EXTENDED REPORT

Investigation of chromosome 2q in osteoarthritis of the hand: no significant linkage in a Tasmanian population


Background: Previous studies have suggested a strong genetic component to osteoarthritis (OA), especially that of the hand, and three linkage studies have suggested the existence of susceptibility loci in disparate regions of chromosome 2q.

Objective: To examine for linkage to 2q in a Tasmanian population of women and men with familial hand OA.

Methods: Hand OA (distal interphalangeal, carpometacarpal, and Heberden’s nodes) was assessed by a combination of hand photographs and radiographs. A non-parametric linkage (NPL) analysis was performed on chromosome 2q of 69 members in 22 families with severe distal interphalangeal joint OA using Genehunter. A quantitative trait linkage analysis of a larger group of 456 members in 68 families was also performed using SOLAR.

Results: The maximum non-parametric linkage score was 1.05 (p=0.15) at marker IL1R1, close to the centromere. All components of hand OA scores had significant heritability in this dataset (28%-35%, all p<0.001). Despite this, the quantitative trait analysis (after adjustment for age and, where appropriate, sex) yielded maximum LOD scores of 0.90 for Heberden’s nodes (both sexes combined), and 1.19 for carpometacarpal OA score (women only).

Conclusions: These results do not provide confirmation of linkage on chromosome 2q in the larger white population with hand OA. They suggest that there are regional variations in the genetic cause of hand OA and that other loci not on 2q may be important in this disease.

Osteoarthritis (OA) is the leading cause of musculoskeletal disability in western countries. Although knee and hip OA cause the greatest disability, hand OA also results in disability. The prevalence of hand OA in the community is very high, with population based estimates ranging from 30% to 52%, and is strongly age dependent, ranging in one survey from 5% in subjects aged 18–39 to 72% in subjects aged 60 and over.

Some studies have suggested that both genetic and environmental factors contribute to the development of OA, with different risk factors at different joints, and different risk factors for men and women. There is recent evidence to suggest that the heritability of hand OA and hip OA may be higher than the heritability of knee OA. In addition, some reports suggest that the genetic component of OA may be greater for women than for men.

Three previous studies have identified chromosome 2q as one of the regions of the human genome most likely to harbour genes for OA, although at divergent regions of this chromatin. Two of these linkage studies investigated hand OA, whereas the third was a study of hip and knee OA.

Taken together, these three studies suggest that there may be OA susceptibility genes on chromosome 2q. The aim of this study, therefore, was to examine for the presence of linkage to 2q in a Tasmanian population of men and women with familial hand OA.

METHODS

Tasmania is an island state of Australia. Its population of 472,000 is predominantly white, and is similar genetically to other Anglo-Celtic populations, although it is more homogeneous than some other populations. At the time of the study, statewide rheumatology specialist services in Tasmania were provided by three rheumatologists in a single practice based in the capital city of Hobart. Subjects were recruited from the records of this practice. All patients who had a doctor’s diagnosis of hand OA and at least one living relative with hand OA were invited to take part, along with their affected and unaffected relatives. Ethical approval was granted for the study by the Royal Hobart Hospital human research ethics committee and all participants provided informed written consent. Participants underwent a comprehensive protocol involving collection of blood, as well as detailed assessment of hand OA, anthropometrics, environmental factors, pain, and function.

Presence of hand OA was assessed using two methods. Radiographic disease was assessed using an atlas for joint space narrowing and osteophytes at the distal interphalangeal (DIP) and first carpometacarpal (CMC) joints from a single anteroposterior radiograph of the hands, performed according to a standardised protocol by the same two investigators (HMC, GJ) simultaneously, with a single score obtained by consensus. The scores for each component at each joint could vary from 0 (no disease) to 3 (severe disease). Scores for DIP disease could vary from 0–48 and CMC disease from 0–12.

A photograph of both hands was taken with an Elicar Medical Macro MS2 camera. Photographs were then scored as to the presence (1) or absence (0) of Heberden’s nodes in each DIP joint by two investigators (GJ, HMC) simultaneously, with a single score obtained by consensus without reference to radiographs. Heberden’s node scores could vary from 0–8.

