Increased concentrations of nitrite in synovial fluid and serum samples suggest increased nitric oxide synthesis in rheumatic diseases

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Abstract
Cytokines induce nitric oxide synthesis by endothelial cells, macrophages and polymorphonuclear leucocytes, indicating a role for nitric oxide in inflammatory processes. Nitric oxide production was therefore measured indirectly as nitrite in serum and synovial fluid samples from patients with rheumatoid arthritis (RA) and osteoarthritis (OA) together with serum samples from healthy volunteers matched for age and sex. Serum nitrite concentrations in patients with RA and OA were significantly higher than in controls. In both disease groups synovial fluid nitrite was significantly higher than serum nitrite, implying nitric oxide synthesis by the synovium. Serum and synovial fluid nitrite concentrations in RA were also significantly higher than those in OA. These data show increased nitric oxide production in RA and OA and suggest a role for nitric oxide as an inflammatory mediator in rheumatic diseases.


Nitric oxide has been shown to be the endothelium derived relaxing factor. It is synthesised by the vascular endothelium from L-arginine and interacts with the haem group of guanylate cyclase to increase cGMP formation in vascular smooth muscle cells. Altered nitric oxide synthesis may affect the pathogenesis of some cardiovascular diseases. The L-arginine to nitric oxide biosynthetic pathway has also been identified in numerous other cells and tissues including the adrenal glands, brain, platelets, and some non-adrenergic non-cholinergic nerve terminals, suggesting a more widespread role for nitric oxide in the regulation of cellular function and communication. In all these cells nitric oxide is synthesised by a constitutive Ca$^{2+}$ calmodulin dependent nitric oxide synthase. Nitric oxide is also generated by macrophages, polymorphonuclear leucocytes, lymphocytes, and hepatocytes, though in these cells an inducible enzyme is responsible for its synthesis. This enzyme is not Ca$^{2+}$ dependent, is induced by cytokines, and its induction is inhibited by glucocorticoids. Moreover, its expression results in the sustained release of nitric oxide in amounts far in excess of those by the constitutive pathway. This enzyme was originally described in macrophages in which nitric oxide is essential for cytotoxicity against tumour cells and protozoa. The latter may require an interaction of nitric oxide with the superoxide anion, which although neutralising the effects of nitric oxide and superoxide may, via peroxynitrite, lead to the formation of hydroxyl radicals. This suggests a role for nitric oxide in inflammatory diseases and especially rheumatoid arthritis (RA), in which there is clear evidence for the release of cytokines and for hypoxic ischemia phenomena and free radical generation. For these reasons we have measured the concentrations of nitrite, a breakdown product of nitric oxide, in serum and synovial fluid from patients with RA and osteoarthritis (OA) and in the serum of controls matched for age and sex.

Patients and methods
Paired serum and synovial fluid samples were obtained from 25 patients with active RA fulfilling the 1987 American Rheumatism Association criteria (19 women, six men, mean age 62.4 years) and 19 patients with primary OA (10 women, nine men, mean age 62.8 years). Diagnostic accuracy was assured by clinical review and concurrent laboratory assessment in all patients, which included determination of serum rheumatoid factor, serum C reactive protein (CRP), synovial fluid white blood cell counts, and, in appropriate cases, microscopic study of synovial fluid samples for crystals. All patients were receiving either therapeutic or diagnostic knee aspiration. Serum samples were also obtained from 16 healthy volunteers matched for age and sex. After centrifugation at 2500 g serum and cell free synovial fluid were stored at −70°C until analysis.

Investigating the role of nitric oxide in RA or any other disease requires a simple measure of nitric oxide. Its short half-life makes its direct measurement impractical, though in aqueous solutions nitric oxide decays to yield equal amounts of nitrite and nitrate, which are used as indices of nitric oxide synthesis in vitro. In vivo dietary intake might obscure any differences between samples or disease groups, however. In this respect British surveys have estimated a mean daily intake from food of 95 mg nitrate and 1.4 mg nitrite, equivalent to plasma nitrate concentrations of 30 μmol/l and undetectable concentrations of nitrite. Lower dietary nitrate intake and our preliminary findings of nitrite concentrations less than 1 μmol/l suggested that nitrite offers a more sensitive index of endogenous nitric oxide production and is less subject to dietary influences than nitrate.

The nitrite content of the samples was determined by chemiluminescence. Nitrite was reduced to nitric oxide by refluxing samples in 15% sodium iodide in glacial acetic acid and...
was carried to the reaction chamber by a nitrogen stream bubbled through the reflux mixture. Nitric oxide is determined by the chemiluminescence of its reaction with ozone. The absolute concentration of nitrite in human samples was determined by reference to a standard solution of sodium nitrite. This method is extremely sensitive and can measure picomolar amounts of nitric oxide. A considerable advantage was that biological samples were analysed without further preparatory steps. This avoids techniques such as deproteinisation with organic acids which, in our experience, completely remove nitrite by converting it to gaseous nitric oxide.

### Discussion

There was no significant loss of nitrite from samples following freeze/thawing or storage at −70°C for up to six months. After 15–18 months storage at −70°C the mean nitrite loss from samples was 11%.

### Results

Mean nitrite concentrations in synovial fluid and serum samples were in the μmol/l range (see table). The nitrite concentration in serum samples from patients with RA was over twice that of serum samples from patients with OA (p<0.005). It was also three times that of controls matched for age and sex (0.44 and 0.147 μmol/l respectively, p<0.001) (figure). The serum nitrite concentration in patients with OA was also higher than that in controls matched for age and sex (0.213 and 0.142 μmol/l respectively; p<0.05).

