Synovial fluid examination for the diagnosis of synovial amyloidosis in patients with chronic renal failure undergoing haemodialysis

J MUÑOZ-GÓMEZ, R GÓMEZ-PÉREZ, M SOLÉ-ARQUES, AND E LLOPART-BUISÁN

From the Departments of 1Rheumatology and 2Pathology, Hospital Clínico y Provincial, Barcelona, Spain.

SUMMARY The diagnosis of synovial amyloidosis is based upon synovial biopsy. Synovial fluid (SF) in seven patients with amyloid arthropathy associated with chronic renal failure undergoing haemodialysis were studied. The SF and synovial samples of 10 consecutive patients with seronegative mono- or oligoarthritis served as a controls. Six of the seven patients with amyloid positive synovial biopsy specimens showed amyloid in their SF. No amyloid was found in the synovial tissue or fluid of the 10 patients in the control group, the sensitivity being 87.7%. The finding of amyloid in SF was highly reproducible, showing its presence in the same joint on several occasions. The deposits were Congophilia resistant to potassium permanganate pretreatment, and the immunohistochemical analysis proved that they contained β2 microglobulin. The high sensitivity and good reproducibility of the method shows that the finding of amyloid in SF is sufficient for the diagnosis of synovial amyloidosis. It is possible to perform immunochemical analysis on the SF sediment. Amyloid arthropathy can therefore be added to the list of conditions in which synovial fluid examination can be clinically helpful.

Key words: β2 microglobulin, potassium permanganate, amyloid arthropathy, immunohistochemical analysis.

The diagnosis of synovial amyloidosis is based upon the demonstration of amyloid in synovial tissue. In 1973 Gordon et al showed the presence of amyloid in small fragments of villi in the synovial fluid (SF) in three patients with rheumatoid-like polyarthritis, who were in fact affected by amyloid arthropathy associated with myeloma. Since then no new publications have appeared confirming the usefulness of the method. We have studied a group of patients with chronic renal failure and amyloid arthropathy undergoing periodical haemodialysis in order to relate SF findings to those of synovial biopsy.

Materials and methods

We have studied the SF in seven patients with chronic renal failure undergoing dialysis (four men, three women; mean age 51 years; range 33–62 years) diagnosed as suffering from synovial amyloidosis by means of synovial biopsy. The period of haemodialysis varied from five to 14 years, mean nine years (Table 1). The aetiology for their chronic renal failure was glomerulonephritis (two patients), back flow hydronephrosis (one patient), nephroangiokeratoma (one patient), chronic nephritis (one patient), and unknown (two patients). No patient had renal amyloidosis. Ten consecutive patients (three men, seven women; mean age 50 years; range 30–75 years) with seronegative mono- or oligoarthritis of more than one year's duration, in whom synovial biopsy had been performed for diagnostic purposes, served as a controls.

Synovial samples and centrifuged synovial fluid sediments were fixed in 10% formal saline, embedded in paraffin, and the sections stained with Congo red and studied under polarising microscopy for the green birefringence typical of amyloidosis (Fig. 1). The samples from patients with chronic renal failure who were undergoing dialysis were treated with potassium permanganate by Wright's technique.
Synovial amyloidosis diagnosed by synovial fluid examination

Table 1

<table>
<thead>
<tr>
<th>Patient</th>
<th>Age (years)</th>
<th>Sex</th>
<th>Dialysis (years)</th>
<th>Amyloid Synovial membrane</th>
<th>Amyloid Synovial fluid</th>
<th>Potassium permanganate synovial fluid*</th>
<th>β₂ Microglobulin</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>59</td>
<td>M</td>
<td>9</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>2</td>
<td>48</td>
<td>M</td>
<td>10</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>ND</td>
</tr>
<tr>
<td>3</td>
<td>47</td>
<td>M</td>
<td>5</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Wrist Hip (also capsule)</td>
</tr>
<tr>
<td>4</td>
<td>54</td>
<td>M</td>
<td>8</td>
<td>+</td>
<td>+</td>
<td>Insufficient quantity</td>
<td>ND</td>
</tr>
<tr>
<td>5</td>
<td>62</td>
<td>F</td>
<td>9</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>6</td>
<td>33</td>
<td>F</td>
<td>14</td>
<td>+</td>
<td>-</td>
<td>ND</td>
<td>+</td>
</tr>
<tr>
<td>7</td>
<td>55</td>
<td>F</td>
<td>8</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td></td>
</tr>
</tbody>
</table>

*+ indicates Congophilia resistant to pretreatment with potassium permanganate. ND = not done.

Immunohistochemical analysis was carried out by the peroxidase-antiperoxidase method and by the avidin-biotin technique using anti-β₂ microglobulin antibody (DAKO). For statistical analysis the P4F program of the BMDP (California University) was used and proportions were compared with the McNemar test.

Results

In six of the seven patients with amyloid positive synovial biopsy specimens the SF showed the presence of amyloid (Table 1). No amyloid was found in the synovial tissue or fluid in the 10 patients in the control group.

In our patients the sensitivity of the study of SF with respect to synovial biopsy specimen was 87.7%. In patient No 6 (amyloid positive in synovial tissue and negative in fluid) the amount of SF was small (2 ml), which might explain the absence of villous fragments; nevertheless, in other patients not included in this series we have found amyloid in similar amounts of SF. There were no statistically significant differences between the findings of the synovial biopsy and those of the SF (McNemar test).

The finding of amyloid in SF proved to be highly reproducible because on examining different samples of SF from the same biopsied joint over long periods of time (average eight months) amyloid was always found to be present (Table 2). The examination of other affected joints in the same patient always showed amyloid in the SF (17 samples from six patients), thus showing that the articular disorder was generalised (Table 2).

The amyloid showed a Congophilia resistance to treatment with potassium permanganate in the synovial fluid sediments of all the patients studied, a property apparently shared with AL amyloid (Fig. 1).
The immunohistochemical analysis confirmed that the amyloid deposits of synovial fluid sediments contained β2 microglobulin. In addition, β2 microglobulin was demonstrated in synovial tissue (knee, wrist, hip) in the three patients in whom it was studied.

**Discussion**

Gordon *et al* showed similar findings in amyloid arthropathy associated with myeloma. In their study amyloid was found in the synovial tissue of the right wrist in three patients and in the SF of the joints studied (first patient, right wrist and right knee; second patient, left shoulder and both knees; third patient both shoulders). In our cases the amyloid arthropathy was associated with patients with chronic renal failure undergoing haemodialysis. It seems likely that amyloid would be found in the SF in any type of synovial amyloidosis. Recently Gejyo *et al* have found that amyloid laden tissue, obtained from a patient with chronic renal failure undergoing dialysis and with carpal tunnel syn- drome, contained β2 microglobulin, suggesting that amyloid arthropathy associated with haemodialysis is due to the presence of β2 microglobulin. This fact has been confirmed by immunohistochemical analysis by others, and we have also been able to demonstrate it in the SF sediment of our patients.

The high sensitivity (88%) of SF examination with regard to the synovial biopsy and the good reproducibility of the method (showing the presence of amyloid in the SF in the same joint on several occasions) show that the finding of amyloid in synovial fluid is sufficient for a diagnosis of synovial amyloidosis to be made. Immunohistochemical analysis of SF sediment may be useful to demonstrate the presence of β2 microglobulin, AA amyloid, and both κ and λ light chains. Amyloid arthropathy can therefore be added to the list of conditions where synovial fluid examination can be clinically useful.

We would like to thank Dr A St J Dixon, Bath Institute for Rheumatic Diseases, Bath, England, for kindly reviewing the manuscript.

**References**