METABOLISM OF CONNECTIVE TISSUE IN LIMB
ATROPHY IN THE RABBIT

BY

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In previous experiments in adult rats (Slack, 1954, 1955), massive tissue atrophy was induced by
denervation and femoral head dislocation of one hind limb. A study of the metabolism of total
collagen, total sulphated polysaccharide, and mixed muscle protein indicated that disuse atrophy does
not imply cessation of all protein or polysaccharide
synthesis. After the onset of atrophy some syn-
thesis of collagen appears to continue for at least
15 weeks. Synthesis of muscle protein continues
for about 6 weeks and of sulphated polysaccharide
for at least 2 weeks. The interesting finding was
that at least some of the normal synthetic processes
continue despite massive removal of muscle protein
and connective tissue.

These earlier experiments did not differentiate
between connective tissue closely associated with
the muscle mass, and connective tissue in the bones,
joints, and associated tendon insertions. Furthermore,
the rat experiments were open to objection because
of the unusual operative technique required
to achieve aseptic atrophy, and the fact that the
bones of the rat appear to be capable of growth
throughout much of the life of the animal. In view
of the unexpected finding of continued connective
tissue synthesis in the atrophying rat limb it was
considered desirable to repeat the experiments in
a larger animal.

In the present experimental work, to be described,
atrophy has been induced in one hind limb of
rabbits by denervation. The metabolism of con-
nective tissue collagen fractions and sulphated poly-
saccharide has been studied in two limb components:
connective tissue associated with the muscle mass
(soft tissue compartment), and that in bones, joints,
and tendon insertions (skeletal compartment).

Both the present work, and the previous experi-
ments in rats, are concerned with disuse atrophy.
Gillespie (1954) showed that, in a paralysed limb,
the main factor in atrophy is, in fact, disuse.
Diminution of blood supply is not a controlling
factor. It also appears from his work that the bony
changes in the paralysed limbs are due to
quantitative differences, and not to alterations in
quality of the bones—as judged by breaking stress,
elasticity, percentage of ash, and specific gravity.
X rays show a loss of density in disused limb bones,
described as decalcification or osteoporosis. Gilles-
pie’s work shows that these terms may be misleading.

More recently Geiser and Trueta (1958) have
described changes in the calcaneum of the rabbit,
atrophying as the result of tendo calcaneus section,
plaster immobilization, and other means. Bone
removal was found to occur up to 5 or 6 weeks after
the onset of atrophy. Thereafter there appeared
to be little further bone removal. This time-
interval corresponds to the time required for com-
plete muscle atrophy and cessation of muscle
protein synthesis in the rat experiments quoted above
(Slack, 1954). It is interesting that Geiser and Trueta,
in their experiments, found evidence of osteoblasts
at all stages of atrophy. In other words, some
synthesis of new connective tissue must be occurring
in the bone despite overall atrophy. This finding
is in agreement with the experiments on collagen
metabolism in atrophying rat limbs quoted earlier.

A considerable volume of work has been published
on changes in muscle proteins following denervation,
tenotomy, and plaster fixation. The interested
reader is referred to Helander (1957) for an excellent
review and results of original work. The impact of
atrophy on the connective tissues appears still to be
receiving less attention than it deserves. It is hoped
that the experiments to be described will, in some
measure, add to our knowledge of connective
tissue behaviour in massive tissue atrophy.

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Experimental Methods

Operative Procedure
Young adult rabbits weighing 1·8 to 2 kg. were subjected to neurectomy under ether anaesthesia and local nerve blocks. In each case an attempt was made to transect, and remove at least a distal one inch of, all motor nerves to the left hind limb. The approach was made ventrally, just lateral to the lower spine, and the sympathetic supply dissected out. Of the animals surviving operation, one-third proved to have incomplete denervation and are not included in this series. Successful operations produced a complete flail hind limb, showing obvious muscle atrophy from the hip level downwards.

