Genetic variation in the hypothalamic–pituitary–adrenal stress axis influences susceptibility to musculoskeletal pain: results from the EPIFUND study

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ABSTRACT

Objectives To determine if genetic variation in genes in the hypothalamic–pituitary–adrenal (HPA) axis, the primary stress response system, influences susceptibility to developing musculoskeletal pain.

Methods Pain and comorbidity data was collected at three time points in a prospective population-based cohort study. Pairwise tagging single nucleotide polymorphisms (SNPs) were selected and genotyped for seven genes. Genetic association analysis was carried out using zero-inflated negative binomial regression to test for association between SNPs and the maximum number of pain sites across the three time points in participants reporting pain, reported as proportional changes with 95% CIs. SNPs were also tested for association with chronic widespread pain (CWP) using logistic regression reporting odds ratios and 95% CI.

Results A total of 75 SNPs were successfully genotyped in 994 participants including 164 cases with persistent CWP and 172 pain-free controls. Multiple SNPs in *SERPINA6* were associated with the maximum number of pain sites; for example, each copy of the T allele of rs941601 was associated with having 16% (proportional change=1.16, 95% CI 1.04 to 1.28, $p=0.006$) more pain sites compared to participants with the CC genotype. *SERPINA6* gene SNPs were also associated with CWP. Significant associations between the maximum number of pain sites and SNPs in the *CRHBP* and *POMC* genes were also observed and a SNP in *MC2R* was also associated with CWP. Associations between SNPs and comorbidity of poor sleep quality and depression explained some of the associations observed.

Conclusions Genetic variation in HPA axis genes was associated with musculoskeletal pain; however, some of the associations were explained by comorbidities. Replication of these findings is required in independent cohorts.

INTRODUCTION

Musculoskeletal pain is common in the general population with up to 33% of people reporting low back pain (LBP) and approximately 11% reporting chronic widespread pain (CWP).¹ Stress has been associated with musculoskeletal pain syndromes^{2,3} and psychological distress has been shown to be a strong predictor for the onset of CWP.⁴

The body's primary stress response system, the hypothalamic–pituitary–adrenal (HPA) axis, has been shown to be hyporesponsive in patients with fibromyalgia and patients with LBP.^{5,6} In a cohort of participants free of CWP those who went onto develop it showed reduced morning and increased evening levels of cortisol and higher levels of

cortisol following a dexamethasone test (HPA axis suppression test). This suggests that individuals developing CWP show a blunting of the cortisol diurnal rhythm and a failure to suppress the HPA axis indicating a hypoactive HPA axis.

It has been hypothesised that the abnormal functioning of the HPA axis in musculoskeletal pain could be due to stressors such as severe adverse events in early life, which have been shown to result in dysfunction of the HPA axis^{7,8} or altered levels of neurotransmitters such as reduced serotonin levels, as observed in fibromyalgia,^{9,10} which stimulates adrenocorticotrophin (ACTH) and corticotrophin releasing hormone (CRH) in response to stress. Genetic variation in genes key to HPA axis function could also play a role in musculoskeletal pain susceptibility. Individuals may respond inefficiently to stressors due to their genetic makeup, which may in turn render them more susceptible to developing musculoskeletal pain.

Twin studies have estimated that genetics explains 50% of the variance in pain syndromes.^{11,12} Pain thresholds, which are reduced in patients with CWP,¹³ have also been shown to have a genetic component,^{14,15} however, one study did not find evidence of this.¹⁶ Attempts to identify the genes involved in CWP susceptibility and experimental pain sensitivity have been limited to date and problematic in their study design. The most prevalent problem is a lack of sufficient sample size to detect the likely modest effects of individual genes. Insufficient representation of the genetic variation within candidate genes and not considering the role of comorbidities are also common problems.¹⁷

In this study we aimed to determine if genetic variation within the HPA axis pathway genes *CRH*, *CRH receptor 1* (*CRHR1*), *CRH binding protein* (*CRHBP*), the ACTH precursor *pro-opiomelanocortin* (*POMC*) and its receptor (*MC2R*), the *glucocorticoid receptor* (*NR3C1*) and *corticosteroid binding globulin* (*SERPINA6*), is associated with susceptibility to musculoskeletal pain while accounting for the effects of comorbidities.

