Raised human cartilage glycoprotein-39 plasma levels in patients with rheumatoid arthritis and other inflammatory conditions

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Abstract

Objective—To evaluate plasma human cartilage glycoprotein (HC gp-39) as a possible marker for the presence and/or activity of rheumatoid arthritis (RA) and other inflammatory conditions.

Background—HC gp-39 is a secretory product of chondrocytes, synovial cells, macrophages, and neutrophils. HC gp-39, also described as YKL-40, was found to be a marker of joint disease and tissue injury in RA and various other diseases.

Methods—Levels of HC gp-39 were determined by a sandwich enzyme linked immunosorbent assay (ELISA) in 47 patients with RA, 47 with osteoarthritis (OA), 24 with systemic lupus erythematosus (SLE), 24 with inflammatory bowel disease (IBD), and in 47 healthy controls. A disease activity score was assessed in the patients with RA, SLE, and IBD.

Results—The plasma level of HC gp-39 in the RA patient group was significantly higher than in the other patient groups and healthy controls. The level in patients with OA, SLE, and IBD was also significantly higher than the HC gp-39 level found in the healthy control group. HC gp-39 levels in patients with RA correlated positively with the ESR and IgM rheumatoid factor level but not with other variables of disease activity. In the patients with SLE and IBD no correlation was found with the disease activity score.

Conclusion—The plasma level of HC gp-39 is increased in inflammatory conditions with and without joint disease (SLE, IBD, OA, and RA). Thus increased levels of HC gp-39 do not only reflect joint disease but also reflect inflammation or tissue degradation in various conditions. Notably, the highest level of HC gp-39 was found in patients with RA. Only in the RA patient group was a correlation between HC gp-39 plasma levels and some laboratory variables of disease activity found.

Human cartilage glycoprotein 39 (HC gp-39) is a protein with an apparent molecular weight of 42 kDa produced by human chondrocytes, synovial cells, macrophages, and neutrophils. Interest in HC gp-39 in RA was stimulated by the finding that HC gp-39 messenger RNA is expressed in cartilage obtained from patients with RA, whereas healthy adult cartilage did not contain this transcript. Further interest in the glycoprotein was prompted by the predominant recognition of HC gp-39 derived peptides, selected for their ability to bind to RA associated DR4 (DRB1*0401) molecules, by peripheral blood T cells from patients with RA. Moreover, it has been shown that HC gp-39 emulsified in complete Freund's adjuvant induced arthritis in BALB/c mice. This arthritis was suppressed by intranasal administration of HC gp-39 before immunisation, suggesting that immunological tolerance of the protein may control disease activity.

The function of the glycoprotein at present is unknown. HC gp-39 shows homology with the chitinase protein family, but no chitinase activity has been found. The glycoprotein has also been described as YKL-40. Levels of HC gp-39 have been determined in serum and synovial fluid (SF) of patients with arthropathies. HC gp-39 serum levels in patients with joint disease were significantly higher than those found in healthy controls. In this study the patient group with joint disease comprised patients with RA, osteoarthritis (OA), crystal arthritis, reactive arthritis, and undifferentiated monarthritides. The serum HC gp-39 concentrations did not differ significantly between patients with inflammatory joint disease and those with OA. HC gp-39 levels in SF of patients with inflammatory or degenerative joint diseases were 10–15-fold higher than serum levels and a correlation was found between HC gp-39 concentration in SF and serum. These findings suggest that a substantial portion of serum HC gp-39 originates from the joint. HC gp-39 levels in serum and SF strongly correlated with serum C reactive protein (CRP) and SF interleukin 6. In a later study it was shown that serum HC gp-39 levels in patients with active RA (56 patients) and the levels in patients with OA (27) were significantly higher than in patients with inactive RA (9), patients with diabetes (35), and healthy controls (329). No differences in HC gp-39 levels were seen between patients with active RA and patients with OA. In this study no information was given on disease variables of the RA patient group. In a recent study the HC gp-39 levels in patients with RA with clinically and biochemically active disease were significantly higher than in healthy controls and patients with RA with low disease activity. Apart from studies in RA and OA, no data have been published on the level of HC gp-39.
**Table 1**  Clinical and demographic characteristics of the subjects investigated