The reproducibility of all measures was assessed by resoring widely varying samples of 45 radiographs and 50

Abbreviations: cM, centimorgans; CMC, first carpometacarpal; DIP, distal interphalangeal; MLS, multipoint LOD score; NPL, non-parametric linkage; OA, osteoarthritis
photographs one week after they were first scored. All intra-class correlations were at least 0.94. All phenotyped subjects, plus an additional 77 spouses, were genotyped using fluorescent automated fragment analysis technology (Applied Biosystems ABI PRISM 377 DNA Sequencers) at the Australian Genome Research Facility, Melbourne, using 19 microsatellite markers from chromosome 2q. These included the 16 markers for 2q from Applied Biosystems ABI PRISM Linkage Mapping set MD-10, and three additional microsatellite markers (IL1R1, D2S2335, and GCG) reported to be in linked regions in previous studies. Genetic distances between the ABI markers were taken from the Marshfield chromosome 2 sex averaged linkage map (http://research.marshfieldclinic.org/genetics/), and the positions of the additional markers were estimated using the same map in combination with the orderings of Whitehead contigs WC2.8 and WC2.11 (accessed via the Genome Database, http://www.gdb.org), physical map distances from the genome browser of the National Center for Biotechnology Information (http://www.ncbi.nlm.nih.gov), and figure 2 in a previous OA linkage study. Pedcheck was used to detect genotyping and pedigree errors.

Both dichotomous trait and quantitative trait analyses were used to test for linkage. For a dichotomous trait, affected patients only analysis, families and individual members were only included if they satisfied identical severity and similar age criteria to those used in a previous Finnish study. This was done so that results between the two studies could be directly compared. Families were included if they contained a proband 55 years old or younger with at least one non-zero Altman atlas DIP joint osteophyte score in each hand, and a DIP joint osteophyte score of 2 or 3 in at least one joint (equivalent to third or fourth degree radiographic OA respectively on the Kellgren-Lawrence scale used in the Finnish study). Relatives were classed as “affected” either if they were the sibling of a proband and had at least one non-zero DIP joint osteophyte score in each hand (at least second degree radiographic OA on the Kellgren-Lawrence scale), or if they were 55 or younger and had at least one non-zero DIP joint osteophyte score in each hand. Non-parametric linkage (NPL) scores were calculated using Genehunter 2 with the Spairs statistic, and Genehunter-Plus was used to calculate the less conservative Kong and Cox z score allowing for incomplete inheritance information. Non-parametric scores and z scores from each pedigree were combined by using the default weighting with scores from all pedigrees weighted equally. All pedigrees were small enough to include all affected persons in a full multipoint analysis. Of the unaffected relatives and older parents who had been genotyped, as many as possible were included in the multipoint analysis (within the constraints on pedigree size imposed by Genehunter) to improve the accuracy of haplotype reconstruction.

Several parametric analyses were also performed using the same criteria outlined above to define affected subjects. For these analyses the remaining subjects had to be classed as unaffected or unknown. Subjects 50 or older without DIP joint osteophytes in both hands were classed as unaffected; all other subjects were classed as unknown (those younger than 50 with insufficient abnormalities to be classed as affected, and those older than 55 with sufficient abnormalities to prevent them being classed as unaffected). For the parametric calculations, Genehunter 2 was used for a multipoint analysis and extra unaffected relatives and older parents were included in two point analyses with Fastlink version 4.1.
A multipoint variance components analysis—testing for the presence of an additive quantitative trait locus on 2q—was carried out using SOLAR. Three trait scores were analysed: (a) the DIP joint radiographic score (possible range 0–48), (b) the CMC joint score (0–12), and (c) the Heberden’s node score (0–8). Each trait was analysed for all subjects combined, as well as for women and men separately. The software was first used to estimate heritability after adjusting for the covariates age, sex, and an age-sex interaction, and correcting for ascertainment by conditioning on the probands. Age stratification was not performed. Evidence for linkage was then assessed at various points on the chromosome by testing the null hypothesis that the additive genetic variance due to a quantitative trait locus at each point was zero. This test for linkage assumes that trait scores are normally distributed. However, simulations have suggested that non-normal trait distributions with a coefficient of kurtosis less than 2 do not yield excessive type I error rates.

### RESULTS

A total of 7116 genotypes were measured. Of these, 82 (1.2%) were eliminated after checking for inheritance errors using Pedcheck.