METHODS

Participants

Pain and psychological data was collected at three time points over a 4-year period via a postal survey as part of a prospective population-based cohort study, EPIFUND (for 'Epidemiology of Functional Disorders'). Participants aged 25–65 years were recruited from three primary care registers in the northwest of England.

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Pain phenotype

Participants were asked to indicate the site of any pain experienced for 1 day or longer in the past month on blank body manikins (front, back and sides) and to indicate if their pain had lasted for more than 3 months. Participants were classified two ways. (1) A 'total pain' score was constructed which ranged from 0 (no pain) to 29 (pain in all sites of the body) using the Manchester coding system^{18 19} (figure 1). For each participant the maximum number of pain sites across the three time points was determined. (2) CWP was classified using American College of Rheumatology criteria. Cases were defined as participants with CWP ≥ 2 time points and controls were pain free at all three time points.

Ascertainment of comorbidities

Anxiety, depression, psychological distress and sleep quality were assessed at baseline. Anxiety and depression were measured using the Hospital Anxiety and Depression (HAD) scale.²⁰ The HAD questionnaire contains seven items on anxiety and seven items on depression in the last week. Each question is answered on a 4-point Likert scale (0–3) with total scores ranging from 0 to 21. Higher scores are indicative of a higher probability of having an anxiety or depressive disorder. The General Health Questionnaire²¹ (12-item version) was used to assess levels of psychological distress. Scores range from 0 to 12 with higher scores indicating higher levels of psychological distress. Sleep quality was assessed using the Estimation of Sleep Problems Scale.²² A total of 4 items on recent sleep problems (sleep onset, sleep maintenance, early wakening and non-restorative sleep) were assessed on a 5-point scale (0–5) resulting in a total score of between 0 and 20, with higher scores indicating poorer sleep quality.

Genetic analysis

Participants were asked to provide a buccal swab sample for genetic analysis with informed consent. The sample collection and DNA extraction methods were adapted from Freeman *et al.*²³

Pairwise tagging single nucleotide polymorphisms (SNPs, $r^2 \geq 0.8$) with a minor allele frequency $\geq 5\%$ were selected for *CRH*, *CRHBP*, *CRHR1*, *POMC*, *MC2R*, *NR3C1* and *SERPINA6* and their 10 kb flanking regions using Tagger.²⁴ SNPs were genotyped using Sequenom MassARRAY technology following the manufacturer's instructions (<http://www.sequenom.com>).

Figure 1 Pain sites used to calculate total pain score (Manchester coding).

Sample and assay quality control thresholds were set to 90%. Allele frequencies were checked for consistency with HapMap data and tested for deviation from Hardy–Weinberg Equilibrium (HWE) and excluded from the analysis if $p \leq 0.01$ in the total population. Linkage disequilibrium (LD) between SNPs was examined using Haploview version 3.32.²⁵

Statistical analysis

Analysis was conducted using STATA V.9.2 (Stata, College Station, Texas, USA). p Values < 0.05 were considered statistically significant.

Total number of pain sites

The distribution of the maximum number of pain sites was positively skewed with 17% of participants reporting no pain. To account for this and overdispersion of the data a zero-inflated negative binomial regression (ZINB) model was used. ZINB tests whether a SNP is associated with the odds of reporting no pain (logit) as well as testing for association between a SNP and the maximum number of pain sites in participants who have reported pain (negative binomial). Participants reporting pain can range from having acute pain at a single site to chronic pain at multiple sites, therefore, treating them as a referent group to determine if a SNP is associated with the odds of reporting no pain is a crude analysis and so the results of the logit portion of the model are not reported. Initially a test for trend was conducted. Where there was no evidence of a trend association, recessive (AA and Aa vs aa) and dominant (AA vs Aa and aa) models of association were tested. The results of the negative binomial portion of the model are reported as the proportional change in the number of pain sites with 95% CIs.