Administration of Isotopes and Separation of Tissues
14 days after operation, six rabbits, selected for complete paralysis of the left hind limb, were injected subcutaneously in the upper abdomen with 10 μc. α-14C glycine per 100 g. body weight, and 200 μc. Na235SO4 per 100 g. body weight. The rabbits were then killed serially from 10 hours after isotope injection to 4 days afterwards.

The skin was stripped from both hind limbs down to the toes, which were cut off. The limbs were cut through and dislocated at the level of the femoral head. All muscle and associated fascia from each limb was removed, and this constituted the soft tissue compartment. The tissue samples were reduced to a homogeneous powder by repeated freezing in liquid nitrogen and pounding in a steel mill. After removal of aliquots for determination of total constituents the tissue fractions were separated as follows:

Collagen Fractions.—Neutral salt-soluble collagen, acidic citrate-soluble, and insoluble collagens were separated and purified, essentially as described by Jackson (1957).

The collagen content of the separated fractions was estimated by the method for hydroxyproline given below.

Sulphated Polysaccharide.—All material remaining, including supernatants, from each stage of collagen fractionation was treated as described in other experiments (Slack, 1958). The method has been shown to give good recovery of sulphated polysaccharide free from protein or hyaluronic acid (Scott, 1955). As isolated, the polysaccharides are therefore those soluble in neutral salt, acid citrate buffer, and 0·1 N NaOH, free or associated with tissue protein in the natural state. The associated protein was hydrolysed by treatment with papain to facilitate its removal.

Separated polysaccharide was estimated gravimetrically. Chemical analyses of the acetone dried preparations were done, to determine sulphate and hexosamine contents, as described below.

Chemical Analyses

Hydroxyproline.—Estimated by the Neuman and Logan method as modified by Miyada and Tappe (1956).

Hexosamine.—Determined by the method of Boas (1953).

Sulphate.—Estimated according to the method described in a previous communication (Slack, 1958).

Radioactivity Measurements

Collagen.—Preparation and isolation of 2,4-dinitro-phenylglycine was as described by Neuberger, Perrone, and Slack (1951). Radioactivities were determined on solid samples of “negligible thickness” according to the method of Henriques, Neuberger (1955), or on solid samples of “infinite thickness” when quantities in excess of 25 mg. crystalline DNP-glycine were available.

Sulphate.—This was separated as the 4-chloro-4'-amino diphenyl derivative and radioactivities were determined as previously described (Slack, 1958).

Results

2 weeks after neurectomy total wasting in the atrophying left limb amounted to 23 per cent. in terms of the weight of the normal right hind limb. The greater part of this wasting occurred in the soft tissues (Table I, opposite); 16 weeks after operation loss of tissue in the atrophying limb amounted to 44 per cent., but at 19 weeks it had increased only to 47 per cent. These figures are similar to the rates of total tissue wasting found in the rat (Slack, 1954).

The amounts of collagen fractions obtained are shown in Table II (opposite). As expected, the greater part of the collagen was in the insoluble phase. Neutral salt-soluble collagen formed only a very small part of the total collagen. The amounts obtained after final purification were so small as to present analytical difficulties. Quantitative radioactivity measurements on the neutral salt-soluble fraction of collagen are probably therefore subject to greater experimental variation than either of the two remaining collagen fractions. Consistently smaller amounts of neutral salt-soluble collagen were obtained from the atrophying limb.
TABLE I
WET WEIGHT OF WHOLE TISSUES IN NORMAL AND ATROPHYING RABBIT LIMBS
14 DAYS AFTER NEURECTOMY AND 16 TO 19 WEEKS AFTER
(Percentage wasting expressed as percentage difference from the opposite normal limb)