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>RA* (n=50)</th>
<th>OA* (n=51)</th>
<th>IBD* (n=26)</th>
<th>SLE* (n=24)</th>
<th>HC* (n=49)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age median (years)</td>
<td>65.0</td>
<td>66.0</td>
<td>34.0</td>
<td>38.0</td>
<td>44.0</td>
</tr>
<tr>
<td>Women (No (%))</td>
<td>33 (66)</td>
<td>41 (80)</td>
<td>17 (66)</td>
<td>22 (92)</td>
<td>21 (43)</td>
</tr>
<tr>
<td>Disease established median (years)</td>
<td>7.5</td>
<td>3.5</td>
<td>5.0</td>
<td>4.0</td>
<td>NA*</td>
</tr>
</tbody>
</table>

RA = rheumatoid arthritis; OA = osteoarthritis; IBD = inflammatory bowel disease; SLE = systemic lupus erythematosus; HC = healthy control; NA = not applicable

**Table 2**  Disease characteristics of 50 patients with RA

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>RA* (n=50)</th>
<th>OA* (n=51)</th>
<th>IBD* (n=26)</th>
<th>SLE* (n=24)</th>
<th>HC* (n=49)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rheumatoid factor positive (%)</td>
<td>79</td>
<td>96</td>
<td>24</td>
<td>32</td>
<td>5.0</td>
</tr>
<tr>
<td>Rheumatoid factor IgM (IU/ml, median)</td>
<td>49</td>
<td>8.0</td>
<td>4.5</td>
<td>4.0</td>
<td>3.12</td>
</tr>
<tr>
<td>Erosive disease (%)</td>
<td>96</td>
<td>96</td>
<td>96</td>
<td>96</td>
<td>96</td>
</tr>
<tr>
<td>Nodular disease (%)</td>
<td>24</td>
<td>24</td>
<td>24</td>
<td>24</td>
<td>24</td>
</tr>
<tr>
<td>ESR* (mm/1st h, median)</td>
<td>32</td>
<td>32</td>
<td>32</td>
<td>32</td>
<td>32</td>
</tr>
<tr>
<td>Tender joint score (median)</td>
<td>5.0</td>
<td>8.0</td>
<td>4.5</td>
<td>4.0</td>
<td>3.12</td>
</tr>
<tr>
<td>Swollen joint score (median)</td>
<td>8.0</td>
<td>8.0</td>
<td>4.5</td>
<td>4.0</td>
<td>3.12</td>
</tr>
<tr>
<td>Patient’s assessment score (VAS*, median)</td>
<td>4.5</td>
<td>4.5</td>
<td>4.5</td>
<td>4.5</td>
<td>4.5</td>
</tr>
<tr>
<td>Doctor’s assessment score (VAS, median)</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
</tr>
<tr>
<td>Modified disease activity score (DAS)</td>
<td>3.12</td>
<td>3.12</td>
<td>3.12</td>
<td>3.12</td>
<td>3.12</td>
</tr>
</tbody>
</table>

*ESR = erythrocyte sedimentation rate; VAS = visual analogue scale.

The patient group with IBD consisted of patients with active ulcerative colitis or Crohn’s disease. These patients had no arthritis. All patients with ulcerative colitis had rectal bleeding and diarrhoea, whereas the patients with Crohn’s disease had a disease activity index (CDAI) of 150 or more. The CDAI included variables such as the number of stools, rating of abdominal pain, rating of general wellbeing, and symptoms such as fistulas, anal fissures, skin lesions, and fever.

**PREPARATION OF SAMPLES**

After the patient’s informed consent had been obtained, peripheral blood samples were collected in heparin coated tubes between 9 00 am and 1 00 pm. Plasma samples were stored at −20°C until use for the determination of HC gp-39 levels.

**MONOCLONAL ANTIBODIES**

Bab/c mice were immunised with HC gp-39 purified from cell culture supernatant of MG63 cells emulsified in complete Freund’s adjuvant (1:1). After serum conversion as determined by ELISA, spleen cells were fused with NS1 cells through electrofusion. Clone selection was carried out by ELISA and selections were confirmed by western blot analysis on culture supernatants containing HC gp-39, and by Biacore experiments. Monoclonal antibody (MoAb) 8B was an excellent reagent in western blot, probably recognising a linear epitope of HC gp-39, whereas MoAb 10B is directed against a conformational epitope distant from that of MoAb 8B (determined by Biacore analysis) (data not shown).