The effects of four variables; depression, anxiety, psychological distress and sleep quality on the maximum number of pain sites was assessed using ZINB and only depression and sleep were independently associated (data not shown). Therefore, SNPs significantly associated with the maximum number of pain sites were tested for association with HAD depression score (using negative binomial regression) and sleep quality score (using ZINB). Where there was evidence of an association between a SNP and depression or sleep quality the model was adjusted for the associated comorbidity (depression and/or sleep) to determine if the SNP associations with the maximum number of pain sites were occurring independently of these comorbid factors. Interactions between SNPs (associated with the maximum number of pain sites) and depression and sleep were also tested to determine if they were acting as effect modifiers on the associations observed with pain.

CWP

SNPs were tested for association with CWP using the Cochran–Armitage trend test. Where there was no evidence of a trend association, recessive and dominant models of association were tested using a χ^2 test. Logistic regression was used to estimate odds ratios (OR) and 95% CI.

Haplotype structure was determined using the confidence bounds method in Haploview.²⁶ The expectation–maximisation algorithm was used to estimate haplotype frequencies. The overall distribution of haplotypes was compared between patients with CWP and pain-free controls using a χ^2 test. The frequency of each individual haplotype was also compared to the combined frequency of the other haplotypes between cases and controls using the χ^2 test. Haplotype inference and association testing was conducted in PLINK.²⁷

Depression and sleep were independently associated with CWP (data not shown). Significant associations with CWP were adjusted for depression and/or sleep if there was evidence of association between the SNP and depression and/or sleep, to determine if the associations were occurring independently of these comorbid factors. Interactions between SNPs (associated with CWP) and depression and sleep were also tested to determine if they were moderating the relationship between the SNP and CWP.

RESULTS

Participants

DNA samples were obtained from 1189 participants with pain data at all time points. Samples from 195 (16%) participants were not successfully genotyped for 90% of the SNPs and were excluded from the analysis. The characteristics of the resulting

994 participants are given in table 1. The nested case-control study of CWP consisted of 164 cases and 172 controls. HAD depression and sleep quality scores in the cases were significantly (Mann–Whitney U test, $p < 0.001$) higher than in the controls (table 1).

Genotyping

Of the 88 SNPs selected, 75 were successfully genotyped and in HWE ($p > 0.01$). Coverage of HapMap SNPs ($r^2 < 0.8$) was 100% for *POMC*, 92% for *CRHR1* and *MC2R*, 87% for *SERPINA6*, 84% for *NR3C1*, 67% for *CRH* and 60% for *CRHBP*.

Total number of pain sites

Four SNPs spanning 5' to intron 4 of *SERPINA6* showed association with the maximum number of pain sites in participants reporting pain. rs941601 and rs746530 both showed a significant trend for an increase in the maximum number of pain sites, in participants reporting pain, with the number of copies of the minor allele. A significant increase in the maximum number of pain sites was observed in participants with two copies of the minor allele (recessive effect) for rs11627241 and rs1998056 compared to participants with zero copies or one copy (table 2).

In participants reporting pain, having one or two copies of the minor allele of rs3769371 in *POMC* was associated with a reduction in the maximum number of pain sites compared to having zero copies (dominant effect). Conversely, rs1875999 in *CRHBP* was associated with an increased maximum number of pain sites in participants with one or two copies of the minor allele compared to participants with zero copies (table 2) and also showed evidence of association with an increased likelihood of depression (proportional change=1.10, 95% CI 1.00 to 1.20, $p=0.047$ for each copy of the minor allele) and poor sleep quality (proportional change=1.15, 95% CI 1.06 to 1.25, $p=0.001$ for each copy of the minor allele in participants reporting sleep problems (sleep quality score ≥ 1)). Adjusting the association between the maximum number of pain sites and rs1875999 for HAD depression score and sleep quality rendered the association non-significant (proportional change=1.01, 95% CI 0.91 to 1.11, $p=0.860$).

There was no evidence of interaction between the SNPs, associated with pain, and depression or sleep on the maximum number of pain sites. There was also no evidence of association between the maximum number of pain sites in participants reporting pain and SNPs genotyped in *CRH*, *CRHR1*, *MC2R* and *NR3C1*.