<table>
<thead>
<tr>
<th>Time Interval after Neurectomy (wks)</th>
<th>Serial No. of Rabbit</th>
<th>Normal Right Hind Limb</th>
<th>Atrophying Left Hind Limb</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>&quot;Tissue Compartment&quot;</td>
<td>Total</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Skeletal</td>
<td>Soft</td>
</tr>
<tr>
<td>2</td>
<td>1001</td>
<td>41</td>
<td>124</td>
</tr>
<tr>
<td></td>
<td>1002</td>
<td>32</td>
<td>108</td>
</tr>
<tr>
<td></td>
<td>1003</td>
<td>38</td>
<td>147</td>
</tr>
<tr>
<td></td>
<td>1004</td>
<td>30</td>
<td>120</td>
</tr>
<tr>
<td></td>
<td>1005</td>
<td>39</td>
<td>135</td>
</tr>
<tr>
<td></td>
<td>1007</td>
<td>48</td>
<td>132</td>
</tr>
<tr>
<td>Average</td>
<td></td>
<td>38</td>
<td>128</td>
</tr>
<tr>
<td>Percentage Wasting</td>
<td></td>
<td>...</td>
<td>...</td>
</tr>
<tr>
<td>16</td>
<td>974</td>
<td>47</td>
<td>149</td>
</tr>
<tr>
<td></td>
<td>975</td>
<td>38</td>
<td>121</td>
</tr>
<tr>
<td>Percentage Wasting</td>
<td></td>
<td>...</td>
<td>...</td>
</tr>
<tr>
<td>19</td>
<td>986</td>
<td>30</td>
<td>90</td>
</tr>
<tr>
<td></td>
<td>987</td>
<td>32</td>
<td>108</td>
</tr>
<tr>
<td>Percentage Wasting</td>
<td></td>
<td>...</td>
<td>...</td>
</tr>
</tbody>
</table>

TABLE II
AMOUNTS OF COLLAGEN FRACTIONS, AND COMPARISON OF TOTAL SEPARATED FRACTIONS
WITH TOTAL COLLAGEN IN ORIGINAL MATERIAL
(Collagen contents determined by the method of Miyada and Tappel (1956))

<table>
<thead>
<tr>
<th>Serial No. of Rabbit</th>
<th>Material</th>
<th>Separated Collagens (g./100 g. fresh tissue)</th>
<th>Total Collagen in Original Material (B)</th>
<th>Difference between A and B as percentage of B</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Neutal Salt-soluble</td>
<td>Acidic Citrate-soluble</td>
<td>Insoluble</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Skeletal</td>
<td>Soft</td>
<td>Skeletal</td>
</tr>
<tr>
<td>1001</td>
<td>Normal</td>
<td>0.0029</td>
<td>0.0021</td>
<td>1.35</td>
</tr>
<tr>
<td></td>
<td>Atrophying</td>
<td>0.0020</td>
<td>0.0014</td>
<td>1.53</td>
</tr>
<tr>
<td>1002</td>
<td>Normal</td>
<td>0.0032</td>
<td>0.0024</td>
<td>1.05</td>
</tr>
<tr>
<td></td>
<td>Atrophying</td>
<td>0.0021</td>
<td>0.0012</td>
<td>1.23</td>
</tr>
<tr>
<td>1003 and 1004</td>
<td>Normal</td>
<td>0.0021</td>
<td>0.0022</td>
<td>1.14</td>
</tr>
<tr>
<td></td>
<td>Atrophying</td>
<td>0.0018</td>
<td>0.0010</td>
<td>1.35</td>
</tr>
<tr>
<td>1005 and 1007</td>
<td>Normal</td>
<td>0.0017</td>
<td>0.0020</td>
<td>1.22</td>
</tr>
<tr>
<td></td>
<td>Atrophying</td>
<td>0.0017</td>
<td>0.0013</td>
<td>1.68</td>
</tr>
</tbody>
</table>

By contrast, larger amounts of acid citrate-soluble collagen were obtained from both skeletal and soft tissue compartments of the atrophying limbs. The insoluble collagen requires special attention.

