Lupus anticoagulant: clinical significance in anticardiolipin positive patients with systemic lupus erythematosus

N J McHugh, D A H Moye, I E James, M Sampson, P J Maddison

Abstract
The significance of anticardiolipin antibodies and the lupus anticoagulant was studied in 58 consecutive patients with systemic lupus erythematosus. On 85 occasions serum IgG and IgM anticardiolipin antibodies were measured by an enzyme linked immunosorbent assay (ELISA), and simultaneous plasma samples tested for lupus anticoagulant activity. The most significant association with clinical events (previous thrombosis or thrombocytopenia occurring in 11/58 patients) was with prolonged tissue thromboplastin inhibition time (TTT) followed by prolonged kaolin cephalin clotting time (KCCT) then raised IgG anticardiolipin antibody concentrations and dilute Russell's viper venom time. Although IgG anticardiolipin antibodies or KCCT were the most sensitive tests in identifying this group, the TTT was the most specific (98%). Nine patients were IgG anticardiolipin antibody positive and lupus anticoagulant negative, of whom one had thrombocytopenia but none had thrombosis. The presence of a lupus anticoagulant in anticardiolipin antibody positive patients increases specificity for certain adverse clinical events.

Anticardiolipin antibodies identify a subgroup of patients with systemic lupus erythematosus with recurrent arterial and venous thrombosis, thrombocytopenia, recurrent pregnancy loss, other clinical features occurring in this group include cerebrovascular disease and livedo reticularis. Patients with high IgG anticardiolipin antibody positive and lupus anticoagulant negative, of whom one had thrombocytopenia but none had thrombosis. The presence of a lupus anticoagulant in anticardiolipin antibody positive patients increases specificity for certain adverse clinical events.

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Lupus anticoagulant and anticardiolipin antibodies in SLE

Lupus anticoagulant antibodies 2 SD=9 GPL, 5 SD=24 GPL, 20 SD=100 GPL; for IgM anticardiolipin antibodies 2 SD=6 MPL, 5 SD=25 MPL, 10 SD=55 MPL.

LUPUS ANTICOAGULANT
Whole blood was collected into Becton Dickinson containers containing 1/10 volume of trisodium citrate. As lupus anticoagulant activity seems to be less evident in the presence of platelets the blood was centrifuged at 2500 g for 20 minutes at 4°C to obtain platelet free plasma. The plasma was stored at −20°C until tested.

In this study three phospholipid dependent coagulation tests were used. The tissue thromboplastin inhibition test (TTIT) is a modification of the one stage prothrombin time, which is a measure of the extrinsic clotting system. The kaolin cephalin clotting time (KCCT) is a modification of the activated partial thromboplastin time and is a measure of the intrinsic clotting system. The dilute Russell’s viper venom test (RVVT) is independent of the intrinsic and extrinsic systems; Russell’s viper venom activates factor X directly in the presence of phospholipid and calcium ions. In both the KCCT and the RVVT the source of phospholipid is cephalin.

KAOLIN CEPHALIN CLOTTING TIME
Patient’s plasma (0-1 ml) was incubated with 0-1 ml of Bell and Alton cephalin at standard concentration, and 0-1 ml of 0-5% light kaolin in Owen’s buffer for three minutes at 37°C. Calcium chloride (0-1 ml, 0-025 mol/l) was then added and the clotting time recorded. The presence of a lupus anticoagulant was indicated by prolongation of the KCCT, not corrected in a 1:1 mixture of patient’s and pooled normal plasma. The 1:1 mixture was tested at 10 and 60 minutes to show that the inhibitory effect was not progressive, unlike the specific factor inhibitors.