CWP

Two SNPs in *SERPINA6* were significantly associated with CWP. Rs941601 and rs8022616 showed a significant trend for increasing and decreasing the odds of having CWP with the number of copies of the minor allele, respectively (table 3). These two SNPs are

Table 1 Participant characteristics

	Total population	CWP analysis	
		Cases	Controls
n	994	164	172
Median age (95% CI)	50.9 (49.8 to 52.0)	52.6 (50.3 to 53.9)	48.5 (46.4 to 51.6)
% Female	58	66	57
Median depression score (95% CI)	3 (2 to 3)	5 (4 to 6)	2 (1 to 2)
Median sleep score (95% CI)	5 (5 to 5)	11 (10 to 13)	3 (2 to 4)

CWP, chronic widespread pain.

Table 2 Significant associations with the maximum number of pain sites in participants reporting pain

Gene	SNP	Genotype	n	Proportional change (95% CI)	p Value
<i>SERPINA6</i>	rs941601	CC	604	Reference	–
		CT	202	1.16 (1.04 to 1.28)	0.006
		TT	16	1.34 (1.09 to 1.64)	0.006
	rs11627241	CC and CT	772	Reference	–
		TT	50	1.27 (1.03 to 1.57)	0.026
	rs1998056	CC and CG	671	Reference	–
		GG	149	1.18 (1.03 to 1.34)	0.017
	rs746530	GG	352	Reference	–
		GA	360	1.10 (1.02 to 1.19)	0.011
		AA	101	1.21 (1.05 to 1.41)	0.011
<i>POMC</i>	rs3769671	AA	740	Reference	–
		AC and CC	65	0.81 (0.67 to 0.99)	0.040
<i>CRHBP</i>	rs1875999	TT	373	Reference	–
		TC and CC	444	1.12 (1.01 to 1.24)	0.036

SNP, single nucleotide polymorphism.

Table 3 Significant associations with CWP

Gene	SNP	Genotype	n (%)		OR (95% CI)	p Value
			Cases	Controls		
<i>SERPINA6</i>	rs941601	CC	117 (71)	138 (81)	Reference	–
		CT	43 (26)	31 (18)	1.61 (1.02 to 2.55)	0.040
		TT	4 (3)	2 (1)	2.59 (1.03 to 6.52)	0.040
	rs8022616	AA	142 (87)	131 (77)	Reference	–
		AG	20 (12)	36 (21)	0.58 (0.34 to 0.97)	0.037
		GG	2 (1)	3 (2)	0.33 (0.12 to 0.93)	0.037
<i>MC2R</i>	rs11661134	GG	132 (84)	158 (92)	Reference	–
		AG and AA	26 (16)	14 (8)	2.22 (1.12 to 4.43)	0.023

CWP, chronic widespread pain; SNP, single nucleotide polymorphism.

A single SNP in *MC2R*, rs11661134, was associated with an increased odds of having CWP in participants with one or two copies of the minor A allele compared to participants with zero copies (OR=2.24, 95% CI 1.14 to 4.39, p=0.02). There was no evidence of interaction between SNPs associated with CWP

One of the strengths of this study is the phenotypes used. Cases had reported CWP at at least two of the three time points,

Figure 2 Linkage disequilibrium and haplotype block structure of *SERPINA6* in the study population. Single nucleotide polymorphisms (SNPs) genotyped and their position in *SERPINA6* are shown with pairwise linkage disequilibrium (LD; colour coded by r^2 (white=0, black=1) and numbered by D' (if no number $D'=1$)) and haplotype block structure as defined by the confidence bounds method.²⁶

Combination of SNPs	Overall distribution p value	Distribution of individual haplotypes			
		Haplotype	Case %	Control %	p Value
rs941601–rs3790036–	0.09	CACCAG	0.21	0.22	0.881
rs2144835–rs11629171–		CATTGA	0.07	0.13	0.019
rs8022616–rs11622665		CGTTAA	0.16	0.17	0.651
		CACCAA	0.14	0.15	0.762
		TATCAA	0.16	0.10	0.041
		CATCCA	0.26	0.23	0.401

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providing a robust phenotype, and were compared to a pain-free control group to avoid erroneous associations due to the presence of non-persistent CWP, WP or regional pain disorders which may also be influenced by genetic variation in the HPA axis. However, this results in a limited sample size. A novel method was used to quantify pain by creating a composite score of the number of body regions, from 0 to 29, making it possible to examine the relationship between genetic variation and musculoskeletal pain in the entire study population thus increasing statistical power.

