Circulating yersinia specific immune complexes after acute yersiniosis: a follow up study of patients with and without reactive arthritis

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SUMMARY The occurrence of immune complexes (ICs) containing Yersinia enterocolitica antigen and immunoglobulin was studied in 216 serum samples from 51 patients with recent yersiniosis at an early stage of the disease and during a follow up of two to 12 months. Twenty eight of the patients developed reactive arthritis. 23 recovered from the infection without any complications. An enzyme linked immunosorbent assay (ELISA) recently developed for detection of ICs containing yersinia antigen(s) and IgM, IgG, or IgA was applied. During the first two months after onset of the infection yersinia specific IgM complexes were demonstrated in higher concentrations in patients with arthritis than in those not developing this postinfection complication (p<0.02). The difference in the occurrence of IgM complexes between the two groups decreased with time. Yersinia specific IgA complexes were demonstrable in four patients with arthritis and in none of the non-arthritic patients. Yersinia specific IgG complexes were occasionally present in both patient groups. Altogether, more than eight months after onset of the infection yersinia specific ICs were detected in six arthritic and two non-arthritic patients, suggesting that in certain individuals yersinia may hide after the initial infection for prolonged periods.

It has been suggested that immune complexes (ICs) may have a role in the pathogenesis of reactive arthritis, particularly since the symptoms of post-infection complications to some extent resemble those seen in IC diseases.1 In a patient with polyarthritis after salmonella infection the level of circulating ICs correlated with severity of the disease.2 There is also evidence of certain similarities between reactive arthritis and IgA mediated glomerulonephritis, suggesting that the same pathogenetic mechanisms may be active in both. IgA nephropathy has been reported in two patients with ankyllosing spondylitis and in one with Reiter’s disease.3 Forsström et al first reported IgA nephropathy connected with yersinia arthritis,4 and later Friedberg et al found yersinia antigens in the glomeruli of five patients out of eight with glomerulonephritis triggered by yersinia infection.5 Interestingly, circulating ICs containing IgA were recently found to correlate with the severity of psoriatic arthritis6 and of an experimentally induced lung injury.7 Kekomäki et al measured ICs in patients with yersiniosis using four antigen non-specific methods detecting mainly IgG complexes; circulating ICs were frequently found both in arthritic and non-arthritic patients.8 A significant disappearance of circulating ICs was seen in arthritic patients early during the follow up when compared with those with prolonged gastroenteritis. Also, Leirisalo et al found circulating, antigen non-specific ICs in patients after acute yersiniosis, irrespective of whether or not they developed arthritis.9

In the present study occurrence of ICs containing yersinia antigen was analysed in the sera of patients with acute yersiniosis at an early stage of the disease and during a follow up. Patients with reactive arthritis were compared with those having recovered from the infection without complications and with healthy controls.

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Patients and methods

Patients

All the 51 patients studied had acute infection with *Y. enterocolitica* O:3. The diagnosis was based on the considerably raised levels of antibodies to *yersinia* detected by ELISA\textsuperscript{10,11} and the clinical picture. *Y. enterocolitica* O:3 was isolated from the faeces of 20 patients. All patients had abdominal pain or diarrhoea, or both. Fever and raised erythrocyte sedimentation rate were usually present. Two of the patients were operated upon for suspected appendicitis; the appendix was not inflamed, and yersiniosis was diagnosed both bacteriologically and serologically. Twenty eight patients developed reactive arthritis and 23 recovered without complications. The follow up period ranged from three to 12 months in 25 patients with arthritis, and in 17 patients without arthritis. From three patients with arthritis and from six patients without arthritis samples were taken only during the first two months. The follow up time was divided into periods of two months; the sample giving the highest IC value was chosen to represent each patient during each period. The patients were divided according to the grade of arthritis into three groups: (a) patients who had only gastroenteritis without any postinfection complications, (b) patients who had grade + arthritis with their own recording of joint symptoms or mild synovitis verified by the physician, and (c) patients with severe grade++ arthritis verified by the physician. Distribution of HLA-B27, age, and sex of the patients is shown in Table 1.

