The poor man’s capillary microscope: A novel technique for the assessment of capillary morphology

Rupert M Bauersachs, Frank Löstner

Abstract

Objectives—To evaluate a new method for rapid and efficient assessment of capillary morphology.

Methods—Nailfold capillary morphology in 18 patients with Raynaud’s phenomenon was investigated with a new bedside test, based on a modified dermatoscope using a novel gel-immersion technique. These findings were compared with those obtained by standard capillary microscopy.

Results—With the standard microscope, six patients had dilated capillaries, six had mega-capillaries, seven had avascular fields, eight had ramified capillaries, six had contorted capillaries, and seven patients showed micro-haemorrhages, respectively. The dermatoscope identified exactly the same patients to have the same capillary abnormalities. One hundred and thirty six fields, eight had ramified capillaries, six had contorted capillaries, and seven patients showed micro-haemorrhages, respectively. The average examination time was 18 (range 8–30) minutes with the standard microscope and 4 (3–8) minutes with the dermatoscope.

Conclusion—A new diagnostic tool for rapid and efficient examination of nailfold capillaries is described for circumstances when a standard microscope is not available. This study shows that the handheld device can be used in clinical routine with sufficient diagnostic efficacy and little expenditure, both timewise and financially.

(Royal Soc of Rheumatology 1997;56:435–437)

Capillary microscopy is a valuable diagnostic tool that can reveal microangiopathy and early diagnostic signs characteristic for scleroderma. Despite several drawbacks, this method has found some support, mainly because of the lack of other simple and inexpensive methods. Unfortunately, too often capillary morphology is not assessed at all because of the lack of equipment or time.

Therefore, there is a need for a quick and simple capillary investigation technique. This study evaluates whether a modified dermatoscope with a new immersion technique can be used as a diagnostic tool for the examination of nailfold capillaries, and how its findings compare with those obtained by a standard capillary microscope.

Methods

We modified a single lens dermatoscope (epiluminescence microscope: Heine Dermatoskop Delta 10, Heine Optotechnik, 82211 Herrsching, Germany (fig 1), which initially had been developed for the inspection of skin lesions and malignant melanoma. For the examination of nailfold capillaries several adaptations had to be made: (1) The single lens magnification was changed from 10× to 12×; (2) the light bulb was tinted green (wavelength approximately 490 nm) to increase the contrast of the red cell column in the capillaries (bulb paint by Conrad Electronic, Hirschau, Germany); (3) the original bulbs were replaced by 2.5 V Kryptogen bulbs (Heine Optotechnik, 82211 Herrsching, Germany); (4) a graded (millimeter) contact slide was used on the covering surface of the device to estimate the size of avascular areas and capillary diameter; the thickness of the lines was 50 and 100 μm, respectively; (5) common immersion oil was substituted by a Carbachol gel (ultrasound transmission gel, General Pharmacy, Hospital Bogenhausen, Munich, Germany), which can easily be applied between the fingernail and the dermato- scope without running off the finger during the examination. We measured a refractive index of 1.348 of the gel.

The examiner using the dermatoscope had no knowledge of the videomicroscope findings and vice versa. The reference microscope was a stereoscopic, epiluminescence microscope (Wild M5B, Heerbrugg, Switzerland), with video recording and analysis (Panasonic AG-6400-E, Matsushita Electric Industrial Co, Ltd, Japan; TM-150 PSN Monitor, JVC, Yokohama, Japan). The magnification was 60-fold to 240-fold; the examination followed a standard procedure. For the dermatoscope examination, a small portion of immersion gel
Because of the limited visibility in some fingers, the total number of comparisons may differ. The kappa test statistic (κ) is used for statistical analysis of the correlation between the findings of the two methods.

**Results**

Using the standard microscope, patients with the following capillary abnormalities were identified: six patients had dilated capillaries and six had mega-capillaries, in seven patients avascular fields were observed, in eight patients ramified capillaries, and in six contorted capillaries, while micro-haemorrhages were found in seven patients. With the dermatoscope exactly the same patients were identified to have the same capillary abnormalities.

For the assessment of capillary morphology of individual fingers, visibility in a total of 144 fingers was considered sufficient in 126 with the videomicroscope and in 124 fingers with the dermatoscope, respectively. Table 1 shows the results of the pairwise comparison of the two methods for individual fingers. Note, that because of impaired visibility in some fingers, on average only 136 pairs were usable for comparison.