One limitation of the study is that ethnicity of participants was not determined; however, participants were derived from predominantly white Caucasian geographic areas. Another limitation of our study is that due to limited power we have chosen not to correct for multiple testing. Indeed correcting for the number of effective tests, accounting for LD between SNPs, using the methodology proposed by Li and Ji³⁵ would result in a p value cut-off of 0.00037; none of the associations observed reached this level of significance. However, this method of correction is comparatively stringent and may result in false negatives if applied. To be certain whether these are true pain susceptibility loci or false positives, independent replication of these findings in larger cohorts is essential.

It was anticipated that similar findings would be detected with the two pain outcomes as the patients with CWP also tend to have a high number of pain sites. There is some consistency between the findings of the CWP and maximum number of pain sites analysis but also some discrepancies. This may be explained by the reduced power in the CWP analysis or because the maximum pain score is capturing acute and chronic pain which may not be influenced by the same genetic factors.

In conclusion, we report the first evidence that genetic variation in the primary stress response system influences susceptibility to musculoskeletal pain in a general population sample. However, the associations reported are modest, sometimes explained by psychological comorbidity and require replication in a large independent cohort to determine whether they play a role in the aetiology of musculoskeletal pain.

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Competing interests None.

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REFERENCES

- Croft P, Rigby AS, Boswell R, *et al*. The prevalence of chronic widespread pain in the general population. *J Rheumatol* 1993;**20**:710–13.
- Hoogendoorn WE, Bongers PM, de Vet HC, *et al*. High physical work load and low job satisfaction increase the risk of sickness absence due to low back pain: results of a prospective cohort study. *Occup Environ Med* 2002;**59**:323–8.
- Diepenmaat AC, van der Wal MF, de Vet HC, *et al*. Neck/shoulder, low back, and arm pain in relation to computer use, physical activity, stress, and depression among Dutch adolescents. *Pediatrics* 2006;**117**:412–16.
- McBeth J, Macfarlane GJ, Benjamin S, *et al*. Features of somatisation predict the onset of chronic widespread pain: results of a large population-based study. *Arthritis Rheum* 2001;**44**:940–6.
- Crofford LJ, Pillemer SR, Kalogeras KT, *et al*. Hypothalamic-pituitary-adrenal axis perturbations in patients with fibromyalgia. *Arthritis Rheum* 1994;**37**:1583–92.
- Griep EN, Boersma JW, Lentjes EG, *et al*. Function of the hypothalamic-pituitary-adrenal axis in patients with fibromyalgia and low back pain. *J Rheumatol* 1998;**25**:1374–81.
- Van VE, Scarpa A. The effects of child maltreatment on the hypothalamic-pituitary-adrenal axis. *Trauma Violence Abuse* 2004;**5**:333–52.
- Weissbecker I, Floyd A, Dedert E, *et al*. Childhood trauma and diurnal cortisol disruption in fibromyalgia syndrome. *Psychoneuroendocrinology* 2006;**31**:312–24.
- Wolfe F, Russell IJ, Viapraio G, *et al*. Serotonin levels, pain threshold, and fibromyalgia symptoms in the general population. *J Rheumatol* 1997;**24**:555–9.
- Russell IJ, Viapraio GA, Lopez YG. Serum serotonin in fibromyalgia syndrome, rheumatoid arthritis and healthy normol controls. *Arthritis Rheum* 1993;**36**:S222.
- Kato K, Sullivan PF, Evengård B, *et al*. Importance of genetic influences on chronic widespread pain. *Arthritis Rheum* 2006;**54**:1682–6.
- MacGregor AJ, Andrew T, Sambrook PN, *et al*. Structural, psychological, and genetic influences on low back and neck pain: a study of adult female twins. *Arthritis Rheum* 2004;**51**:160–7.