RUSSELL’S VIPER VENOM TIME
This was performed by an adaptation of the method of Austin and Rhymes. The test reagent was prepared by diluting 0-2 ml of a 1/1000 solution of Russell’s viper venom with 3 ml of a 1/8 dilution of cephalin. This combination was found to give a RVVT of 22 seconds with normal plasma. Patient’s plasma (0-1 ml) was incubated with 0-1 ml of reagent at 37°C for 30 seconds. Calcium chloride (0-1 ml, 0-025 mol/l) was then added and the clotting time recorded. The presence of a lupus anticoagulant was indicated by prolongation of the KCCT, not corrected in a 1:1 mixture of patient’s and pooled normal plasma.

TISSUE THROMBOPLASTIN INHIBITION TEST
This was performed by an adaptation of the method of Schleider et al. A 1/1000 dilution of Manchester thromboplastin (0-01 ml) in 0-9% NaCl was incubated with 0-1 ml of patient’s plasma for five minutes. Calcium chloride (0-1 ml, 0-025 mol/l) was then added and the clotting time recorded. The ratio of the patient’s clotting time to the clotting time of normal pooled plasma was calculated. The presence of a lupus anticoagulant was indicated by a ratio of >1-3, not corrected in a 1:1 mixture of patient’s and pooled normal plasma.

All the above tests were performed on a Roche Cobas fibrocoagulometer.

STATISTICAL ANALYSIS
Spearman’s rank correlation and the χ² test were used as indicated.

Results
ANTICARDIOLIPIN ANTIBODY AND LUPUS ANTICOAGULANT POSITIVITY
Seventeen patients had an initial increase of IgG anticardiolipin antibodies, 10 in the moderate range and seven in the low range. Five of these patients and four others had raised IgM anticardiolipin antibodies (six in the low range). Of the 17 patients with raised IgG anticardiolipin antibody concentrations nine were negative for lupus anticoagulant in all three assays and eight were positive in at least one assay. There was no significant difference in the median IgG anticardiolipin antibody concentration between these last two groups.

Thirteen patients were lupus anticoagulant positive on their first sample in at least one assay. Twelve patients had an abnormal KCCT; eight an abnormal RVVT, and six an abnormal TTIT. Four patients were abnormal in all three clotting assays, another five abnormal in two assays, and four patients abnormal in one assay alone.

CORRELATION BETWEEN ANTICARDIOLIPIN ANTIBODIES AND COAGULATION ASSAYS
There was a highly significant correlation (Spearman’s) between anticardiolipin antibody concentrations and all assays used for measuring lupus anticoagulant with the exception of IgM anticardiolipin antibodies and the KCCT, which just failed to reach significance (table 1). Similarly, there was a highly significant association between the individual coagulation assays.

Table 1 Relation between anticardiolipin antibodies (aCL) and coagulation assays

<table>
<thead>
<tr>
<th>IgG aCL</th>
<th>IgM aCL</th>
<th>KCCT</th>
<th>TTIT</th>
</tr>
</thead>
<tbody>
<tr>
<td>+</td>
<td>NS</td>
<td>+</td>
<td>+</td>
</tr>
</tbody>
</table>

*KCCT = kaolin cephalin clotting time; TTIT = tissue thromboplastin inhibition test; RVVT = Russell’s viper venom time.

<table>
<thead>
<tr>
<th>IgM aCL</th>
<th>KCCT*</th>
<th>TTIT</th>
<th>RVVT</th>
</tr>
</thead>
<tbody>
<tr>
<td>+</td>
<td>NS</td>
<td>+</td>
<td>+</td>
</tr>
</tbody>
</table>

* = p<0.001 (Spearman’s rank correlation); NS = not significant.

Twenty three patients had further paired serum and plasma samples taken at a later date and...
four of these were also bled on a third occasion. The figure shows the serial results for those patients positive in any one of the IgG anticardiolipin antibody or coagulation tests. Nine patients who were IgG anticardiolipin antibody positive on the first occasion all remained positive (figure A). Only one of the other 14 patients (patient A) became IgG anticardiolipin antibody positive and this at low levels. The relation between disease activity, pregnancy, and treatment and anticardiolipin antibody concentrations in a larger number of patients over a longer period is the subject of a separate study.