**HC GP-39 ELISA**

Polystyrene micro-ELISA strip-plates (Greiner) were coated overnight at 4°C with 100 μl/well of 1 μg/ml anti-HC gp-39 capture antibody (HC gp-39 8B) in phosphate buffered saline (PBS). The supernatant was discarded and the wells were incubated for one hour at room temperature with 200 μl PBS, 0.05 % Tween 20 (PBST) to block excess free binding sites. Subsequently, plates were washed three times with PBST and incubated for one hour at room temperature with 100 μl of sample or recombinant HC gp-39 standard in PBST. Plasma samples were diluted 1:2 or 1:4 in PBST. After this incubation the plates were washed three times with PBST and incubated for one hour at room temperature with horseradish peroxidase labelled anti-HC gp-39 detection antibody (HC gp-39 10B) at a dilution of 1:1000 in PBST. The plates were washed three times with PBST and three times with demi-water. Finally, the colour reaction was developed by incubating the wells with 100...
Table 3 Descriptive statistics of human cartilage glycoprotein-39 plasma levels in the different groups

<table>
<thead>
<tr>
<th>Subject groups</th>
<th>No</th>
<th>Geometric mean (ng/ml)</th>
<th>RSD* (%)</th>
<th>Median (ng/ml)</th>
<th>Duncan grouping</th>
</tr>
</thead>
<tbody>
<tr>
<td>RA*</td>
<td>47</td>
<td>38.3</td>
<td>112.3</td>
<td>31.0</td>
<td>A</td>
</tr>
<tr>
<td>IBD*</td>
<td>24</td>
<td>24.8</td>
<td>72.0</td>
<td>22.5</td>
<td>B</td>
</tr>
<tr>
<td>OA*</td>
<td>47</td>
<td>24.7</td>
<td>76.6</td>
<td>20.6</td>
<td>B</td>
</tr>
<tr>
<td>SLE*</td>
<td>24</td>
<td>22.6</td>
<td>95.2</td>
<td>21.1</td>
<td>B</td>
</tr>
<tr>
<td>HC*</td>
<td>47</td>
<td>15.5</td>
<td>42.0</td>
<td>14.8</td>
<td>C</td>
</tr>
</tbody>
</table>

*HC = healthy control; RSD = relative standard deviation; RA = rheumatoid arthritis; IBD = inflammatory bowel disease; OA = osteoarthritis; SLE = systemic lupus erythematosus.
†Duncan grouping (Duncan’s multiple range test): means with the same letter are not significantly different (p>0.05); means with different letters differ significantly (p<0.05).

Results

Table 1 summarises the demographic and clinical characteristics of the different patient groups and healthy controls. Table 2 gives the disease characteristics of the RA patient group. It can be concluded that the RA patient group is a typical cross section of the RA population visiting the outpatients clinic of a tertial referral centre. The RA patient group was a heterogeneous group with respect to disease characteristics: the duration of disease varied from 1 to 56 years, the swollen joint count from 1 to 22, the tender joint count from 0 to 20, and the ESR from 2 to 89 mm/1st h. The clinical variables of the patients with SLE, of whom 87.5% had a history of arthritis, also varied considerably: the duration of disease varied from 1 to 18 years and the SLEDAI varied from 0 to 24. For patients with IBD, the disease duration varied from 2 to 31 years. In the latter patient group a disease activity score index (CDAI) was measured only for the 13 patients with Crohn’s disease; this varied from 106 to 366 (one patient with a CDAI below 150 (106) was mistakenly included). In the OA patient group most patients had localised OA (71%).

Table 3 gives the HC gp-39 plasma levels for the various groups (for 11 subjects—three patients with RA, four patients with OA, two patients with IBD, and two healthy controls—the levels could not be reported). HC gp-39 plasma levels differed significantly between patient groups and the control group (ANOVA, p=0.0001) (table 3, fig 1). From Duncan’s test it was found that the mean of the HC gp-39 plasma levels in the RA group was significantly higher than the means found in the other patient groups and the healthy control group (table 3, p<0.05). The means of the HC gp-39 plasma levels in the OA, SLE, and IBD patient groups were also significantly higher than that of the healthy control group (p<0.05). No significant difference was observed between the mean plasma levels of HC gp-39 between the OA, SLE, and IBD patient groups (table 3).