Insoluble collagen, separated from fresh tissues as described, was found to be smaller in amount per unit weight atrophying limb as compared with the opposite normal limb. On the other hand, total collagen estimated by hydroxyproline was higher per unit weight of tissue in the atrophying limb (Table III, overleaf). The hydroxyproline figures agree with previous experience (Harkness, 1957; Slack, 1954), and it is therefore probable that the extraction procedures used have resulted in a greater loss of purified collagen in the case of atrophying tissue. Reference to the final column of Table III shows that some 15 per cent. of the initial total collagen was lost by the extraction procedure on normal tissues. In atrophying tissue at least 25 per cent. was lost. In general, the atrophying soft tissue showed a greater loss than the skeletal tissue. Some of the difference between estimated total collagen and collagen fractions separated and isolated in the atrophying limbs can be accounted for by increased acid citrate-soluble fraction. But the greater part of the collagen lost...
in the extraction procedure seems to result from altered stability of insoluble collagen at the stage of 0.1 N NaOH extraction.

The amounts of sulphated polysaccharides recovered from the tissues are shown in Table III. In most cases there was a decrease in polysaccharide in the atrophying tissues, both as recovered fractions of polysaccharide and as measured by total hexosamine. There was, however, considerable variation in the actual amounts of polysaccharide recovered per unit weight fresh tissue in normal and atrophying limbs. The variations were such as to permit only a general statement that atrophying tissue, both skeletal and soft, shows some reduction of total polysaccharide per unit weight of fresh tissue.

The data obtained on the metabolism of collagen are given in Table IV. These are admittedly scanty. From the available figures it would appear that the greater reduction of synthesis of collagen occurred in the neutral salt-soluble fraction. The synthesis of this precursor fraction was reduced in both skeletal and soft tissue. At 14 days after neuroectomy the greatest reduction in new synthesis of precursor collagen was evidenced in the soft tissues (Fig. 1, opposite). But, despite this reduction in synthesis of the collagen, all the neutral salt fractions of collagen from the atrophying tissues showed some evidence of continuing synthesis of collagen.

From Table IV it is also evident that some newly synthesized collagen becomes incorporated in both the insoluble collagen fraction and the acid citrate-soluble fractions. The figures do not permit further evaluation beyond the fact that some incorporation of new glycine is being achieved in these collagen fractions, even during active atrophy of the limb tissues.

### Table III

**AMOUNTS OF SULPHATED POLYSACCHARIDES RECOVERED FROM SKELETAL AND SOFT TISSUES OF NORMAL AND ATROPHYING HIND LIMBS OF RABBITS 14 DAYS AFTER NEURECTOMY**

(Amounts determined gravimetrically on acetone-dried specimens)

<table>
<thead>
<tr>
<th>Serial No. of Rabbit</th>
<th>Material</th>
<th>Sulphated Polysaccharides* (mg./100 g. fresh tissue)</th>
<th>Total Separated Polysaccharide</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Neutral Salt Extract</td>
<td>Acidic Citrate Extract</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Skeletal Soft</td>
<td>Skeletal Soft</td>
</tr>
<tr>
<td>1001 Normal</td>
<td></td>
<td>19 12</td>
<td>6 4</td>
</tr>
<tr>
<td>1001 Atrophying</td>
<td></td>
<td>14 19</td>
<td>4 9</td>
</tr>
<tr>
<td>1002 Normal</td>
<td></td>
<td>24 19</td>
<td>14 10</td>
</tr>
<tr>
<td>1002 Atrophying</td>
<td></td>
<td>16 18</td>
<td>10 11</td>
</tr>
<tr>
<td>1003 and 1004 Normal</td>
<td></td>
<td>22 11</td>
<td>12 7</td>
</tr>
<tr>
<td>1003 and 1004 Atrophying</td>
<td></td>
<td>19 16</td>
<td>18 3</td>
</tr>
<tr>
<td>1005 and 1007 Normal</td>
<td></td>
<td>16 7</td>
<td>13 5</td>
</tr>
<tr>
<td>1005 and 1007 Atrophying</td>
<td></td>
<td>7 15</td>
<td>8 7</td>
</tr>
</tbody>
</table>

* Analyses of separated polysaccharides gave figures for sulphate content varying from 10.2 to 12.9 g./100 g. polysaccharide, and of hexosamine 24.2 to 33.6 g./100 g. polysaccharide.