Two series of patient sera were studied. The first series consisted of 69 samples from 15 patients (eight women and seven men), which had earlier been studied for occurrence of antigen non-specific ICs by Kekomäki et al.\textsuperscript{8} Of these patients, nine had gastroenteritis only and six developed arthritis. The sera had been stored for six years at –20°C without thawing. To take into account the effect of long storing period on the results sera from 30 healthy blood donors which had been stored for the same time and under the same conditions were studied. The other series included 147 serum samples from 36 patients with yersiniosis, 22 of them with arthritis and 14 with gastroenteritis. The sera were stored at –20°C for no longer than two years and not thawed more than three times before the final experiments. The effect of thawing has been discussed in a previous paper describing the methodology in closer detail.\textsuperscript{12} Sera from 50 healthy blood donors were used as controls (Table 1).

Detection of *Yersinia* specific immune complexes

We have recently developed an ELISA for the detection of Ig class specific, *yersinia* ICs, which is described in detail elsewhere.\textsuperscript{12,13} In this assay, *yersinia* specific ICs were captured via the antibody to polystyrene microtitre plates by rabbit antihuman immunoglobulins, and the existence of *Y. enterocolitica* O:3 antigens was demonstrated using Fab fragments of alkaline phosphatase conjugated antibody against the same serotype (anti-yersinia (Fab)-AP conjugate). To standardise the test each plate included three controls: a positive serum with high amount of *yersinia*-immunoglobulin complexes of all three immunoglobulin classes, buffer (5% normal rabbit serum–PBS), and a pool consisting of sera from 50 healthy blood donors.

The results were expressed as enzyme immunoassay units (EIUs) as follows:

\[
\text{EIU}_{\text{sample}} = \frac{\text{OD}_{\text{sample}} - \text{OD}_{\text{buffer}}}{\text{OD}_{\text{positive standard}} - \text{OD}_{\text{buffer}}} \times 100
\]

where OD is the optical density.

EIUs of 50 normal sera were measured separately and mean values and standard deviations were calculated. EIUs over 2 SD added to the EIUs of pooled normal control sera included on each plate were taken as positive. Grade of positivity was determined as follows: + with EIUs 2 SD to 4 SD above the mean of normal controls, ++ with values 4 SD to 6 SD, and +++ with EIUs over 6 SDs above the mean of normal controls.

Specificity of binding of anti- *yersinia* (Fab)-AP conjugate to the samples

The specificity of anti-yersinia (Fab) has earlier been shown by inhibiting its binding to *Y. enterocolitica* O:3 coated wells by several Gram negative bacteria.\textsuperscript{13} In the present work the specificity was further controlled by inhibiting its binding to the ICs detected. For this purpose a serum highly positive for *yersinia* specific ICs of IgM, IgG, and IgA classes was used. The serum was used in the ELISA as described above, except that *Y. enterocolitica* O:3 bacteria were added in different concentrations simultaneously with the anti- *yersinia* (Fab)-AP conjugate.

<table>
<thead>
<tr>
<th>Grade of arthritis</th>
<th>Men</th>
<th>Women</th>
<th>Age* (years)</th>
<th>HLA-B27 +</th>
<th>–</th>
</tr>
</thead>
<tbody>
<tr>
<td>++</td>
<td>12</td>
<td>7</td>
<td>35 (2-62)</td>
<td>15</td>
<td>4</td>
</tr>
<tr>
<td>+</td>
<td>5</td>
<td>4</td>
<td>36 (22-50)</td>
<td>1</td>
<td>8</td>
</tr>
<tr>
<td>–</td>
<td>10</td>
<td>13</td>
<td>30 (14-72)</td>
<td>6</td>
<td>17</td>
</tr>
<tr>
<td>Normal controls</td>
<td>25</td>
<td>25</td>
<td>39 (19-64)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Values are mean (range).
Determination of Rheumatoid Factors
Rheumatoid factors were determined in all sera positive for yersinia specific ICs by both the Waaler-Rose test and an ELISA for IgM rheumatoid factors.14