Sixty nine subjects in 22 families were classed as affected using the dichotomous trait definition. The three largest families contained 10, seven, and five affected members; the other 19 families contained two to four affected members. There were 67 affected sibling pairs (38 independent affected sibling pairs), two affected avuncular pairs, and 39 affected cousin pairs. The affected subjects (44 women and 25 men) had a mean age of 53.2 (SD 6.2) years and ranged in age from 34 to 67 years. Twenty one of the affected subjects were graded with a maximum Altman DIP osteophyte score of 1 in any joint—equivalent to second degree radiographic OA on the Kellgren-Lawrence scale. Of the remaining subjects, 29 had a maximum Altman score of 2, and 19 had a maximum score of 3.

The maximum NPL score was 1.05 (p=0.15) at marker IL1R1, close to the centromere (fig 1). The Kong and Cox z score reached a maximum of 1.81 (p=0.035) at the same marker. There was a similar lack of evidence for linkage when several dominant and recessive parametric models were tested, using both Fastlink and Genehunter (including the dominant model for which suggestive linkage was reported in the Finnish study; results not shown).

For the quantitative trait analysis, phenotypic data were available for a total of 456 members (295 women and 161 men) in 68 families, with two to 21 members per family. Some characteristics of these members are given in table 1. They ranged in age from 24 to 92 years. Radiographic DIP joint scores, CMC joint scores, and Heberden’s node scores were non-zero for 63%, 53%, and 51% of members respectively and ranged from 0 to their maximum values (48, 12, and 8 respectively). Spearman’s correlation was 0.73 between radiographic DIP scores and Heberden’s node scores, 0.64 between radiographic DIP scores and CMC scores, and 0.50 between CMC scores and Heberden’s node scores (all p values<0.001). All scores were strongly associated with age and sex: age and sex and their interaction term explained 43% of the variance in DIP scores, 38% of the variance in CMC scores, and 31% of the variance in Heberden’s node scores.

After adjusting for these covariates, the estimated heritabilities of the DIP, CMC, and Heberden’s node scores were 35%, 35%, and 28% respectively, and all differed significantly from zero (p<0.001). The coefficients of kurtosis for the covariate adjusted scores were all less than 0.7.

There was no evidence of an additive quantitative trait analysis on 2q influencing any of the three traits (table 2). When women and men were analysed together, the maximum LOD score was 0.90 for Heberden’s nodes at a locus 140 cM distal to IL1R1. When women and men were analysed separately, the maximum LOD score across 2q for any analysis was 1.19 for CMC joints in women.

### DISCUSSION

Despite a comprehensive approach with high quality phenotypic data, there was no evidence of linkage on chromosome 2q in the Finnish population to support previous linkage findings in other populations.

In an analysis of 99 siblings from 44 families in England affected with nodal OA, two point linkage (pointwise p values<0.05) was reported for three microsatellite markers spanning approximately 70 cM in the regions 2q23–q35 in the middle of 2q. In a Finnish study, a genomewide scan was conducted on 64 siblings from 27 sibships with severe osteoarthritis in their distal interphalangeal joints. Suggestive linkage was reported for 2q, very close to the centromere (two point LOD score 2.34, p=5.1×10−4). A second linkage peak was detected some 20 cM distal (two point LOD score 1.48; p=4.5×10−4). Multipoint analysis yielded p values of 1×10−5 and 7×10−4 respectively at these two loci.

### Table 1

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Mean (SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>55.2 (14.7)</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>164.7 (9.9)</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>74.6 (16.2)</td>
</tr>
<tr>
<td>Distal interphalangeal joint score</td>
<td>12.0 (13.4)</td>
</tr>
<tr>
<td>Carpometacarpal joint score</td>
<td>3.1 (3.6)</td>
</tr>
<tr>
<td>Heberden’s node score</td>
<td>2.7 (3.0)</td>
</tr>
</tbody>
</table>