### Table IV

**SPECIFIC RADIOACTIVITIES OF COLLAGEN FRACTIONS FROM SKELETAL AND SOFT TISSUES OF NORMAL AND ATROPHYING HIND LIMBS OF RABBITS 14 DAYS AFTER NEURECTOMY**

<table>
<thead>
<tr>
<th>Serial No. of Rabbit</th>
<th>Time after Isotope</th>
<th>Material</th>
<th>Collagen Radioactivities*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Neutral Salt-soluble</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Skeletal</td>
</tr>
<tr>
<td>1003 and 1004 10 hrs</td>
<td>Normal Atrophying</td>
<td>581 320</td>
<td>1,324 344</td>
</tr>
<tr>
<td>1005 and 1007 24 hrs</td>
<td>Normal Atrophying</td>
<td>174 102</td>
<td>374 120</td>
</tr>
<tr>
<td>1001 2 days</td>
<td>Normal Atrophying</td>
<td>68 54</td>
<td>150 56</td>
</tr>
<tr>
<td>1002 4 days</td>
<td>Normal Atrophying</td>
<td>14 38</td>
<td>20 26</td>
</tr>
</tbody>
</table>

* Counts derived from 14C labelled glycine and radioactivity expressed as count/min./mg. collagen.
METABOLISM OF CONNECTIVE TISSUE IN LIMB ATROPHY IN THE RABBIT

The amounts of sulphated polysaccharide recovered from the tissues are shown in Table III and their specific activities in Table V. The quantities shown in Table III are acetone dried weights of separated polysaccharides. A series of total hexosamine estimations were done on aliquots of original material. These indicated that the amounts of polysaccharide actually obtained in a relatively pure state were of the order of only one-tenth of that presumably present on the basis of total hexosamine estimation. By no means all the hexosamine in total tissue samples derives from mucopolysaccharide, but the polysaccharide actually recovered from the tissues may well be only a portion of that bound to connective tissue proteins in the living tissue.

The polysaccharide extracted from the tissues by neutral salt was small in relation to the total extracted polysaccharide, but showed relatively high specific activity in both normal and atrophying

**Table V**

SPECIFIC RADIOACTIVITIES OF POLYSACCHARIDE SULPHATE IN THREE EXTRACTS OF SKELETAL AND SOFT TISSUES OF NORMAL AND ATROPHYING HIND LIMBS OF RABBITS 14 DAYS AFTER NEURECTOMY

<table>
<thead>
<tr>
<th>Serial No. of Rabbit</th>
<th>Time after Isotope</th>
<th>Material</th>
<th>Radioactivities*</th>
<th>0.1N NaOH Extract</th>
<th>0.1N NaOH Extract</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Neutral Salt Extract</td>
<td>Acidic Citrate Extract</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Skeletal</td>
<td>Soft</td>
<td>Skeletal</td>
</tr>
<tr>
<td>1003 and 1004</td>
<td>10 hrs</td>
<td>Normal Atrophying</td>
<td>686</td>
<td>774</td>
<td>9</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Normal Atrophying</td>
<td>525</td>
<td>470</td>
<td>12</td>
</tr>
<tr>
<td>1005 and 1007</td>
<td>24 hrs</td>
<td>Normal Atrophying</td>
<td>620</td>
<td>542</td>
<td>22</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Normal Atrophying</td>
<td>585</td>
<td>481</td>
<td>40</td>
</tr>
<tr>
<td>1001</td>
<td>2 days</td>
<td>Normal Atrophying</td>
<td>505</td>
<td>409</td>
<td>37</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Normal Atrophying</td>
<td>551</td>
<td>462</td>
<td>28</td>
</tr>
<tr>
<td>1002</td>
<td>4 days</td>
<td>Normal Atrophying</td>
<td>200</td>
<td>237</td>
<td>22</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Normal Atrophying</td>
<td>307</td>
<td>351</td>
<td>36</td>
</tr>
</tbody>
</table>

