Abnormal lymphocyte function is secondary to drug-induced autoimmunity

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Pugh, S., Pelton, B., Raftery, E. B., and Denman, A. M. (1976). Annals of the Rheumatic Diseases, 35, 344–348. Abnormal lymphocyte function is secondary to drug-induced autoimmunity. Abnormal lymphocyte function has been frequently reported in patients with connective tissue diseases but its significance has been uncertain. Sequential studies of lymphocyte function were carried out in patients receiving the β-adrenergic blocking drug practolol (Eraldin) both before and during the development of autoimmune complications. No evidence was obtained that abnormal lymphocyte function presaged the onset of autoimmunity, and when these tests did show deficient responses these could be correlated with disease activity in general.

Abnormal tests of lymphocyte function, such as reduced numbers of T-cells and impaired responses to mitogens, have been reported in autoimmune and chronic inflammatory diseases (Lockshin and others, 1975). Although there are theoretical reasons for attributing autoimmune disease to depressed T lymphocyte function, it is not clear whether the reported abnormalities are of primary pathogenetic importance or merely secondary to disease activity.

In 1973 we described 3 cases of a systemic lupus erythematosus (SLE) syndrome induced by practolol (Raftery and Denman, 1973). Since then other important complications have been described such as the oculocutaneous syndrome (Felix, I., and Dahl, 1974; Wright, 1975) and sclerosing peritonitis (Brown and others, 1974). Moreover, the majority of patients with these side effects have serum autoantibodies (Amos, Brigden, and McKerron, 1975).

In a preliminary attempt to elucidate the pathogenesis of these complications we noted that lymphocytes from patients with the SLE syndrome had abnormal responses to mitogens in vitro, whereas no immune response to the drug itself could be detected. This suggested that sequential observations of lymphocyte function in patients receiving practolol would make it possible to determine whether abnormal lymphocyte function precedes the development of autoimmunity.

This paper shows that depressed lymphocyte transformation in patients receiving practolol could be correlated with the activity of the disease which necessitated treatment with this drug, or was induced by it. No evidence was obtained that abnormal lymphocyte function predisposed the patient to the development of autoimmunity.

Patients and methods

The patients who were studied fall into three categories. Group 1 consisted of 10 patients with definite practolol toxicity in the form of skin lesions in 6 and the oculocutaneous syndrome in the remainder. All had antinuclear antibody (ANA). These patients had already developed complications by the time they were investigated.

Group 2 The second group consisted of 122 consecutive patients with ischaemic heart disease seen at this centre as inpatients or outpatients. The incidence of ANA (titre >1:10) in this group was 23%. These patients were studied on at least two occasions over a period of 18 months. 77 patients were receiving practolol, and 45 were not receiving a beta blocking drug (see Table I).

Group 3 contained 20 consecutive patients (mean age 60 years, range 48–72) of whom 18 had recently developed angina pectoris and 2 cardiac arrhythmias. Patients who were already receiving treatment when first seen, who had an ESR >10 mm/h, or who had any associated disease, were excluded. The patients were given practolol (300–800 mg/d) but, apart from glyceryl trinitrate, received no other drugs. Blood samples were taken before starting practolol, and 1, 2, 3, 6, and 9 months after starting the drug.

Laboratory tests

Autoantibodies

Sera were examined for autoantibodies by indirect immunofluorescence using multiple block sections of
Table I  Patients in group 2

<table>
<thead>
<tr>
<th>Patients</th>
<th>No.</th>
<th>Age (years)</th>
<th>Dosage (mg/d)</th>
<th>Other drugs</th>
<th>Associated diseases</th>
</tr>
</thead>
<tbody>
<tr>
<td>On practolol</td>
<td>77</td>
<td>63 (41-80)</td>
<td>300-800</td>
<td>Glyceryl trinitrate</td>
<td>Acute infections</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Digoxin</td>
<td>Recent myocardial infarction</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Frusenide</td>
<td>Rheumatoid arthritis</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Methyl dopa</td>
<td>Hypertension</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Guanethidine</td>
<td>Diabetes</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Reserpin</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Phenformin</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Antibiotics</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Aspirin</td>
<td></td>
</tr>
<tr>
<td>Not on practolol</td>
<td>45</td>
<td>60 (39-81)</td>
<td>—</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

rat liver, stomach, kidney, and human thyroid, and rabbit antihuman immunoglobulin (Burroughs Wellcome) at 1:10 dilution.