### Table 2

<table>
<thead>
<tr>
<th>Subjects</th>
<th>Quantitative trait (maximum value)</th>
<th>Maximum multipoint LOD score* (locus [cM])</th>
</tr>
</thead>
<tbody>
<tr>
<td>All (n=456)</td>
<td>Distal interphalangeal radiographic score (48)</td>
<td>0.80 (140)</td>
</tr>
<tr>
<td></td>
<td>Carpometacarpal radiographic score (12)</td>
<td>0.61 (146)</td>
</tr>
<tr>
<td></td>
<td>Heberden’s node score (8)</td>
<td>0.90 (140)</td>
</tr>
<tr>
<td>Women (n=295)</td>
<td>Distal interphalangeal radiographic score (48)</td>
<td>0.66 (136)</td>
</tr>
<tr>
<td></td>
<td>Carpometacarpal radiographic score (12)</td>
<td>1.19 (126)</td>
</tr>
<tr>
<td></td>
<td>Heberden’s node score (8)</td>
<td>0.81 (136)</td>
</tr>
<tr>
<td>Men (n=161)</td>
<td>Distal interphalangeal radiographic score (48)</td>
<td>0.15 (35)</td>
</tr>
<tr>
<td></td>
<td>Carpometacarpal radiographic score (12)</td>
<td>0.04 (105)</td>
</tr>
<tr>
<td></td>
<td>Heberden’s node score (8)</td>
<td>0.40 (90)</td>
</tr>
</tbody>
</table>

*Distances in centiMorgans (cM) measured distal to marker IL1R1.
In a United Kingdom linkage study of knee and hip OA\(^6\) a genomewide scan was conducted on a much larger set of 481 families, each containing at least one sibling pair who had had hip or knee joint replacements. A multipoint LOD score (MLS) of 1.22 was reported at a locus between the linkage sites on 2q reported in the English nodal OA study.\(^{11}\) This MLS increased to 2.19 (p = 7.4 x 10\(^{-7}\)) in the families concordant for hip only disease.

For our dichotomous trait analysis, we had a similar number of affected people, 69, as in the Finnish study,\(^7\) which had 64, and a similar number of affected sibling pairs (67 v 50). The same degree of DIP joint radiographic OA was required in both studies for subjects to be classed as affected, and the mean age of those affected was five years younger in our study (53.2 years v 58.9 years). However, our study failed to reproduce the suggestive linkages reported close to the centromere. Nevertheless, it is worth noting that the highest linkage peak identified in our analysis (NPL score of 1.05) occurred at one of the markers (IL1R1) where suggestive linkage was identified in the Finnish study. Our result may represent a minor contribution of this locus in a more heterogeneous population. However, larger sample sizes would be needed to investigate this hypothesis.

For most disease models the variance components analysis with 436 phenotypic scores will have greater power to detect linkage than the dichotomous trait analysis with 69 affected people. Using simulations and theoretical results for the power of variance components analysis\(^{12-15}\) based on the total number of relative pairs in our sample (439 sibling pairs, 589 aunt pairs, 559 cousin pairs—including both concordant and discordant relative pairs), we have about 70% power to detect a LOD score of 2 at a quantitative trait locus with heritability of 30%. However, in our variance components analysis, no loci had a LOD score of 2 at a quantitative trait locus with heritability of 20%.

For our multipoint analysis we have about 70% power to detect a LOD score of 2 at a quantitative trait locus with heritability of 30%. However, in our variance components analysis, no loci had a LOD score of 2 at a quantitative trait locus with heritability of 20%.

Considering that our sample size was comparable with or greater than sample sizes in previous hand OA studies reporting linkage, it seems likely that loci outside 2q contribute to susceptibility or severity of hand OA in the Tasmanian population, and loci in 2q make at most a small contribution, indicating the need for genotyping in other regions of the genome.

### ACKNOWLEDGEMENTS
We thank the families for their participation. Thanks also go to Sr Catrina Boon who coordinated fieldwork for this study and Michele Brown for technical assistance. This work was funded by the Cooperative Research Centre for Discovery of Genes for Common Human Diseases and Cerylid Biosciences Pty Ltd. The CRC for Discovery of Genes for Common Human Diseases is established and supported by the Australian Government’s Cooperative Research Centres Program. HC is a recipient of the McGill Fellowship of the Arthritis Foundation of Australia.

### Authors’ affiliations
- J Stankovich, M M Sale, H M Cooley, A Reilly, J L Dickinson, G Jones, Menzies Centre for Population Health Research, University of Tasmania, GPO Box 252–23, Hobart, Tasmania, 7001, Australia
- M Bahlo, The Walter and Eliza Hall Institute of Medical Research, Royal Melbourne Hospital, Victoria, 3050, Australia

### REFERENCES

www.annrheumdis.com