Basic calcium phosphate crystals induce synthesis and secretion of 92 kDa gelatinase (gelatinase B/matrix metalloprotease 9) in human fibroblasts

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Abstract
Objective—Synovial fluid basic calcium phosphate (BCP) crystals are associated with severe joint degeneration accompanied by synovial hypertrophy. The metalloprotease 92 kDa gelatinase (MMP-9) has been implicated in the degradation of extracellular matrix in osteoarthritis, but the ability of BCP crystals to induce gelatinase in human fibroblasts or in adult porcine chondrocytes has not previously been studied. The hypothesis that the mitogenic response to BCP crystals is accompanied by induction and secretion of MMP-9 was studied.

Methods—MMP-9 messenger RNA (mRNA) was detected by northern blot and reverse transcription-polymerase chain reaction (RT-PCR). Gelatinase secretion was identified by western blot and zymography of conditioned media.

Results—BCP crystals caused a concentration dependent induction of MMP-9 mRNA accumulation and protein secretion in human fibroblasts but not in adult porcine chondrocytes.

Conclusion—BCP crystals induce MMP-9 production by HF but not adult porcine chondrocytes. Fibroblast MMP-9 may be an important mediator of the joint destruction associated with synovial fluid BCP crystals.

Concurrence of deposits of calcium containing crystals such as basic calcium phosphate (BCP) (hydroxyapatite, tricalcium phosphate, and octacalcium phosphate) and degenerative joint disease is well established. The prevalence of BCP crystals in synovial fluid (SF) from patients with knee osteoarthritis (OA) is between 30 and 60%. Ample data support the role of these crystals in cartilage degeneration as their presence correlates strongly with radiographic evidence of cartilage degeneration and larger joint effusions are seen in affected knee joints when compared with joint fluid from OA knees without crystals. The pathogenicity of these crystals is further supported by their association with a distinctive type of non-inflammatory destructive arthropathy affecting the shoulders and knees called Milwaukee Shoulder Syndrome. Milwaukee Shoulder Syndrome is characterised by severe glenohumeral degeneration with loss of articular cartilage and dissolution of the rotator cuff. Large, non-inflammatory joint effusions and pronounced synovial proliferation with frequent formation of pedunculated loose bodies are typical.

The biological effects of BCP crystals that may promote articular damage have been investigated in vitro. BCP crystals induce mitogenesis in a number of cell types, including fibroblasts and chondrocytes. This property presumably accounts for the synovial proliferation characteristic of both Milwaukee Shoulder Syndrome and BCP crystal associated OA. BCP crystals also induce coordinate synthesis of the metalloproteases collagenase and stromelysin in human fibroblasts. Secretion of these enzymes permits degradation of many matrix components including type II collagen, fibronectin, laminin, and proteoglycan. These two enzymes have been postulated to mediate, at least in part, the joint destruction associated with the presence of BCP crystals in SF.

The 92 kDa gelatinase (gelatinase B/matrix metalloprotease 9) (MMP-9), a 92 kDa metalloprotease, has also recently been implicated in the degradation of extracellular matrix in joint degeneration as it is expressed in osteoarthritic, but not in normal, human articular cartilage. It is also inducible in normal human cartilage and in normal human fibroblasts treated with interleukin 1 (IL-1). Gelatinases may influence the breakdown of cartilage in OA by accelerating degradation of matrix components or by digesting the matrix molecules such as types IV, V, and XI collagens as well as denatured collagens (gelatins). Although MMP-9 may contribute to the pathogenic effects of BCP crystals in joint degradation, the ability of BCP crystals to induce gelatinase in human fibroblasts or cartilage has not previously been studied. We hypothesised that the mitogenic response to BCP crystals is accompanied by the induction and secretion of MMP-9 in both human fibroblasts and adult porcine chondrocytes.

Methods
CRYSTAL SYNTHESIS
BCP crystals were synthesised by a modification of published methods. Mineral prepared by this method had a calcium/phosphorus molar ratio of 1.59 and contained partially carbonate substituted hydroxyapatite with admixed octacalcium phosphate by Fourier transform infrared spectroscopy. Retention of the crystal character was confirmed by x ray diffraction and Fourier transform infrared spectroscopy. The crystals were crushed and
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PROBES AND REAGENTS
The MMP-9 complementary DNA (cDNA) probe was a 2.3 Kb EcoR1 fragment from the pETNG clone. The pHcGAP plasmid containing glyceraldehyde-3-phosphate dehydrogenase (GAPD) cDNA was from the American Type Culture Collection. The MMP-9 antibody was a polyclonal antibody raised in rabbits against a synthetic peptide of MMP-9. Enhanced chemiluminescence was performed with a kit from Amersham (Arlington Heights, IL). Type I gelatin from porcine skin was from Sigma Chemical Co, St Louis, MO. Recombinant tumour necrosis α (TNFα) and interleukin 1α (IL1) were obtained from Genzyme (Cambridge, MA). Cycloheximide (IL1) was obtained from Genzyme (Cambridge, MA).

