IgE and IgE-rheumatoid factors in circulating immune complexes in rheumatoid arthritis

K. MERETEY, A. FALUS, C. C. ERHARDT, AND R. N. MAINI

From the National Institute of Rheumatology and Physiotherapy, Budapest, Hungary, and the Division of Clinical Immunology, Kennedy Institute of Rheumatology, 6 Bute Gardens, Hammersmith, London W6 7DW

SUMMARY The sera of 21 patients with rheumatoid arthritis (RA), 11 patients with systemic lupus erythematosus (SLE), and 20 healthy subjects were analysed for the presence of IgE in immune complex fractions. These fractions were isolated by polyethylene glycol precipitation and gel filtration. Thirteen sera from RA patients contained IgE immune complexes (IC) and 11 of these were from patients with extra-articular manifestations. One SLE and none of the control sera contained such material. The serum IgE level did not correlate with IgE content of the IC fractions. Higher mean serum IgE levels were found in RA patients with extra-articular complications than in controls or RA patients with joint disease only, but the differences did not reach statistical significance. IgE anti-rabbit IgG (IgE rheumatoid factors) could be demonstrated in some IgE positive IC fractions. Antibodies to IgE, in 2 instances characterised as belonging to IgG class, were also found in ICs. This suggests the presence of anti-IgE complexes. It is suggested that IgE, including some with rheumatoid factor activity, is contained in complexes which may be involved in some extra-articular manifestations of RA.

Immune complexes (ICs) that occur in rheumatoid arthritis (RA) are heterogeneous in respect of both size and composition. The ICs in RA consist mostly of IgG, IgM, and complement factors. Rheumatoid factors are also regularly described in these ICs and may be involved in the evolution of systemic manifestations.

In a recent study antinuclear antibodies of IgE type were found to occur in a complexed form in the sera of RA patients. IgE-type complexes have also been described in the sera of atopic patients.

A double polyethylene glycol (PEG) precipitation test was developed to detect IgE in the immune-complex-rich fraction isolated from sera. By this technique IgE-containing complexes were found in some RA patients.

The present studies were undertaken in order to characterise these IgE-type ICs in rheumatoid arthritis and to correlate them with clinical manifestations.

Materials and methods

Patients. Twenty-one RA and 11 SLE patients, as well as 20 healthy persons, were included in this study. All patients with RA had classical or definite RA, and patients with SLE had 4 or more ARA criteria. Fourteen RA patients had extra-articular manifestations (nodules, pulmonary disease, vasculitis and neuropathy) and were evaluated as a separate group.

Sera were obtained in the active phase of the disease (elevated erythrocyte sedimentation rate and clinical assessment), and aliquots were stored at −70°C until used.

Isolation of immune complexes. Immune-complex-rich fractions were prepared as described previously. Briefly, sera were diluted with 3% PEG (PEG 6000, Fluka, Germany) to a final concentration of 1:50. After mixing and overnight incubation at 4°C the precipitates were washed with 3% PEG and redissolved in the original volume of the serum sample. Protein content was measured by the Lowry method. IgG, IgM, and IgA levels in ICs by radial immunodiffusion, and the C3 and C4 concentrations by means of rocket electrophoresis.

Detection of IgE in ICs. We could detect IgE-type complexes by the double PEG precipitation test described previously. The redissolved PEG precipitates (50 μl) were incubated with 50 μl of 125I-anti-IgE (Phaedebas RAST Isotope Unit, Pharmacia, Sweden). The labelled anti-IgE bound to the ICs
were reprecipitated by 3% PEG (final concentration) and the pellet counted for radioactivity (Autogamma counter, Gamma Works, Hungary). Pellets with more than 3% of the added radioactivity were taken as a positive result (2 SD above the mean of the nonspecific binding in 10 normal sera and 100 µg aggregated IgG). IgE level in selected IC fractions and sera were measured by the Phadebas PRIST test (Pharmacia, Sweden). Gel filtration of the sera was performed on a Sephadex G-200 column in 0.1 M borate buffer at pH 8.1. Peaks were collected concentrated by PEG 20 000 (Serva) to the original volume and checked for IgE content by PRIST.

IgE rheumatoid factors (RFs) in ICs. Rabbit IgG-Sephadex gel was used as solid phase immunosorbent to separate RFs (a gift from F. Peterfy, HUMAN Institute for Sera and Vaccine Production, Budapest). Sera, or isolated PEG fractions, were incubated with an aliquot of the immunosorbent, and after thorough washings the amount of IgE-RF activity was determined by the subsequent binding of 125I anti-IgE.

