Human heart sarcolemmal sheath antibodies in children with non-suppurative sequelae of group A streptococcal infections: a follow up study

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Abstract

The kinetics of the human heart sarcolemmal sheath antibody were studied in children with acute rheumatic fever who had no carditis, children with acute rheumatic fever who had carditis and developed rheumatic heart disease, and in children with acute poststreptococcal glomerulonephritis. The children with rheumatic fever and those who developed valvular heart disease were given continuous secondary antistreptococcal prophylaxis. The titre of antibody at onset was significantly higher than that of the controls in children with acute rheumatic fever and carditis and in children with acute poststreptococcal nephritis. The difference in the antibody titre between children with rheumatic fever who had no carditis and controls was not statistically significant. After a mean follow up of three years, however, a high titre was only maintained in children with rheumatic fever who developed valvular heart disease.

Heart reactive antibodies were produced in rabbits immunised by group A streptococci; these antibodies bound to the sarcolemmal sheath of the cardiac myofibres, skeletal muscle, and the smooth muscles of the vessel walls.1-3 By fluorescent absorption studies Kaplan identified two cross reactive systems in the cell wall of the streptococci: one was type specific whereas the other was universal to all serotypes.1-3 These heart reactive antibodies were different from those detected in patients with postcardiomyopathy syndrome, Chagas’ disease, Dressler’s syndrome, and in patients with rejected cardiac transplants.1-3 The heart reactive antibodies in patients with rejected cardiac transplants were absorbed only by cardiac tissue but not the streptococcal antigen, showing that such antibodies were not induced by streptococci.1-3 Zabriskie et al detected heart reactive antibodies in patients with acute rheumatic fever, acute poststreptococcal glomerulonephritis and in non-complicated streptococcal infections; the antibody was fourfold higher in patients with rheumatic fever.4 The titre declined slowly over a period of two to three years. These heart reactive antibodies were absorbed by the streptococcal membrane, however, thus differentiating them from those demonstrated by Kaplan, which were absorbed by the cell wall of the streptococci. Furthermore, the heart reactive antibodies described by Zabriskie et al in patients with acute rheumatic fever were also different from those produced in rabbits immunised by the streptococcal membrane.4 Van de Rijn et al purified heart reactive antibodies from patients with acute rheumatic fever and showed that they bound strongly to the human heart sarcolemmal sheath (HHSS), poorly to skeletal muscles, and did not bind to the smooth muscles of the vessel walls.5 To the best of our knowledge no studies have attempted to explore the kinetics of these antibodies after their purification by van de Rijn et al.5 These antibodies may have a role in the pathogenesis of acute rheumatic fever. In this paper we report our findings on the kinetics of these antibodies in children with acute rheumatic fever who had no carditis, children with rheumatic fever and carditis who developed rheumatic heart disease, and children with glomerulonephritis.

Patients and methods

PATIENTS

Four groups of subjects were studied. Group 1 comprised 15 children with an initial attack of acute rheumatic polyarthritis; group 2 consisted of 15 children with an initial attack of rheumatic fever which progressed to rheumatic heart disease. Two other groups of children, matched for age and sex with the two groups with rheumatic fever and rheumatic heart disease, were included in this study. A group of 13 children with acute glomerulonephritis (group 3) and a group of 27 normal children (group 4) served as controls (see table 1). All children were followed up for three or more years.

Groups 1 and 2 were treated monthly with benzathine penicillin G. Children of group 1, 2, and 3 were bled within 48 hours of admission to hospital and on subsequent follow up visits. All serum samples were separated and stored at −20°C until tested. Of the 15 children with rheumatic fever and who developed rheumatic heart disease, 11 had mitral incompetence, seven had aortic incompetence, the valvular lesions were single or in combination. The diagnoses of rheumatic fever and glomerulonephritis were based on internationally accepted criteria.6 7

METHODS

The heart reactive antigen was prepared using the technique described by Van de Rijn et al.5 Briefly, human heart was removed five hours after death, cleaned from fat, ground for five minutes at 4°C, and centrifuged at 14 000 g for 30 minutes. The sediment was washed three times with saline and autolysis performed by
incubating one volume of the sediment with 100 volumes of distilled water for 24 hours at 4°C. This was repeated three times, and the sediment was then suspended in 0.05 M TRIS-HCl buffer (pH 7.5) containing 1 mM Mg^{2+} and treated with 1 mg RNase/ml and 1 mg DNase/ml at 37°C for three hours. A few drops of chloroform were added as antibacterial agent, and the suspension was then centrifuged at 14 000 g for 30 minutes. The sediment was washed three times under the same conditions and lyophilised. The sheath antigen was further extracted with 1% sodium dodecyl sulphate at 100°C in a water bath for three minutes, as described by Dale and Beachey.\(^5\) The sediment was collected after centrifuging at 18 000 g for 30 minutes, and this will be referred to as the HHSS antigen. The HHSS was ultrasonically disintegrated at 23 kHz (Soniprep 150, MSE, England) in a concentration of 1 mg/ml for 2 × 10 minutes and then applied to microtitre plates as antigens.

Indirect microsolid phase radioimmunoassay was carried out on disposable microtitre plates (Cooke, England/United States), using a sandwich technique.\(^6\) Antigen was fixed at a concentration of 0.5 µg/50 µl phosphate buffered saline per well; the serum samples were diluted in phosphate buffered saline containing 1% bovine serum albumin. Specific antibodies were detected by protein A (Pharmacia, Sweden) labelled with iodine-125 (Amersham, England). Dosimetry was carried out with a gammacounter (Crystal II multidetector, RIA system, Packard-Nebraa, United States). Results were calculated as counts per minute and compared with negative and positive serum standards. Positivity was expressed in dilution titles. Each specimen was tested in fourfold dilution ranging from 40 to 40 960. The antestreptolysin O and anti-DNAse B antibody titres were determined as described previously.\(^10\)\(^11\)

Statistical analysis for differences in distribution was by the Mann-Whitney U rank test.