- Wolfe F, Ross K, Anderson J, *et al*. Aspects of fibromyalgia in the general population: sex, pain threshold, and fibromyalgia symptoms. *J Rheumatol* 1995;**22**:151–6.
- Norbury TA, MacGregor AJ, Urwin J, *et al*. Heritability of responses to painful stimuli in women: a classical twin study. *Brain* 2007;**130**:3041–9.
- Nielsen CS, Stubhaug A, Price DD, *et al*. Individual differences in pain sensitivity: Genetic and environmental contributions. *Pain* 2007;**136**:21–29.
- MacGregor AJ, Griffiths GO, Baker J, *et al*. Determinants of pressure pain threshold in adult twins: evidence that shared environmental influences predominate. *Pain* 1997;**73**:253–7.
- Limer KL, Nicholl BI, Thomson W, *et al*. Exploring the genetic susceptibility of chronic widespread pain: the tender points in genetic association studies. *Rheumatology (Oxford)* 2008;**47**:572–7.
- MacFarlane GJ, Croft PR, Schollum J, *et al*. Widespread pain: is an improved classification possible? *J Rheumatol* 1996;**23**:1628–32.
- Hunt IM, Silman AJ, Benjamin S, *et al*. The prevalence and associated features of chronic widespread pain in the community using the 'Manchester' definition of chronic widespread pain. *Rheumatology (Oxford)* 1999;**38**:275–9.
- Zigmond AS, Snaith RP. The hospital anxiety and depression scale. *Acta Psychiatr Scand* 1983;**67**:361–70.
- Goldberg D. *A User's Guide to the General Health Questionnaire*. Windsor: NFER-Nelson, 1988.
- Jenkins CD, Stanton BA, Niemcryk SJ, *et al*. A scale for the estimation of sleep problems in clinical research. *J Clin Epidemiol* 1988;**41**:313–21.
- Freeman B, Powell J, Ball D, *et al*. DNA by mail: an inexpensive and noninvasive method for collecting DNA samples from widely dispersed populations. *Behav Genet* 1997;**27**:251–7.
- de Bakker PI, Yelensky R, Pe'er I, *et al*. Efficiency and power in genetic association studies. *Nat Genet* 2005;**37**:1217–23.
- Barrett JC, Fry B, Maller J, *et al*. Haploview: analysis and visualisation of LD and haplotype maps. *Bioinformatics* 2005;**21**:263–5.
- Gabriel SB, Schaffner SF, Nguyen H, *et al*. The structure of haplotype blocks in the human genome. *Science* 2002;**296**:2225–9.
- Purcell S, Neale B, Todd-Brown K, *et al*. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* 2007;**81**:559–75.
- Claes S, Villafuerte S, Forsgren T, *et al*. The corticotropin-releasing hormone binding protein is associated with major depression in a population from Northern Sweden. *Biol Psychiatry* 2003;**54**:867–72.
- McBeth J, Silman AJ, Gupta A, *et al*. Moderation of psychosocial risk factors through dysfunction of the hypothalamic-pituitary-adrenal stress axis in the onset of chronic widespread musculoskeletal pain: findings of a population-based prospective cohort study. *Arthritis Rheum* 2007;**56**:360–71.
- Van Den EF, Moorkens G, Van HB, *et al*. Hypothalamic-pituitary-adrenal axis function in chronic fatigue syndrome. *Neuropsychobiology* 2007;**55**:112–20.
- Rajeevan MS, Smith AK, Dimulescu I, *et al*. Glucocorticoid receptor polymorphisms and haplotypes associated with chronic fatigue syndrome. *Genes Brain Behav* 2007;**6**:167–76.
- Smith AK, White PD, Aslaksen E, *et al*. Polymorphisms in genes regulating the HPA axis associated with empirically delineated classes of unexplained chronic fatigue. *Pharmacogenomics* 2006;**7**:387–94.
- Stevens A, Ray DW, Zeggini E, *et al*. Glucocorticoid sensitivity is determined by a specific glucocorticoid receptor haplotype. *J Clin Endocrinol Metab* 2004;**89**:892–7.
- Rautanen A, Eriksson JG, Kere J, *et al*. Associations of body size at birth with late-life cortisol concentrations and glucose tolerance are modified by haplotypes of the glucocorticoid receptor gene. *J Clin Endocrinol Metab* 2006;**91**:4544–51.
- Li J, Ji L. Adjusting multiple testing in multilocus analyses using the eigenvalues of a correlation matrix. *Heredity* 2005;**95**:221–7.