Antibodies to IgE in PEG precipitates. Test for anti-IgE was performed on heat-treated serum or PEG precipitates. It was confirmed in separate experiments that heat treatment of sera at 56°C for 4 hours destroyed most of the immunoreactivity of IgE molecules. Such heat treated sera were incubated with a standard amount of IgE (10 IU/ml Phadebas standard), and after overnight incubation the decrease in the available IgE was determined by PRIST. IgE was simultaneously measured in the heat-treated samples. Significant decrease of the measurable IgE content after incubation with heated serum, or with IC fractions, was taken as an evidence for the presence of anti-IgE activity.

IgG was isolated from ICs by diethyl aminoethyl (DEAE) cellulose ion exchange chromatography and its anti-IgE activity measured as described above.

Statistical analysis was done by comparing the mean values by Student's t test. Serum IgE values were evaluated after e-log conversion of the results in order to compensate for the nonhomogeneity of the data.15

Results
Occurrence of complexes containing IgE in serum samples of RA and SLE patients. The isolated PEG precipitates from normal sera failed to bind significant amounts of anti-IgE (Fig. 1). The mean binding of the controls was 1.64 ± 0.18%, and none of the samples bound more than 3% of the added 125I anti-IgE. Low binding was found in SLE sera (1.99 ± 0.29). High values were observed in RA cases. In patients with extra-articular manifestations 11 out of 14 sera were positive. The extra-articular RA group differed significantly from all other groups (mean 6.8 ± 1.12, p<0.01). Patients with RA who had only joint disease showed a moderate increase in the mean value (2.49 ± 0.47% binding activity), but only 2 sera of the 7 studied showed binding higher than 3%. This group differed significantly both from normal persons and from extra-articular RA cases (p<0.05).

IgE-level of sera. We determined the serum IgE levels of the samples to compare them with the presence of IgE-complexes (Table 1). The geometric mean of sera from normal persons and RA patients

Table 1 IgE level of sera

<table>
<thead>
<tr>
<th>Groups</th>
<th>n</th>
<th>IgE KU/l*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal controls</td>
<td>20</td>
<td>59 (45–82)</td>
</tr>
<tr>
<td>RA ‘joints only’</td>
<td>14</td>
<td>100 (79–192)</td>
</tr>
<tr>
<td>SLE</td>
<td>11</td>
<td>78 (49–123)</td>
</tr>
</tbody>
</table>

*Geometric mean calculated after e-log conversion of data and range of values within 1 SE.

Table 2 Clinical features of RA and SLE patients related to the presence of IgE ICs

<table>
<thead>
<tr>
<th>RA/ SLE</th>
<th>+</th>
<th>–</th>
</tr>
</thead>
<tbody>
<tr>
<td>Nodules</td>
<td>6</td>
<td>0</td>
</tr>
<tr>
<td>Pulmonary involvement</td>
<td>6</td>
<td>0</td>
</tr>
<tr>
<td>Renal involvement</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Other features</td>
<td>4</td>
<td>0</td>
</tr>
</tbody>
</table>

*PEG fractions bind more (+) or less (-) than 3% of the added 125I anti-IgE.

Fig. 1 Occurrence of IgE-containing PEG precipitates (immune complexes). Ordinate shows the percentage binding of 125I-anti-IgE. Mean ±SEM.
with joint disease alone were the lowest. The range and the mean of IgE concentration in SLE cases and in RA with extra-articular manifestations were raised, and they showed that in some patients the IgE levels were much higher than in the other groups. However, the mean differences were not significant. The IgE levels did not correlate with the amount of IgE in ICs (r = 0.31).

The relationship of IgE-ICs to clinical features in RA and nephropathy in SLE. The extra-articular features of patients with RA are summarised in Table 2. We could conclude that extra-articular features, especially pulmonary involvement and nodules, correlated well with the presence of IgE-containing immune complexes. In contrast, patients with SLE nephropathy did not have IgE ICs in their serum.

Composition of PEG fractions in RA cases. Immunochemical analysis of PEG IC fractions revealed that IgG and IgM were the major components in all RA sera. IgA and complement (C3 and C4) were also present in some ICs. All of the components could be detected more frequently in the extra-articular RA group than in the ‘joints only’ group. The IgE content of PEG precipitates correlated with both the protein (r = 0.53) and IgG content (r = 0.62). This indicated either that IgG and IgE ICs occurred in the same patients or that Ig and IgE were present in a complexed form.