The mean nitrite concentration in synovial fluid in patients with RA was 0.91 μmol/l, which was over twice the 0.44 μmol/l in serum samples (p<0.01). The highest nitrite concentration recorded was 4.27 μmol/l in the synovial fluid of a patient with active RA (CRP 124 mg/l, normal <10 mg/l) and was eight times that of the paired serum sample. The nitrite content of synovial fluid from patients with OA (0.354 μmol/l) was less than that of synovial fluid from patients with RA (p<0.01), but still higher than serum nitrite in patients with OA, which was 0.213 μmol/l (p<0.05). Synovial fluid nitrite was higher than the paired serum sample in 22/25 patients with RA and 16/19 patients with OA.

Synovial fluid nitrite and serum nitrite concentrations were significantly correlated in patients with RA (r=+0.315; p<0.05). In patients with RA there was no significant correlation for serum nitrite v CRP, synovial fluid nitrite v CRP, the ratio synovial fluid/serum nitrite v CRP, nor synovial fluid nitrite v synovial fluid white blood cell count.

### Nitrite concentrations in synovial fluid (SF) and serum samples from patients with rheumatoid arthritis (RA), osteoarthritis (OA) and in serum samples from controls matched for age and sex.

<table>
<thead>
<tr>
<th>Group</th>
<th>Mean (SEM) concentration of nitrite in synovial fluid (μmol/l)</th>
<th>Mean (SEM) concentration of nitrite in serum (μmol/l)</th>
<th>p Value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patients with RA (n=25)</td>
<td>0.910 (0.162)</td>
<td>0.440 (0.052)</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>RA matched controls (n=16)</td>
<td>0.147 (0.016)</td>
<td>0.064 (0.002)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Patients with OA (n=19)</td>
<td>0.354 (0.029)</td>
<td>0.213 (0.049)</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>OA matched controls (n=11)</td>
<td>0.142 (0.018)</td>
<td>0.085 (0.003)</td>
<td>&lt;0.05</td>
</tr>
</tbody>
</table>

*RA synovial fluid v OA synovial fluid, p<0.01; RA serum v OA serum, p<0.005.
Nitric oxide synthesis in rheumatic diseases

inducible nitric oxide pathway plays a part in inflammatory disorders.

The most intriguing finding was that serum nitrite concentrations were increased in patients with RA and OA compared with controls matched for age and sex. The origin of this nitrite is not clear. While synovial inflammation might increase serum nitrite when synovial fluid cleared by the lymphatic system enters the systemic circulation and by equilibration with the vascular compartment within the synovium. This may not entirely account for the higher serum nitrite concentration in RA compared with controls, however, and seems an unlikely explanation in patients with OA. A possible source of increased nitrite is the systemic vasculature and other cells in which the induction of nitrite has been shown. Although difficult to reconcile with current concepts of OA, this is compatible with the systemic nature of RA, in which circulating cytokines are evident and in which vasculitis is a common complication. The precise part played by nitric oxide in inflammation remains to be elucidated. Actions of nitric oxide relevant to this process include its role in the cytotoxic mechanism of activated macrophages, inhibition of iron-sulphur centred enzymes and its anti-proliferative effects.

Nitric oxide is also implicated in chemotaxis of polymorphonuclear leucocytes, and t-arginine, the precursor of nitric oxide, has been shown to have in vivo immunomodulatory actions. The latter include enhanced wound healing and T cell function, notably increased natural and lymphokine activated killer cell activity.

The powerful vasodilating and platelet inhibitory actions of nitric oxide are also pertinent to rheumatoid synovitis. Microvascular injury is prominent in the latter and includes thrombus plugging and features typical of hypoxia. Evidence suggests that the latter is due to hypoxic perfusion injury consequent on impaired synovial perfusion during joint exercise. In this situation nitric oxide induced vasodilation appears to be the removal of superoxide by reaction with nitric oxide might be protective. In vitro, however, the latter reaction yields hydroxyl radicals via the peroxynitrite anion. If the latter occurs to a considerable degree in vivo it may far outweigh the favourable actions of nitric oxide with the net effect of increasing microvascular and synovial tissue injury.

Further investigation is required to establish whether the overall effect of nitric oxide generated by the inducible enzyme is pro-inflammatory. There is clear evidence that induced nitric oxide synthesis leads to cellular injury, which is inhibited by glucocorticoids. The latter occurs at glucocorticoid concentrations within the therapeutic range and appears to be specific as the inhibitory effect is blocked in a concentration dependent manner by corticosterone, a partial agonist of glucocorticoid receptors.

We have shown highly significant differences between nitrite concentrations in synovial fluid and serum samples and in serum samples from patients with RA and OA compared with controls matched for age and sex. This suggests increased endogenous nitric oxide synthesis in these rheumatic diseases and nitric oxide production by the synovium and probably other tissues. It is likely that increased nitric oxide production may play a part in the pathogenesis of these and other diseases in which cytokines have a prominent role. It is possible that some cases of endogenous nitric oxide synthesis and this study shows that nitrite may be measured in samples from humans without complex preparatory steps. These results should encourage other workers to use nitrite concentrations to examine alterations of nitric oxide biosynthesis in other diseases.