IMMUNOBLOTTING
Gelatinase protein secretion into conditioned media was confirmed by gelatin substrate gels. Samples of conditioned media (20 µl) were electrophoresed through 10% polyacrylamide gels under reducing conditions, transferred onto nitrocellulose membranes, and western blot was performed using a 1:1000 dilution of antibody to MMP-9 as described previously. Immunoreactive bands were detected using enhanced chemiluminescence reagents (Amersham Corp) and autoradiography.

ENZYME ACTIVITY
Gelatinase enzyme activity in conditioned media was determined using gelatin substrate gels. Aliquots (115 µl) of conditioned media were electrophoresed on 10% polyacrylamide gels containing 1 mg/ml type I gelatin under non-reducing conditions. After electrophoresis, gels were washed in 2.5% Triton X-100 for one hour to allow proteins to renature. Gels were then incubated overnight in substrate buffer (50 mM TRIS, pH 8.0, 150 mM NaCl, 10 mM CaCl2, 1 µM ZnCl2, 0.02% Na azide) at 37°C, rinsed in water, and stained in 0.5% w/v Coomassie brilliant blue in 10% acetic acid and 30% isopropyl alcohol. Destained gels showed a clear zone of lysis where gelatin degradation, an index of gelatinase activity, had occurred.

RESULTS
BCP CRYSTAL INDUCTION OF MMP-9 mRNA
While BCP concentrations as low as 9 µg/cm² resulted in significant accumulation of MMP-9 mRNA, a positive dose responsiveness was demonstrated with maximal accumulation seen after stimulation with 18 or 36 µg/cm² (fig 1). BCP crystal induced accumulation of MMP-9 mRNA in HP was detectable within 16 hours after stimulation and continued for at least 24 hours (data not shown). TPA, a known inducer of MMP-9, served as a positive control. As it has been demonstrated that new protein synthesis is required for accumulation of...
Figure 1  BCP crystal induced accumulation of 92 kDa gelatinase (MMP-9) messenger RNA (mRNA). Confluent cultures of human fibroblasts on 100 mm plates were incubated in DMEM and 0.5% FBS for 24 hours and were then stimulated with BCP crystals in the quantities shown. Control cultures were stimulated with TPA (200 nm) or left unstimulated. BCP crystals were also added to some cultures treated with the protein synthesis inhibitor, cycloheximide (CHX, 10 µg/ml). After 24 hours, all plates were harvested and total RNA isolated. Northern blot analysis was performed using a specific cDNA probe for MMP-9 (GL) followed by autoradiography. The MMP-9 probe was then removed by washing, and the blot was reprobed with glyceraldehyde-3-phosphate dehydrogenase (GAPD) cDNA, to ensure analysis of equivalent quantities of total mRNA under each condition. The positions of the 18S and 28S ribosomal bands are shown.

Figure 2  BCP crystal induced stimulation of MMP-9 synthesis. Confluent, quiescent human fibroblast cultures incubated in DMEM containing 0.5% FBS were stimulated with either BCP crystals (B) at varying concentrations, TPA (200 nm) or TNF (10 ng/ml); fibroblasts in 0.5% FBS alone (C) were used as a control. The culture medium was harvested after 48 hours and analysed by western blotting using a specific polyclonal antibody to MMP-9. Molecular weight markers are shown on the left.

Identification of MMP-9 Protein in Conditioned Media from Cultures Treated with BCP Crystals

Figure 2 shows the induction by BCP crystals, TPA or TNF of the secretion of a protein of molecular weight 92 kDa, immunoreactive with a polyclonal antibody to MMP-9, into conditioned media. The secretion of this protein was maximum 48 hours after stimulation, the longest incubation tested. This protein was not evident in control cultures incubated in 0.5% FBS alone. In contrast, in adult porcine chondrocytes there was no evidence of secretion of a 92 kDa protein, immunoreactive with a polyclonal antibody to MMP-9, into conditioned media when cultures were either unstimulated or treated with BCP crystals, TPA, TNF or 10% FBS (data not shown).