Detection of IgE RFs in immune complexes. IgE IC-positive and negative serum samples were selected and fractionated on a Sephadex G-200 column. In 3 of the 6 sera fractionated IgE was found in the excluded peak, indicating that it was in a complexed form (Table 3). The concentrated first peaks were tested for their binding to solid-phase rabbit IgG, and for IgE-class anti-IgG activity, measured by sequential binding as described. Two of the 3 IgE IC-positive peaks contained IgE-type rheumatoid factors.

PEG fractions were also similarly tested for IgE RF (rheumatoid factor) activity on the rabbit IgG immunosorbent. Of the 7 IgE-positive PEG fractions 3 contained IgE RF activity.

Antibodies to IgE in immune complexes. The first peak of Sephadex G-200 gel filtrations were heat treated to destroy their IgE, and their anti-IgE content was estimated. Anti-IgE was found in one of the 3 IgE IC-positive macromolecular peaks studied (Table 4). Among the 6 PEG fractions positive for IgE ICs 5 contained anti-IgE activity. We isolated IgG from 9 PEG IC-rich fractions by ion exchange chromatography, and 2 bound IgE.

Discussion

It has been shown that IgE-mediated reactions are important in the localisation of circulating immune complexes in the serum sickness model in rabbits. Such a role for IgE in man has not been fully investigated. Our results suggest that IgE-containing fractions of serum, presumed to be immune complexes, are found in some patients with rheumatoid arthritis, especially those with extra-articular complications. However, since IgE ICs represent only a minor fraction of the whole immune complex pool present in such sera, it is not yet possible to state whether they play a special role in the pathogenesis of tissue lesions. It is likely that IgE ICs will passively traverse abnormally permeable vessels, but we suggest that they may localise in specific sites in tissues rich in IgE receptors.

The respiratory and gastrointestinal tracts are privileged sites for IgE-production. Recent studies have shown that IgE-rich complexes are detectable in the blood of asthmatic patients and in patients with food allergy. The finding of IgE ICs in rheumatoid patients with pulmonary manifestations raises the possibility of a similar origin. Our patients were not atopic, and IgE ICs were also found in patients with nonpulmonary extra-articular manifestations. However, we cannot exclude the involvement of IgE antibodies to extrinsic antigens in the sera of our patients.

All conditions with IgE-type immune complexes are possibly associated with an increased synthesis of IgE. Though high IgE levels occur in allergy and in some infections, data on the serum IgE level in RA and in patients with RA on drugs are contradictory. A recent study has shown that serum IgE level was significantly higher in seropositive than seronegative RA, and it was suggested that drug therapy may be a contributory factor.
We found normal IgE levels in all groups, but a tendency for a higher value was observed in the extra-articular RA group. One can suggest that in patients with extra-articular symptoms an exaggerated polyclonal immunoglobulin response leads to the increased level of IgE. Indeed, it has been suggested that high IgE levels may occur in some forms of immunodeficiency as part of abnormal immunoregulation, but no such data are available in autoimmune patients.

In this study the occurrence of IgE rheumatoid factors in immune complex fractions is described. Although a novel finding, it is not in itself surprising, since other isotype have been already described. It will be important to establish whether these autoantibodies have a special clinical significance. Other IgE autoantibodies have been described, for example. IgE-type granulocyte-specific antigen-antibodies were reported in sera of patients suffering from RA or Felty’s syndrome. IgE-type anti RNA antibodies were also thought to be present in studies in which RNA could induce histamine release from basophils of RA patients.

IgM-type anti-IgE antibodies were detected in sera of healthy persons and in various patients. Our preliminary observations suggest that IgG class of anti-IgE antibodies may also be present in sera of some RA patients. While IgM-type antibodies agglutinated well, they did not release histamine. IgG type anti-IgE autoantibodies, however, might interact with cytophilic IgE and release histamine.

The finding of IgE ICs in RA contrasts with their absence in the nephropathy of SLE. Little is known about the reasons for the patterns of localisation of immune complexes in different organs in various diseases. IgE ICs and rheumatoid factors may be an additional factor which should be considered in the evolution of the subgroups of multisystem disease seen in RA.

This work was supported by the Scientific Research Council, Ministry of Health, Hungary (6-07-0304-01-2) (M). The authors wish to thank Lendvary Maria for the excellent technical assistance and Miss Carole Irving for typing the manuscript. We also acknowledge the support that the Arthritis and Rheumatism Council give to the Kennedy Institute.

References