Nine of the 23 patients studied serially had an abnormal KCCT on at least one occasion (figure B), six of these nine had at least one abnormal TTIT (figure C), and six of these nine had at least one abnormal RVVT (figure D). Most patients who were lupus anticoagulant positive by one assay in the initial sample remained positive by that respective assay in the second or third samples. Conversely, 14 of the 23 patients studied serially were lupus anticoagulant negative in all three assays on first sampling and remained so on serial samples.

Although there were occasional discordant fluctuations between the IgG anticardiolipin antibody concentrations and the coagulation times most often any variation was in the same direction. IgG anticardiolipin antibody concentrations returned to normal in only one patient (patient D, figure A), and this patient had no evidence of a lupus anticoagulant. An initially raised KCCT in three patients became normal. In one patient (patient I, figure B) this corresponded with the start of prednisolone treatment and a fall in the IgG anticardiolipin antibody concentrations and normalisation of the TTIT. In another patient (patient F, figure B) this corresponded with an increase of prednisolone dose during a pregnancy and a fall in IgG anticardiolipin antibody concentrations and normalisation of the TTIT. In this patient a third sample taken 12 months after the pregnancy when the patient was no longer receiving prednisolone showed a return to more abnormal values in these three assays.

**THROMBOSIS AND THROMBOCYTOPENIA**

The patients were divided on the basis of the occurrence of adverse features previously described in association with antiphospholipid antibodies. Thus 11 of the 58 patients had a well reported thromboembolic event or a persistent idiopathic thrombocytopenia (platelet count below 120,000 x 10^3/l on at least two occasions not related to drug treatment), or both. Four had thrombocytopenia alone, three had had major cerebrovascular accidents, two had thrombocytopenia and a previous pulmonary embolism, one a deep vein thrombosis and a pulmonary embolus, and one a deep vein thrombosis alone.

Detection of raised IgG anticardiolipin antibody concentrations was one of the two most sensitive tests in identifying this group of patients but had the lowest specificity (table 2). This was because of a large number of 'false positive' tests, which also meant a lesser degree of significance compared with the KCCT or the TTIT. Although the specificity improved by

![Figure A](image1.png)

![Figure B](image2.png)

![Figure C](image3.png)

![Figure D](image4.png)

**Table 2** Thrombosis or thrombocytopenia, or both (11/58 patients)

|                | Sensitivity (%) | Specificity (%) | p Value
<table>
<thead>
<tr>
<th></th>
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</thead>
<tbody>
<tr>
<td>IgG aCL* &gt;2 SD</td>
<td>63</td>
<td>79</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>IgM aCL &gt;2 SD</td>
<td>36</td>
<td>87</td>
<td>NS</td>
</tr>
<tr>
<td>KCCT*</td>
<td>63</td>
<td>91</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>TTIT*</td>
<td>45</td>
<td>98</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>RVVT*</td>
<td>27</td>
<td>89</td>
<td>NS</td>
</tr>
<tr>
<td>IgG aCL+LAC+*</td>
<td>54</td>
<td>96</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

*aCL= anticardiolipin antibodies; KCCT=kaolin cephalin clotting time; TTIT=tissue thromboplastin inhibition test; RVVT=Russell's viper venom time; LAC=lupus anticoagulant.

*+LAC=lupus anticoagulant positive in any one of the three coagulation assays.

*+ Test.
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Table 3  Thrombosis/thrombocytopenia

<table>
<thead>
<tr>
<th>Positive (n=11)</th>
<th>Negative (n=47)</th>
</tr>
</thead>
<tbody>
<tr>
<td>IgG aCL+ve LAC+ve</td>
<td>6</td>
</tr>
<tr>
<td>IgG aCL+ve LAC−ve</td>
<td>8</td>
</tr>
<tr>
<td>IgG aCL−ve LAC+ve</td>
<td>1</td>
</tr>
<tr>
<td>IgG aCL−ve LAC−ve</td>
<td>3</td>
</tr>
</tbody>
</table>

*aCL=anticardiolipin antibodies; LAC=lupus anticoagulant.
†LAC+ve=lupus anticoagulant positive in any one of three coagulation assays.