Lymphocyte transformation

Blood mononuclear cells separated by Ficoll-Trisiosil sedimentation were cultured by a standard micromethod using Eagle's Dulbecco medium and pooled normal human serum. Dose response curves to mitogen stimulation were obtained using purified phytohaemagglutinin (PHA; Burroughs Wellcome), concanavalin A (Con A; Calbiochem), and pokeweed mitogen (PWM; Gibco) (Lance and Knight, 1974). Culture conditions were standardized throughout but the isotope used in different studies was either 14C thymidine 0.08 μCi, specific activity 62 mCi/mmol, or 3H thymidine 0.167 μCi, specific activity 150 mCi/mmol (Radiochemical Centre, Amersham) as will be indicated in the results.

Lymphocyte subpopulations

Rosette formation with sheep red cells was used as a marker for T lymphocytes (Wybran, Carr, and Fudenberg, 1972). Surface immunoglobulin detected by indirect immunofluorescence was used as a marker for B lymphocytes (Brown and Greaves, 1974). The specificity of the anti-immunoglobulin serum (Burroughs Wellcome) was verified by immuno-electrophoresis before use.

Practolol blood levels were measured by a colorimetric method.

Statistics

All analyses were performed on log transformed data.

Results

Of the first group of patients with established practolol toxicity who were studied the in vitro lymphocyte responses of the 6 patients with skin rashes were normal but those from 2 of the 4 patients with the oculocutaneous syndrome were clearly depressed (Fig. 1). This observation confirmed that some patients with practolol toxicity and associated autoimmunity develop such abnormalities.

![FIG. 1 Dose response curves from 4 patients with the oculocutaneous syndrome. Hatched area represents normal range. Unstimulated cultures 120 (50-280) dpm mean ± 2 SD. PHA = phytohaemagglutinin; Con A = concanavalin A; PWM = pokeweed mitogen](http://ard.bmj.com/)

Table II  Patients from second series divided into groups according to ESR, ANF, and whether or not they were taking practolol. The number of patients with abnormal responses (defined in text) in each group is shown together with the total number in that group

<table>
<thead>
<tr>
<th>Patients</th>
<th>ANF ESR&lt;20 mm/h</th>
<th>Negative ESR&gt;20 mm/h</th>
<th>ANF ESR&lt;20 mm/h</th>
<th>Positive ESR&gt;20 mm/h</th>
</tr>
</thead>
<tbody>
<tr>
<td>Off practolol</td>
<td>3/18 (16%)</td>
<td>7/13 (53%)</td>
<td>0/3</td>
<td>0/2</td>
</tr>
<tr>
<td>On practolol</td>
<td>6/36 (16%)</td>
<td>11/17 (64%)</td>
<td>3/13 (23%)</td>
<td>5/11 (45%)</td>
</tr>
</tbody>
</table>

Recent myocardial infarction

Rheumatoid arthritis

Hypertension

Diabetes
Accordingly, these studies were repeated in a large number of patients taking practolol in order to determine the correlation between lymphocyte function and the appearance of ANA used as a marker for the development of autoimmunity (group 2). Depressed lymphocyte responses, defined as subnormal response to one or more mitogens in at least the two lowest concentrations, were observed in a high proportion of these patients (Table II). The results were analysed with respect to disease activity, practolol treatment, and the development of ANA. Depressed responses to mitogens could only be correlated with a raised erythrocyte sedimentation rate (ESR) (Fig. 2), a test which reflects disease activity in general, but not with the presence of ANA. In addition, no correlation was found between lymphocyte function and practolol treatment with respect to dose, blood level, or duration of treatment. These findings suggested that the depression of lymphocyte function seen in patients with practolol toxicity is more likely to reflect non-specific disease activity than an event of primary pathogenetic importance.

In order to confirm this impression another group of carefully defined patients (group 3) was studied at regular intervals before and during treatment with practolol over a 9-month period. 4 of the patients were withdrawn before the end of the study with complications (Table III). With one exception none of the patients in this group showed any depression of lymphocyte transformation nor was there any

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**Table III  Patients from group 3 withdrawn before the end of study**

<table>
<thead>
<tr>
<th>Patient</th>
<th>Dose (mg/d)</th>
<th>Time on practolol (m)</th>
<th>Reason for withdrawal</th>
</tr>
</thead>
<tbody>
<tr>
<td>H.S.</td>
<td>300</td>
<td>3</td>
<td>ANA rose from 1:10-1:80. Complained of dry eyes. No signs, intercellular antibody negative</td>
</tr>
<tr>
<td>L.C.</td>
<td>300</td>
<td>3</td>
<td>ANF rose from 0 to 1:160. Complained of muscular pain</td>
</tr>
<tr>
<td>K.S.</td>
<td>600</td>
<td>6</td>
<td>Developed typical hyper-keratotoic skin lesions</td>
</tr>
<tr>
<td>K.G.</td>
<td>600</td>
<td>6</td>
<td>Developed polymyositis</td>
</tr>
</tbody>
</table>