Detection of Antigen Non-Specific Immune Complexes
Two methods of detecting antigen non-specific ICs, consisting mainly of IgG, were used. Platelet labelled staphylococcal protein A test (PIPA) was performed as described by Kekomäki and Penttinen15 and conglutinin binding ELISA (KgB-ELISA) according to Wager and Lindström.16

Detection of Anti-Yersinia Antibodies
Antibodies to Y enterocolitica O:3 were measured by an ELISA.10 11

Tissue Typing
The peripheral blood lymphocytes were typed for HLA-B27 antigen by a two stage microlymphocytotoxicity test (Histognost-B27, Behring Institut, Behringwerke AG, Marburg, W Germany).

Statistical Analysis
The statistical significance of the differences in the amount of yersinia specific ICs between patients with and without reactive arthritis was evaluated with Mann-Whitney’s U test.

Results
Yersinia Specific Immune Complexes
Since differences were not found in the occurrence of yersinia specific ICs between the patients with grade + and grade ++ arthritis, they are here presented as one group.

Yersinia specific IgM complexes
During the first two months after onset of the infection yersinia-IgM complexes occurred in the sera of 10 of 23 arthritic patients, and five sera were highly (++, ++++) positive (Fig. 1). Four of 23 non-arthritic patients were positive for yersinia specific IgM complexes; one was highly positive (++). The difference between arthritic and non-arthritic patients was significant (p<0.02). When followed up further, high values seen in patients with arthritis decreased and the difference between the two groups diminished (Fig. 1). At four to six months, however, six of 20 patients with arthritis and only one of seven patients without arthritis had IgM complexes. At eight to 12 months four arthritic and two non-arthritic patients had IgM complexes. (One arthritic patient had IgM complexes both at eight to 10 and at 10 to 12 months, see Fig. 1.) In addition, the complexes were still found in one arthritic patient at 15 months after onset of the infection. Altogether, yersinia-IgM complexes were found in 12 of the 28 (43%) patients with arthritis and in four of the 23 (17%) patients without arthritis.

Yersinia specific IgG and IgA complexes
Yersinia-IgG complexes were detected in three of 28 (11%) arthritic patients and three of 23 (13%) non-arthritic patients. During the first two month follow up four of 28 (14%) patients with arthritis had yersinia-IgA complexes, three of them in high amounts (++, ++++). One of the patients still had a positive level of IgA complexes at 10–12 months. Non-arthritic patients had no IgA complexes.

HLA-B27
Occurrence of yersinia specific ICs of any Ig class did not correlate with the occurrence of HLA-B27 (Table 2).

Individual follow up
For an individual presentation two patients were chosen from both the arthritic and non-arthritic group on the basis of having the highest level of yersinia specific ICs within two months of the onset of infection (Fig. 2). All four still had detectable levels of yersinia-immunoglobulin complexes at...


Table 2  Distribution of yersinia specific ICs in patients according to HLA-B27

<table>
<thead>
<tr>
<th>Grade of arthritis</th>
<th>Yersinia specific ICs</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive</td>
</tr>
<tr>
<td></td>
<td>HLA-B27+</td>
</tr>
<tr>
<td>++</td>
<td>8*</td>
</tr>
<tr>
<td>+</td>
<td>1</td>
</tr>
<tr>
<td>-</td>
<td>1</td>
</tr>
</tbody>
</table>

*Values show number of patients with yersinia specific ICs.

10–12 months after the onset of infection. Interestingly, both of the patients with arthritis had prolonged joint symptoms during the year. The patients without arthritis had not suffered from any symptoms after recovering from the gastroenteritis.