Discussion

The lupus anticoagulant is an acquired inhibitor of coagulation, which has immunological reactivity towards anionic phospholipid.22 It is likely that lupus anticoagulants prolong phospholipid dependent coagulation by acting at the junction of the intrinsic and extrinsic coagulation pathways, interacting with the phospholipid portion of the prothrombinase complex, without inhibiting the activity of specific coagulation factors. This is shown by the fact that the inhibition of in vitro coagulation is immediate and does not increase progressively with incubation time.22 In vivo the lupus anticoagulant is associated with thrombosis and thrombocytopenia rather than haemorrhage, probably owing to the extra presence of platelet and endothelial surfaces, but the exact mechanism is uncertain.

The introduction of solid phase assays for measuring antiphospholipid antibodies has allowed more rapid screening of patient groups at risk of harbouring a lupus anticoagulant, and an abundance of reports confirming the aforementioned associations (reviewed in ref 9). Standardisation of the assay for measuring anticardiolipin antibody concentrations may be more easily achievable than the many tests advocated for measuring lupus anticoagulant and facilitates comparison between patient groups. In our study determination of raised IgG anticardiolipin antibody concentrations was an effective though not perfect screening test for the presence of lupus anticoagulant, which was present in eight of 13 such patients. Also raised IgG anticardiolipin antibodies and an abnormal KCCT were the two most sensitive tests in identifying a group of patients with thrombosis or thrombocytopenia. If the presence of a lupus anticoagulant is strongly suspected, however, coagulation studies should be performed in anticardiolipin antibody negative patients to exclude the presence of an anticoagulant. Five such patients were found in our study, including one patient with florid livedo reticularis and severe mitral valve disease and another with previous deep vein thrombosis.

Although there was a strong correlation between anticardiolipin antibody concentrations and coagulation times in our study this does not necessarily imply identity. Indeed anticardiolipin antibodies may be present in the absence of any apparent coagulation defect and in that case has less clinical significance. This study confirmed that the presence of a lupus anticoagulant was more specific for a group of patients with thrombosis or thrombocytopenia than was the presence of either IgG or IgM anticardiolipin antibodies. Furthermore, there was no significant difference in the median IgG anticardiolipin antibody concentration between IgG anticardiolipin antibody positive patients with or without a lupus anticoagulant. In those patients studied serially parallel changes in anticardiolipin antibody concentrations and coagulation times usually occurred, but there were also the occasional discordant fluctuations as has been previously noted.24 Closer concordance might be found if antibodies to anionic phospholipids other than cardiolipin were measured. Phosphatidylycerine, which occurs in more abundance
in platelet membranes, is a more likely in vivo target for an immune response, and has been shown to overcome specifically the effect of a lupus inhibitor in vitro.\textsuperscript{25}

Several coagulation tests have been advocated for both screening and confirming the presence of a lupus anticoagulant.\textsuperscript{2,3,15} It was not the intention of this study to find the most optimal assay for either of these purposes. We chose three representative assays that could be more conveniently performed in our routine haematology tests and that between them could detect abnormalities in intrinsic (KCCIT) and extrinsic (TTIT) clotting pathways. The RVVT, which is independent of both systems, is less likely to cause false positive results in the reported RVVT, which is independent of both systems, not the intention of this study to assay of a lupus shown either specific most sensitive recently reported all assays as laboratories methods, a standardisation may achieved until better work was achieved in the determination of IgG anti-antiphospholipid syndrome. J Rheumatol 1988; 15: 1742-6.