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**Fig. 2** Mean dose response curves from patients in group 2. ○ = ESR < 20, n=63; ● = ESR > 20, n=43. Hatched area represents range from a group of normals. *P < 0.005 for all points except PHA and Con A at 2 μg where *P < 0.05 ('t' test). Unstimulated cultures 170 (20–1300) cpm mean ± 2 SD

**Fig. 3** Sequential dose response curves from patient K.G. who developed polymyositis. ● = after 3 months on practolol; ■ = after 6 months on practolol. Hatched area represents the normal range. Unstimulated cultures ● = 84 dpm, ■ = 94 dpm. (Normal range 50–280 dpm)
significant change in numbers of circulating T and B lymphocytes. However, one patient developed vague muscular aches and pains and an ANA titre of 1:10 three months after starting practolol treatment. At this time her lymphocyte response to mitogens were normal. Since her angina was poorly controlled, the dose of practolol was increased from 300 to 600 mg daily. Three months later she developed a severe polymyositis with proximal muscle weakness. Her creatinine phosphokinasen level was 1400 and the ANA titre had increased to 1:180. Her lymphocyte responses were now obviously and reproducibly abnormal. The relationship between treatment with practolol and this patient's disease is uncertain. However, its onset clearly preceded the development of abnormal lymphocyte responses (Fig. 3).

Discussion

Abnormal lymphocyte function has been described in several autoimmune and chronic inflammatory diseases (Lockshin and others, 1975) although there is little agreement about the reproducibility or significance of these findings. Some of the variation in results can be attributed to technical factors but in any event there has been no easy way of determining whether such abnormalities are of primary pathogenetic importance or the result of nonspecific disease activity. Autoimmunity has been attributed to the breakdown of a normal suppressor function mediated by T lymphocytes (Allison, Denman, and Barnes, 1971). This could follow a number of events including the destruction of T lymphocytes by virus infection or their progressive loss with age in genetically susceptible individuals. Drugs which block β-adrenergic receptors could conceivably inactivate lymphocyte populations directly. The ability of drugs which react with adrenergic receptors on lymphoid cells to modulate their activity in vitro has been frequently shown (Hadden, Hadden, and Middleton, 1970; Sherman, Smith, and Middleton, 1973). It seemed plausible, therefore, that β-adrenergic blocking drugs might selectively interfere with lymphocyte subpopulations in vivo, thereby abrogating control mechanisms which normally prevent the emergence of autoimmune disease.

Thus, serial tests of lymphocyte function in patients receiving practolol, a proportion of whom would be expected to develop side effects, gave some hope of determining whether the ablation of T lymphocyte function predisposes to autoimmune disease. In the event no evidence was obtained that selective immunosuppression precedes the development of autoimmune disease. Abnormal responses to mitogens were found in a high proportion of the patients studied but these could be correlated with disease activity in general. In contrast no correlation was found between abnormal lymphocyte function and the development of antinuclear antibody. Furthermore, autoimmune disease when it did ensue was not foreshadowed by abnormalities of lymphocyte function. Thus, this study failed to produce any evidence that drug-induced autoimmunity, at least, results from the prior eradication of T lymphocyte function. Nevertheless, lymphocyte transformation and population markers are of limited value as tests of immunosuppressive potency even in patients receiving cytotoxic drugs (Swanson and Schwarz, 1967). In vitro systems of greater physiological relevance (Platts-Mills and Ishizaka, 1975) may show that drugs such as practolol do indeed have selective immunosuppressive effects.

It is also apparent from this study that current tests of lymphocyte function cannot be used to predict the onset of autoimmune disease and other complications in patients receiving practolol and by inference, other drugs with similar side effects. Moreover, a variety of tests has failed to show that patients receiving practolol develop antibodies to the drug itself. The protein nature of these complications is of great interest with respect to spontaneous autoimmune disease since our studies show that the relationship between practolol and its side effects could not have been deduced from any obvious pharmacological or immunogenic effects of the drug. On present evidence other mechanisms must be invoked.

The authors are indebted to Dr. Stella Knight for help in standardizing lymphocyte function tests and to Dr. H. K. Adam for assaying blood levels of practolol at the ICI Laboratories, Alderley Park, Macclesfield, Cheshire.

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