Within the group of patients with RA a significant positive correlation was found between the level of HC gp-39 and the ESR (r=0.006), and between HC gp-39 and the IgM rheumatoid factor level (p=0.007) (table 4). There was no correlation with other clinical variables, such as the duration of symptoms of disease, the presence of extra-

Figure 1 Human cartilage glycoprotein-39 (HC gp-39) levels in different patient groups. IBD = inflammatory bowel disease; OA = osteoarthritis; RA = rheumatoid arthritis; SLE = systemic lupus erythematosus.
articular manifestations, the radiological stage of disease, the swollen or tender joint count, or the DAS. In the SLE and IBD patient groups the level of HC gp-39 did not correlate with any of the disease activity variables (SLEDAI and CDAI for Crohn’s disease) respectively, plus disease duration measured.

Discussion
The present study shows that serum levels of HC gp-39 in patients with RA are significantly higher than those in healthy controls. This finding is in agreement with previous studies that also showed raised levels of HC gp-39 in patients with arthritis.1 2 8 9 10 This study further showed that HC gp-39 levels are raised in OA, SLE, and IBD (compared with healthy controls), though lower levels were found in these conditions than in the RA patient group. It is recognised, in our study, that the patients in the RA and OA group represent a relatively older population with a higher percentage of women than in the healthy control group. Previously, however, no age or sex differences were found in serum HC gp-39 levels in subjects younger than 70,11 thereby suggesting that these variables are less relevant. Indeed, we did not find a significant correlation between plasma HC gp-39 levels and age in the RA and OA patient groups.

In contrast with the findings in previous studies, our study shows a difference in HC gp-39 levels for patients with RA and OA. The lower levels found in patients with OA are thought to be related to the fact that a substantial fraction (71%) of patients with OA had localised (two areas or fewer) joint disease. This study is the first to demonstrate raised levels of HC gp-39 in patients with SLE and IBD. This is in line with previous studies which showed that HC gp-39 levels can be found in conditions not related to RA. In patients with alcoholic cirrhosis a correlation was found between the plasma HC gp-39 level and the degree of liver fibrosis.3 In that study it was shown that HC gp-39 is released from the hepatosplanchnic area, providing evidence that the glycoprotein is produced by the liver. Immunohistochemistry showed that HC gp-39 is localised in fibrotic areas of the liver. In addition, raised HC gp-39 levels were found in patients with recurrent breast cancer and colorectal cancer, where the protein proved to be a prognostic marker for short survival.3 12 More recently, it has been shown that diabetic patients have increased levels of HC gp-39.13 Our present observations further add to the view that the glycoprotein is not joint-specific and that it is upregulated in different conditions not necessarily involving the joint.

HC gp-39 levels in patients with RA correlated positively with the ESR and surprisingly also with IgM RF level. No correlation, however, was found with clinical variables of disease activity. Further, raised HC gp-39 levels in patients with SLE and IBD did not correlate with disease activity scores. Previously, raised HC gp-39 levels in patients with RA were found to be positively correlated with the ESR, the CRP level, and the joint counts for swelling and pain in patients with RA.12 This discrepancy might be attributed to the fact that in our study the age, disease duration, and disease activity of the RA patient group varied considerably. In the study by Harvey et al an increased level of HC gp-39 was found in patients with active RA only, but no details were given about disease activity variables.14

HC gp-39 is produced by human synoviocytes, chondrocytes, and by macrophages, neutrophils, and monocytes.1 4 11 Further evidence suggests that HC gp-39 is also expressed by peripheral blood mononuclear cells in patients with RA.20 We reasoned that the glycoprotein, whether produced locally at the site of inflammation or systemically, is widely expressed and thus may be a prominent target for specific T cells.2 Such a situation creates possibilities for the development of a specific immunomodulatory treatment. In mouse studies it was shown that intranasal application of the protein suppressed inflammation by generation of HC gp-39 specific modulatory T cells, which may then travel to the joints.2 21

The data above support the hypothesis that HC gp-39 is a promising candidate antigen for tolerance induction in RA. The results of the present and previous studies support this hypothesis by finding the highest level of HC gp-39 expression in patients with RA. It remains to be clarified whether the raised HC gp-39 levels found in other inflammatory conditions are relevant to the autoimmune response underlying the disease process.

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