**Specificity of Binding of Anti-Yersinia (Fab)-AP Conjugate to the Samples**

Addition of *Y. enterocolitica* O:3 bacteria in concentrations of 60 µg/ml or more simultaneously with anti-yersinia (Fab) conjugate resulted in clear inhibition of the highly positive values given by an uninhibited sample. The inhibition was observed in the assays of yersinia-specific ICs of all three Ig classes. These findings, together with previous experiments, confirm the specificity of the binding of the anti-yersinia (Fab)-AP conjugate.

**Determination of Rheumatoid Factors**

All the sera positive for yersinia specific ICs were negative for rheumatoid factors.

**Yersinia Specific ICs Compared with ICs Detected by Antigen Non-Specific Methods**

Fourteen samples, each from different patients, had earlier been analysed for the occurrence of circulating ICs by two antigen non-specific methods, KgB-ELISA and PIPA. Yersinia-IgM complexes were detected in three of these 14 sera. KgB-ELISA gave a positive result in all three and PIPA in one. Yersinia-IgG complexes were observed in one non-arthritic patient, who had also ICs detected by non-specific methods, while yersinia-IgA complexes were found in none of the samples. Among the sera negative for yersinia specific complexes of any isotype, ICs were detected by one or both of these non-specific methods in nine cases. In sera of two patients ICs were not detected by any of the methods.

The reason for the difference in the occurrence of ICs detected by antigen specific and antigen non-specific methods might be that (a) sera with a positive result in antigen non-specific assays, but

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**Fig. 2  Follow up of yersinia specific ICs in the sera of four patients.** Two patients with the highest level of yersinia specific ICs at the early stage of the disease were chosen from both the arthritic (A+) and non-arthritic (A−) group. Yersinia-IgM complexes (□), IgG complexes (○), and IgA complexes (△) are presented separately.
negative for yersinia specific complexes, do not contain yersinia antigen within the complexes, or
that (b) our antiserum against yersinia does not recognise all yersinia antigens in ICs; our antiserum
was raised by using bacteria grown at room tempera-
ture, which does not favour expression of the plasmid associated proteins.

COMPARISON OF THE OCCURRENCE OF
YERSINIA SPECIFIC ICs AND IgA CLASS
ANTI-YERSINIA ANTIBODIES
Since IgA-anti-yersinia antibodies are known to persist even for years in patients developing
arthritis,10 11 17 they were studied in the samples
taken more than eight months after the onset of
infection. IgA class antibodies were found in seven
out of eight patients with prolonged occurrence of
yersinia-IgM complexes, the eighth patient without
persisting IgA had IgA complexes. IgA-anti-yersinia
antibodies were also observed in 11 arthritic patients
and in one non-arthritic individual, who had no
yersinia specific complexes. Tissue typing of the
eight patients with prolonged occurrence of yersinia
specific ICs showed that three of the six arthritic
patients were HLA-B27 positive and the two non-
arthritic patients were HLA-B27 negative.

CLINICAL FEATURES IN PATIENTS WITH
PERSISTING ICs
The six patients with arthritis and with persisting ICs
showed some interesting features. At the early stage
of the infection their erythrocyte sedimentation
rates were 101, 87, 75, 57, 50, and 2 mm/h, whereas
values of 24 and 12 mm/h were recorded for the two
patients without arthritis. Five of the six arthritic
patients had prolonged joint symptoms over eight
months after the onset of the infection. One of them
had to be pensioned off owing to development of
severe chronic arthritis. In another ankylosing
spondylitis is suspected. One patient had developed
chronic persisting hepatitis; no possible aetologic
background other than yersinia infection has been
found. A patient with grade ++ arthritis lasting up
to one year has thereafter had mild intermittent
arthralgia. One of the patients has suffered from
polyarthritis for over two years.

Only two of the eight patients with persisting ICs
had not received antibiotics; they both were in the
arthritic group.

Discussion
Our findings show that patients developing arthritis
after yersinia infection have significantly more
(p<0·02) yersinia specific IgM complexes than
patients without this postinfection complication.
The difference applies to the first two months after
onset of the infection and disappears thereafter.
Also Kekomäki et al found that antigen non-
specific ICs as measured by PIPA decreased
significantly with time in patients with arthritis.8 In
the present series yersinia specific IgA complexes
were detected in four out of 28 patients with
arthritis, but in none of the 23 patients without
arthritis.