Identification of MMP-9 Enzyme Activity in Conditioned Media from Cultures Treated with BCP Crystals

To assess gelatinase activity in the conditioned media from human fibroblast cultures stimulated with BCP crystals, aliquots of conditioned media were analysed by SDS-PAGE on polyacrylamide gels containing gelatin substrate (fig 3). Conditioned media from cultures treated with TPA were used for comparison. Bands of lytic activity at 92 kDa consistent with MMP-9 were seen in conditioned media from human fibroblast cultures treated with BCP crystals or TPA. No lytic activity at 92 kDa was found in conditioned media taken from unstimulated control cultures. Lytic activity at 72 kDa resulted under all conditions consistent with 72 kDa gelatinase but seemed to be increased in the conditioned media of cultures treated with BCP crystals, TNF or TPA.

In contrast, in adult porcine chondrocytes there was no evidence of lytic activity at 92 kDa in conditioned media from cultures treated with TPA, IL1 or TNF although lytic activity at 72 kDa was apparent under all conditions (fig 3).

Discussion

This is the first demonstration of the induction of MMP-9 mRNA, followed by production and secretion of active MMP-9 enzyme, in human fibroblasts treated with BCP crystals. The time course of induction indicated that it occurred in coordination with collagenase and stromelysin. Connective tissue gelatinases are
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thought to be important in both normal matrix turnover and in pathological remodelling of cartilage because of their ability to act synergistically with collagenase and stromelysin in the degradation of the macromolecules of extracellular matrices. Both collagenase and neutral protease activity in synovial fluid from patients with Milwaukee Shoulder Syndrome was reported in the original and subsequent descriptions of this arthropathy. The neutral protease activity was probably caused by stromelysin but unfortunately, the presence of gelatinase was not evaluated at that time.

We were unable to demonstrate accumulation of MMP-9 mRNA in normal adult porcine chondrocytes treated with BCP, TPA, IL1 or TNF; nor was there evidence of production or secretion of MMP-9, or both, in conditioned media as assessed by western blot and zymogram. Using PCR, constitutive expression of MMP-9 mRNA was identified in adult porcine chondrocytes but it was not increased after treatment with either BCP crystals, TPA, TPA or IL1. The chondrocyte data reported here contrast with studies of articular chondrocytes from human osteoarthritic cartilage in which MMP-9 mRNA was found with associated secretion of active enzyme. Expression of MMP-9 was increased in fibrillated cartilage. Active MMP-9 was also identified in the concentrated conditioned media from normal human articular chondrocytes treated with IL1.

Such variation of expression of MMP-9 between APC and human osteoarthritic cartilage and human articular chondrocytes may reflect interspecies differences. None the less, adult porcine chondrocytes are capable of producing other MMPs as we have shown the induction of collagenase in adult porcine chondrocytes treated with BCP crystals. and epidermal growth factor both induce collagenase and stromelysin in adult porcine chondrocytes. However, our data suggest that, in vivo, synovial fibroblasts are probably the most important source of intra-articular MMPs resulting in matrix degradation associated with BCP crystals and that chondrocytes are unlikely to produce biologically significant quantities of MMP-9. Although neutrophils secrete MMP-9, BCP crystal associated joint effusions contain few of these cells. Therefore, neutrophils are a possible but improbable source of MMP-9 in BCP crystal associated arthropathies.

In this study, there was constitutive production of 72 kDa gelatinase in human fibroblast conditioned media as assessed by western blot and in human fibroblasts and adult porcine chondrocytes as assessed by zymography. Enzyme activity in human fibroblasts, but not adult porcine chondrocytes conditioned media seemed to be slightly increased after treatment with BCP crystals, TPA or TPA. Although recent studies have shown that both 92 kDa and 72 kDa gelatinases have similar substrate specificity for type IV and V collagens as well as gelatin, the full complement of substrates of either gelatinase has yet to be explored. Because of this, the relative intensities of the zones of lysis created by the 72 and 92 kDa gelatinases on the gelatin substrate gel may be misleading in terms of their physiological significance.

Although MMP-9 mRNA accumulation and protein secretion in response to BCP crystals was less than that noted in response to TPA, enzyme activity in conditioned media from both TPA and BCP crystal treated cultures was comparable. The level of induction of MMP-9 in fibroblasts in response to BCP crystals is probably biologically significant. The data presented in this paper support our hypothesis that the mitogenic response to BCP crystals is accompanied by 92 kDa gelatinase induction and secretion. The coordinate induction and subsequent secretion of 92 kDa gelatinase, collagenase and stromelysin, probably from synovial fibroblasts, may underlie the pronounced, non-inflammatory degradation of cartilage and other matrix rich joint structures found in association with synovial fluid BCP crystals.