**Results**

The upper limit of the normal titre of HHSS antibody was taken as the titre exceeded by no more than 20% of the normal controls. The log of the geometric mean titre of the upper limit of normal was thus taken as 2.4 (see figure). Table 1 shows the basic data of the four groups. Table 2 shows the logs of the geometric mean titre of the antibodies in the four groups at the onset and after three or more years. Table 3 shows the statistical differences of the antestreptolysin O titre and anti-DNAse B antibodies in the four groups at onset. Table 4 shows the statistical differences of the HHSS antibody in the four groups at onset and at three or more years. The figure shows the distribution of HHSS antibody titres in the three groups at onset and at three or more years and also in the controls.

Table 3 Statistical differences in the logs of the geometric mean titre of antestreptolysin O titre and anti-DNAse B at onset

<table>
<thead>
<tr>
<th>Group</th>
<th>p Value at onset</th>
</tr>
</thead>
<tbody>
<tr>
<td>RHD v control</td>
<td>0.0001 0.01</td>
</tr>
<tr>
<td>ARF v control</td>
<td>0.0001 0.003</td>
</tr>
<tr>
<td>AGN v control</td>
<td>0.0001 0.02</td>
</tr>
</tbody>
</table>

For abbreviations see table 2.

Table 4 Statistical differences in the logs of the geometric mean titre of human heart sarcolemmal sheath antibodies at onset and at ≥3 years

<table>
<thead>
<tr>
<th>Group</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>RHD v controls</td>
<td>0.002*</td>
</tr>
<tr>
<td>ARF v controls</td>
<td>0.1</td>
</tr>
<tr>
<td>AGN v controls</td>
<td>0.05*</td>
</tr>
<tr>
<td>RHD v ARF</td>
<td>0.0001 0.03*</td>
</tr>
<tr>
<td>RHD v AGN</td>
<td>0.14 0.2</td>
</tr>
<tr>
<td>AGN v ARF</td>
<td>1.0 0.3</td>
</tr>
</tbody>
</table>

*Significant difference.
†For abbreviations see table 2.
Discussion

Evidence has accumulated over the past 30 years suggesting that the cross reactive antigens in both heart and streptococci are membrane associated. Through the purification of the heart reactive antibodies (HHSS antibodies) described by Zabriskie et al. the cross reactive streptococcal membrane antigen was shown to be composed of four polypeptide chains with a molecular weight ranging between 22,000 and 32,000 daltons.  

Data from our study show that the titres of the HHSS antibody at onset were significantly higher in patients with rheumatic heart disease than in controls (tables 2 and 4), but the titres were not significantly different between the group with rheumatic heart disease and the other two groups. At three or more years, however, the titres of the HHSS antibody were significantly higher in the group with rheumatic heart disease than the group with acute rheumatic fever (tables 2 and 4). In fact the titre in patients with rheumatic heart disease had increased between onset and \( \geq 3 \) years, whereas it decreased in the group with rheumatic fever. This in an important finding as both groups were receiving continuous antistreptococcal secondary prophylaxis, and showed that a persistently high titre of the HHSS antibody was only maintained in patients with rheumatic fever who developed rheumatic heart disease. This behaviour is similar to that of the group A specific polysaccharide antibody.  

The titre of the HHSS antibody in the group with glomerulonephritis was higher than that in the group with rheumatic fever at the onset, though the difference was not statistically significant. This is a surprising finding and not in agreement with the results of Zabriskie et al. and van de Rijn et al. The titre of the HHSS antibody at three or more years was markedly higher in the group with glomerulonephritis than in the group with rheumatic fever (table 2), but the difference was not statistically significant. For interpretation of these findings, however, it should be stressed that the groups with rheumatic fever and rheumatic heart disease were receiving continuous antistreptococcal prophylaxis, whereas the group with glomerulonephritis was not. This is shown by the changes in the titres of antistreptolysin O and anti-DNA\(_B\) antibodies. The titres of these two antibodies at the onset were significantly higher in the three groups than in the controls (tables 2 and 3). At three or more years after the onset the titres of these antibodies in groups with rheumatic fever and rheumatic heart disease were less than those at the onset (tables 2 and 3).

On the other hand, the changes of the titres of these two antibodies in the group with glomerulonephritis were minimal; furthermore, the titres at three or more years were higher than those in the group with rheumatic fever and rheumatic heart disease. Possibly, in the group with glomerulonephritis, who were not receiving prophylaxis, streptococcal infections might have contributed to the rise of the HHSS antibody titre. This may be similar to the situation in which streptococcal infections which resulted in recurrences of rheumatic fever were reported to cause a rise in the HHSS antibody titres.  

In conclusion, the finding that a high titre of the HHSS antibody in patients with acute rheumatic fever was only maintained in those who developed valvular heart disease is significant as this antibody binds to the HHSS. Further studies with longer periods of follow up would certainly be worthwhile.

Supported by a grant MK015 from the University of Kuwait.

6 Ad hoc committee to revise the Jones criteria (modified) of the council on rheumatic fever and congenital heart disease, of the American Heart Association: Jones criteria (revised) for guidance in the diagnosis of rheumatic fever. Circulation 1983; 69: 204A.