There are very few examples in the literature of
ICs containing a known antigen during a follow up.
HBsAg-IgM complexes, however, have been shown
to persist in the patients with a chronic liver disease
after acute viral, type B hepatitis. Such complexes
were described as an early marker of chronic
evolution; HBsAg-IgA or -IgG complexes were
not found in these patients.18 19 Inman et al demon-
strated a 60 000 dalton component specific for
Lactobacillus casei in ICs of a patient with infective
docarditis up to one month after onset of the infection.20

There may be several explanations for the differ-
ence in the occurrence of yersinia specific ICs in
patients with and without arthritis. Better absorp-
tion of the antigen through the gut of patients with
arthritis would lead to increased levels of yersinia
specific circulating complexes. Patients with arthritis
often have only a mild diarrhoea, if any at all,21
which would mean ineffective bacterial elimination
and increased penetration of the antigen from the
gut. The patients with arthritis might also have
increased permeability of the gut leading to higher
levels of yersinia specific ICs, like patients with
ankylosing spondylitis, who were recently reported
to have a significant increase in intestinal permea-
bility when compared with the controls.22

The properties of the bacterial strain itself could
also be responsible for higher levels of yersinia
specific complexes observed in patients with
arthitis. One possibility is molecular mimicry, similarity
of the antigen to the patient's own structures, which
could lead to an ineffective elimination of the
antigen not recognised as foreign. Cross reactivity
between HLA-B27 and certain antigens of arthritis
causing bacteria has recently been shown by the use
of monoclonal antibodies.23 24 A hypothesis of cross
tolerance had earlier been suggested on the basis of
cross reactions found between klebsiella and HLA-
B27 positive cells from patients with ankylosing
spondylitis.25 In our study, however, a high level
and prolonged occurrence of yersinia specific com-
plexes were demonstrated both in HLA-B27 posi-
tive and negative patients (Table 2).

Prolonged IC formation was observed both in
arthritic and in non-arthritic patients and in both
HLA-B27 positive and negative individuals. Even
more than eight months after onset of the infection.
yersinia specific complexes were demonstrated in six patients with arthritis and in two without arthritis. The two non-arthritic patients were those who showed the highest level of yersinia-IgM complexes among the non-arthritic patients at an early stage of the disease. Five of the six patients with arthritis had also persisting IgA class anti-yersinia antibodies, and the one without detectable IgA class antibodies had IgA complexes. Interestingly, five of the arthritic patients still had prolonged joint symptoms one year after onset of the infection. Also the two non-arthritic patients with prolonged occurrence of yersinia specific complexes had persisting IgA anti-yersinia antibodies, which are not usually seen in patients without arthritis. These eight patients with prolonged occurrence of yersinia specific ICs together with persisting IgA are demonstrative examples of the possibility that yersiniae or some parts of it hide within the patient’s body for prolonged periods.

There were also several cases where our test for yersinia specific ICs was negative, possibly owing to the attachment of free antibodies to antihuman immunoglobulins bound to the solid phase, thereby preventing detection of ICs. The ability of the method to detect ICs of all three Ig classes has earlier been demonstrated, however, with preformed ICs. Also, sera with yersinia specific ICs had a high level of anti-yersinia antibodies. There were only two exceptions: in two patients a high level of yersinia-IgM complexes was observed at an early stage of the disease in the absence of anti-yersinia antibodies, which later on became detectable. Therefore, demonstration of yersinia specific ICs may also have diagnostic significance at an early stage of the yersinia infection.

Prolonged occurrence of yersinia specific ICs in eight patients together with the persisting IgA anti-yersinia antibodies possibly indicates persistence of the antigen within the patient. The role of yersinia specific IgM complexes at an early stage of the disease and their significance in the pathogenesis of the arthritis will be studied further.

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