JC virus. One study demonstrated a correlation between JC viruria and treatment with corticosteroids and cytotoxic drugs, whereas others found no relationship. The development of PML does not seem to depend on distinct immunosuppressant drugs: corticosteroids, cyclophosphamide, chlorambucil, azathioprine, cyclosporin A, and leflunomide (patient No 1) have all been associated with the occurrence of PML in case reports.

Low CD4+ T cell counts may predispose for opportunistic CNS infections such as cerebral toxoplasmosis. However, CD4+ T cell depletion alone is not sufficient to define a high risk of PML. Indeed, experience with HIV infection showed that 11% of patients with a clinical manifestation of PML have a CD4+ cell count above 0.2 × 10⁹/l, whereas others found no relationship. The development of PML does not seem to depend on distinct immunosuppressant drugs: corticosteroids, cyclophosphamide, chlorambucil, azathioprine, cyclosporin A, and leflunomide (patient No 1) have all been associated with the occurrence of PML in case reports.

Low CD4+ T cell counts may predispose for opportunistic CNS infections such as cerebral toxoplasmosis. However, CD4+ T cell depletion alone is not sufficient to define a high risk of PML. Indeed, experience with HIV infection showed that 11% of patients with a clinical manifestation of PML have a CD4+ cell count above 0.2 × 10⁹/l, whereas others found no relationship.

**Table 1 summarises the clinical manifestations of PML.**

Abnormalities of the motor function are most common. Fever, headache, neck stiffness, or impaired consciousness are rare. PML lesions begin as small foci in the white matter and expand concentrically, either at one or several sites. In later stages of the disease MR images are strongly suggestive of the diagnosis. Typically, PML appears as bilateral, asymmetrically distributed, confluent (>90%), predominantly subcortical white matter lesions which develop close to the grey-white matter junction and in the periventricular region. They show high signal intensity on T2 weighted imaging and low signal intensity on T1 weighted series. The grey matter, especially the thalamus, can be affected in up to 50% of patients (table 2).

Usually, lesions of PML are not enhanced by gadolinium-DTPA. Rarely, a faint peripheral enhancement is seen, which
is interpreted as an indicator of an immune response to viral antigens in long term survivors.\textsuperscript{55,56} Despite a rather characteristic presentation of advanced PML in MRI analysis the differential diagnosis remains a challenge; because cases of central nervous system SLE mimicking PML have been reported.\textsuperscript{57} Therefore, the next step in confirming a suspected diagnosis of PML must be analysis of the CSF for the presence of JC virus DNA by PCR. This test reaches a sensitivity of 93% and a specificity of 99% while PCR analysis performed in urine or blood samples was not specific for PML.\textsuperscript{58} Whether the viral load correlates with the prognosis is still being debated. The firm diagnosis—unfortunately often post mortem—is made by histological examination, showing enlarged oligodendrocytes with an expanded cytoplasmic compartment and intranuclear inclusions ("ground glass"), which represent the polyoma viruses. Therefore, stereotactic brain biopsy is strongly recommended in all cases of suspected PML and negative JC virus PCR in CSF samples.

The primary differential diagnoses include other viral infections (HIV, HSV, CMV), toxoplasmosis, neuropsychiatric SLE (in patients with known SLE), lymphoma, toxic encephalitis after chemotherapy, vasculitis, and neuroarosarcoisitis (tables 2 and 3). Interestingly, the vascular lesions shown by cerebral angiography in patient No 3 strongly argue against PML as the cause of the primary symptoms in this patient. On the other hand, it has to be kept in mind, that several infectious agents have to be ruled out in the assessment of patients with possible cerebral vasculitis, because a variety of pathogens have a propensity to affect blood vessels—for example, aspergillus, candidiasis, cisticercosis, coccidiodomyositis, 	extit{Strongyloides stercoralis}, arbovirus, VZV, and hepatitis C virus infection.\textsuperscript{59,60}

The prognosis for PML is usually poor. No effective treatment is available at present; anecdotal reports show some efficacy of cidofovir, interferon alfa, and cytosine arabinoside.\textsuperscript{61} Nevertheless, PML may remit if the underlying immunodeficiency improves,\textsuperscript{62} therefore discontinuation of immunosuppressive agents should receive a high priority.\textsuperscript{63}

MRI findings of early white matter lesions caused by JC virus are non-specific and may be indistinguishable from early brain abscesses. However, contrast enhancement or mass effects of the lesions visualised by MRI helps to distinguish cerebral abscesses from PML and HSV encephalitis. Differential diagnosis includes primary brain tumours, metastasis, or cerebrovascular events.

Symptoms in patients with brain abscess are headache, fever, focal neurological deficits, confusion, meninigitis and seizures, all of which may also occur in patients with cerebral metastasis or CNS involvement due to ANA or ANCA positive vasculitis. However, fever and meningitits are rarely seen in systemic rheumatic diseases, with the exception of WG.\textsuperscript{64} If a spinal tap is contraindicated owing to the mass effect of the brain abscess, the preferred diagnostic procedures are stereotactic brain biopsy followed by histological and microbiological analysis.

Typical microbes which can cause brain abscess formation are 	extit{Toxoplasma gondii}, fungi (aspergillus, candida, or cryptoccoci), mycobacteriosis, 	extit{Listeria monocytogenes}, and 	extit{Nocardia asteroides}. Thirty two cases of SLE associated nocardiosis have been reported.\textsuperscript{65} Lungs, skin, and brain were the organs most commonly affected. Nocardiosis of the CNS was found in up to 30% of these patients and was associated with a high mortality.\textsuperscript{66} Nocardia should be kept in mind as a possible pathogen in patients whose infections do not respond to third
authors’ affiliations

K Warnatz, H H Peter, F Petschner, P Vaith, S M Weiner, Department of Rheumatology and Clinical Immunology, Medizinische Klinik, University Hospital, Freiburg, Hugstetter Strasse 55, 79106 Freiburg, Germany

M Schumacher, Department of Neuroradiology, University Hospital, Freiburg

L Wiese, Department of Neurology, University Hospital, Freiburg

A Prasse, Department of Pulmonary Medicine, University Hospital, Freiburg

B Voll, Department of Neuropathology, University Hospital, Freiburg

References


Infectious CNS disease in systemic rheumatic diseases

57


56 Pfister HW, Echemoid D. Infectious inflammatory diseases of the central nervous system from the neurological and internal medicine viewpoint. Internist (Berl) 2001;42:991–8.