**Discussion**

We present a new diagnostic tool for quick and efficient examination of nailfold capillaries for circumstances when standard microscope equipment is not available. The method is based on a novel immersion technique, which uses a highly viscous immersion gel. This study shows that the hand held device can be used with sufficient diagnostic efficacy in clinical routine with little expenditure, both timewise and financially, as the dermatoscope costs less than a standard ophthalmoscope. The dermatoscope seems to be comparable, if not superior to the ophthalmoscope as judged by data from an earlier report using the same statistical approach (κ statistic 0.816 to 0.929 for the ophthalmoscope compared with a videomicroscope), even though this would have to be confirmed by direct comparison. For ramified and contorted capillaries there were about 10% false negative examinations, which illustrates the limits of this method. Still, the practical advantages, the good and straightforward visualisation of capillaries, and the wide eye field with the dermatoscope is obvious from the very first clinical use. The lack of photographic documentation does not seem to be an important problem, as pathological findings should lead to an extended examination with a standard microscope. The standard 10 × magnification of the dermatoscope may also yield sufficient results, however, it seems that capillary morphology can more readily be assessed with the higher 12 × magnification, even though the depth of focus is more limited.

The dermatoscope, with or without modification, may serve not only the poor man as a simple capillary microscope, but also the rich man, who may not have the time to use his more elaborate apparatus whenever indicated.

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**Table 1** Capillary morphology of individual fingers in 18 patients with Raynaud’s phenomenon, assessed with a standard capillary microscope in comparison with the modified dermatoscope

<table>
<thead>
<tr>
<th>Capillary Abnormality</th>
<th>Dermatoscope</th>
<th>Microscope</th>
<th>Present</th>
<th>Absent</th>
<th>κ</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dilatation</td>
<td>Dermatoscope</td>
<td>Present</td>
<td>37</td>
<td>3</td>
<td>0.93</td>
</tr>
<tr>
<td></td>
<td>Microscope</td>
<td>Absent</td>
<td>2</td>
<td>92</td>
<td></td>
</tr>
<tr>
<td>Mega-capillaries</td>
<td>Dermatoscope</td>
<td>Present</td>
<td>27</td>
<td>2</td>
<td>0.97</td>
</tr>
<tr>
<td></td>
<td>Microscope</td>
<td>Absent</td>
<td>0</td>
<td>104</td>
<td></td>
</tr>
<tr>
<td>Avascular fields</td>
<td>Dermatoscope</td>
<td>Present</td>
<td>27</td>
<td>4</td>
<td>0.93</td>
</tr>
<tr>
<td></td>
<td>Microscope</td>
<td>Absent</td>
<td>0</td>
<td>104</td>
<td></td>
</tr>
<tr>
<td>Ramified capillaries</td>
<td>Dermatoscope</td>
<td>Present</td>
<td>32</td>
<td>13</td>
<td>0.78</td>
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<tr>
<td></td>
<td>Microscope</td>
<td>Absent</td>
<td>2</td>
<td>88</td>
<td></td>
</tr>
<tr>
<td>Contorted capillaries</td>
<td>Dermatoscope</td>
<td>Present</td>
<td>100</td>
<td>12</td>
<td>0.81</td>
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<td></td>
<td>Microscope</td>
<td>Absent</td>
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<td>22</td>
<td></td>
</tr>
<tr>
<td>Micro-haemorrhages</td>
<td>Dermatoscope</td>
<td>Present</td>
<td>35</td>
<td>4</td>
<td>0.94</td>
</tr>
<tr>
<td></td>
<td>Microscope</td>
<td>Absent</td>
<td>0</td>
<td>97</td>
<td></td>
</tr>
</tbody>
</table>

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This 63 year old woman with chronic renal failure, on haemodialysis, developed painful, hard swellings in her hands and elbows.

Radiographs confirmed calcinosis, and material aspirated from a finger was confirmed as hydroxyapatite.

Soft tissue calcification in chronic renal failure may be visceral or non-visceral. Various calcium crystals may be responsible, most commonly calcium phosphates and oxalates.\(^1\)

Visceral calcification, in heart, lung and muscle, is usually an amorphous compound similar to whitlockite (anhydrous calcium carbonate). Non-visceral calcification in arteries, skin, and periarticular tissues is more commonly hydroxyapatite.\(^2\) Deposits may undergo a fibrotic reaction and become encapsulated, appear multiloculated, and spontaneously discharge through the skin.


**Contributors:** Gary D Wright, Michael Doherty. City Hospital, Nottingham, NG5 1PB, United Kingdom.