* Radioactivity expressed as count/min./0.1 mg. sulphate.
limbs. There was no unequivocal evidence of a marked reduction of metabolic activity in this fraction from the atrophying limbs. But how much of this polysaccharide fraction was bound to neutral salt-soluble collagen, and how much to the tissue proteins, is at present uncertain.

The acidic citrate extract of polysaccharide appears to have similar specific activity-time relations to the alkali extract, but the specific activities were much smaller at all intervals of time. By far the greater part of the total polysaccharide was not liberated until the final stage of extraction with 0.1 N alkali. This portion of the polysaccharide was that bound by insoluble connective tissue proteins. The specific activities (of this bound polysaccharide) were higher in the skeletal fractions than in the soft tissue, but in all cases much less than the specific activity found in the neutral salt-extractable polysaccharide. The figures suggest a lower rate of turnover of this polysaccharide fraction from the soft tissues in the atrophying limb, and some reduction in specific activity in the skeletal compartment (Fig. 2).

**Discussion**

From the limited evidence of these experiments and from previous work (Slack, 1954), it seems clear that massive tissue atrophy does not imply a complete cessation of connective tissue synthesis. It might be expected, there does seem to be a reduction of synthesis of both collagen and polysaccharides in both skeletal and soft tissues of the atrophying limb. This reduced synthesis is most evident in the soft tissues at 14 days of atrophy. Since the soft tissues are atrophying at a much greater rate than the skeletal compartment this finding is not surprising. In the case of collagen, the greatest reduction of synthesis occurs in the neutral, salt-soluble, precursor collagen of the atrophying limb. Both the acidic citrate-soluble and insoluble collagens also show some evidence of a reduced rate of metabolic turnover.

It is regrettable that insufficient polysaccharide could be isolated from the precursor collagen fraction to permit adequate analysis. There does appear, however, to be a marked difference in metabolic behaviour between the main polysaccharide (0.1 N alkali) fractions from skeletal and soft tissues. In the skeletal tissue the main bulk of polysaccharide from the atrophying limb shows evidence of continuing synthesis in substantial amounts. On the other hand, that from the soft tissues of the atrophying limb shows definite evidence of a slowing down of synthesis. From these and earlier experiments in the
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(Slack, 1954), and from the work of Gillespie (1954), Geiser and Trueta (1958), and others, it seems clear that massive tissue atrophy is a metabolically active process. It would seem to be an energy-requiring process. Therefore, a continued blood supply, and some continuing synthesis of new connective tissue components would appear, ordinarily, to be an essential requirement.

Summary

(1) Massive tissue atrophy was achieved in the left hind limb of six rabbits by extensive neurctomies and sympathectomy.

(2) 14 days after operation, \( ^{14} \text{C} \) glycine and \( ^{35} \text{S} \) labelled sodium sulphate were injected subcutaneously, and the animals were killed at 10 hrs, 24 hrs, 2 days, and 4 days afterwards.

(3) From the atrophying limb and opposite normal limb in each case, the tissues were separated into a soft tissue compartment (muscle, fascia, and tendon) and a skeletal compartment (bones, joints, and tendon insertions).

(4) Collagen was separated in three fractions: neutral salt-soluble precursor collagen, acidic citrate-soluble collagen, and insoluble collagen. Sulphated polysaccharide was separated in three similar fractions.

(5) In the case of collagen, there is evidence of continued synthesis in both skeletal and soft tissues of the atrophying limbs. The most marked reductions in metabolic activity were found in the soft tissue compartment (which is atrophying most rapidly), and particularly in the neutral salt-soluble collagen.

(6) The main portion of the sulphated polysaccharide, the final alkali-soluble fraction, also shows some reduced metabolic activity, most marked in the soft tissues. The behaviour of polysaccharide associated with the precursor collagen could not be assessed.

(7) The findings provide additional evidence that, although synthesis of collagen and polysaccharide is almost certainly reduced in massive tissue atrophy, there is no complete cessation of these processes.

We wish to thank Prof. J. H. Kellgren for many suggestions and continued encouragement, and Miss K. Broady and Mr. D. Myhill for their technical assistance.

REFERENCES


Métabolisme du tissu conjonctif dans l’extrémité atrophiée du lapin

RÉSUMÉ

(1) On provoqua une atrophie massive des tissus de la patte postérieure gauche de six lapins par des neurctomies et sympathectomies étendues.

(2) Quatorze jours après l’opération, on injecta par voie souscutanée les radio-isotopes \( ^{14} \text{C} \)-glycine et \( ^{35} \text{S} \) sulfate de soude et on sacrifia les animaux 10 heures, 24 heures, 2 jours et 4 jours plus tard.

(3) Dans tous les cas les tissus de la patte atrophiée et de la patte correspondante normale furent divisés en partie molle (muscle, fascia et tendon) et squelettique (os, articulation et insertion tendineuse).

(4) Le collagène fut divisé en trois portions: collagène précurseur, soluble en présence de sel neutre, collagène soluble en présence de citrate acide et collagène insoluble. Le polysaccharide sulfaté fut divisé en trois portions similaires.

(5) En ce qui concerne le collagène, on trouva des indices que sa synthèse continue aussi bien dans le tissu squelettique que le recevant le plus rapidement, et particulièrement dans le collagène soluble en présence de sel neutre.

(6) La portion principale du polysaccharide sulfaté, celle qui se dissout finalement dans l’alcali, accusa aussi une réduction de l’activité métabolique, plus marquée dans les tissus mous. On ne put pas déterminer le comportement du polysaccharide associé au collagène précurseur.

(7) Ces résultats offrent des preuves additionnelles montrant que, bien que la synthèse de collagène et de polysaccharide soit presque certainement réduite dans une atrophie massive des tissus, elle ne cesse pas entièrement.

Metabolismo del tejido conjuntivo en la extremidad atrofiada del conejo

SUMARIO

(1) Se produjo una atrofia masiva de los tejidos de la pata trasera izquierda de seis conejos por neurctomias y sympathectomias extensas.

(2) Catorce dias después de la operación se inyectaron por vía subcutánea los radio-isotopes \( ^{14} \text{C} \)-glicina y \( ^{35} \text{S} \) sulfato de sodio y los animales fueron sacrificados 10 horas, 24 horas, 2 días y 4 días después.

(3) En todos los casos los tejidos de la pata atrofiada y de la pata correspondiente normal fueron divididos en dos partes: una blanda (músculo, fascia y tendon) y la otra esquelética (hueso, articulación e inserción tendineos).


(5) En el caso de colágeno, encontrárnonse indicios
de que su síntesis continua tanto en el tejido esquelético como en el blando de las patas atrofiadas. La mayor reducción de la actividad metabólica fue encontrada en el tejido blando (que se atrofia más rápidamente), particularmente en el colágeno soluble en presencia de sal neutra.

(6) La porción principal del polisacarido sulfatado, aquella que se disuelve finalmente en el álcali, también acusó una reducción de la actividad metabólica, más pronunciada en los tejidos blandos. No se pudo determinar el comportamiento del polisacarido asociado al colágeno precursor.

(7) Estos resultados ofrecen datos adicionales mostrando que, aunque la síntesis del colágeno y del polisacarido se vea casi ciertamente reducida en una atrofia masiva de los tejidos, este proceso no cesa enteramente.