Measurement of cytokine and adhesion molecule expression in synovial tissue by digital image analysis

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Abstract

Objective—Digital image analysis (DIA) offers the opportunity to quantify the stained area and staining intensity when synovial tissue (ST) is investigated by immunohistochemical analysis. This study aimed at determining the sensitivity of DIA compared with semiquantitative analysis (SQA).

Methods—Paired ST samples were obtained from the knee joint of 10 patients with rheumatoid arthritis (RA) with active disease and after follow up when complete clinical remission was achieved. ST samples of 10 subjects with non-inflammatory knee pain served as controls. Immunohistochemistry with antibodies against interleukin 1 β (IL1 β) and vascular cell adhesion molecule 1 (VCAM-1) was applied using two staining protocols with 3-amino-9-ethylcarbazole (AEC) or *p*-diethylaminobenzaldehyde (DAB) as dye. All sections were analysed semiquantitatively (0–4) and DIA of up to a maximum of 60 high power fields (HPF). The average integrated optical density was calculated as the product of the stained area (corrected for total tissue area) and the optical density.

Results—Both SQA and DIA enabled the assessment of differences in IL1 β and VCAM-1 expression between ST from active RA, RA in remission, and controls. SQA and DIA showed excellent correlations (IL1 β $r_s=0.867$; $p<0.0001$; VCAM-1 $r_s=0.828$; $p<0.0001$). A limited analysis of one region with six HPF still allowed adequate discrimination compared with an extended analysis of three regions with a total of 60 HPF. In general, the red dye (AEC) resulted in better discrimination than the brown (DAB) staining.

Conclusion—DIA offers a reliable, reproducible, and sensitive analysis of ST sections stained for cytokines and adhesion molecules.

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analysis (SQA).⁴ A possible limitation of SQA is its sensitivity to change after treatment, as documented in studies on cellular markers.⁵ Digital image analysis (DIA) has proved to be a sensitive and reliable tool in the evaluation of cellular markers of the infiltrate,^{6,7} and it has been applied for the measurement of cytokine and adhesion molecule expression in ST.⁸ In the absence of comparative studies between DIA and SQA, this study aimed at documenting the sensitivity of both techniques in the measurement of expression of cytokines and adhesion molecules in paired ST samples of patients with RA with active disease who achieved clinical remission and controls without joint inflammation.

Methods

We studied 10 patients with RA (1987 American College of Rheumatology (ACR) criteria) for expression of interleukin 1 β (IL1 β) and vascular cell adhesion molecule 1 (VCAM-1) in ST. In all 10 patients an arthroscopy was performed under regional anaesthesia in an actively inflamed knee joint,⁹ and in the same knee joint after the establishment of complete clinical remission (1981 ACR criteria¹⁰). ST samples were taken under direct vision with a 4.0 mm biopsy forceps. Controls were 10 patients who had undergone an arthroscopy for unclassified pain without signs of joint inflammation at arthroscopy and during follow up. All patients gave informed consent and the ethical committee of the Repatriation General Hospital approved the study protocol.

ST samples were either snap frozen in liquid nitrogen or fixed in formalin and subsequently embedded in paraffin. Both frozen and paraffin embedded blocks were cut in 5 μ m sections. Frozen sections were stained with monoclonal antibodies against VCAM-1 (anti-CD106; Ig11B1, Sanbio). Paraffin embedded sections were stained with monoclonal antibodies against IL1 β (LP-712; Genzyme, Cambridge, MA).¹¹ For the staining we applied a secondary and a tertiary antibody together with Fast Red (FRV, red) for IL1 β , or 3-amino-9-ethylcarbazole (AEC, red) for VCAM-1, and Mayer's haemalum counterstaining.¹¹

After staining all sections were analysed by both SQA and DIA. Two independent observers performed SQA on all sections using a five point scale.¹¹ Three acquisition methods were used for DIA: one high power field (HPF, $\times 400$ magnification, surface ± 0.125 mm²) in six representative regions (total six HPF, 0.75 mm²), and 20 consecutive HPF in one representative

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Table 1 Mean score (standard error of the mean) of semiquantitative analysis (SQA) and digital image analysis (DIA) on six high power fields (HPF) and 60 HPF for interleukin(IL)1 β and vascular adhesion molecule (VCAM)-1. Depicted are the paired results for patients with active rheumatoid arthritis and after the achievement of clinical remission and healthy controls

		Active disease (A)	Remission (R)	Controls (C)	A v R (p value)	R v C (p value)
IL1 β	SQA	3.0 (0.3)	1.4 (0.3)	1.0 (0.2)	<0.005	NS
	DIA					
	60 HPF	21 760 (3 733)	10 446 (3 034)	4 475 (659)	0.005	NS
	20 HPF	22 291 (3 289)	10 506 (3 073)	4 438 (713)	0.005	NS
VCAM-1	6 HPF	26 288 (5 292)	12 996 (3 993)	4 504 (741)	<0.01	NS
	SQA	3.1 (0.4)	2.2 (0.4)	1.0 (0.3)	NS	<0.05
	DIA					
	60 HPF	67 009 (8 503)	39 674 (9 337)	11 747 (4 876)	<0.05	0.01
	20 HPF	68 879 (10 458)	40 546 (7 749)	11 966 (4 534)	0.01	0.007
	6 HPF	83 409 (12 918)	45 758 (10 763)	10 789 (4 494)	<0.05	0.005

Figure 1 Plots of (A) semiquantitative analysis with digital image analysis of 20 high power fields (HPF) in one single region; (B) digital image analysis of 20 high power fields (HPF) in three regions with 20 high power fields (HPF) each (total 60 HPF); and (C) the difference between the two digital image analyses with the average between the two techniques as suggested by Bland and Altman.¹² Depicted are the semiquantitative score on a five point scale and the integrated optical density (IOD) for interleukin 1 β staining in synovial tissue of all subjects studied and the Spearman correlation coefficient (r_s).

region (total 20 HPF, 2.5 mm²) or 20 consecutive HPF in three representative regions (total 60 HPF, 7.5 mm²), respectively.^{7,8} For the analysis of all digitised images we used a specialised program written in the programming language QUIPS using the Qwin v2.2 program (Leica, Cambridge, UK) on a personal computer. We calculated the integrated optical density as the product of staining area and intensity⁸ corrected for the area for each section stained.

STATISTICAL ANALYSIS

Wilcoxon signed rank tests were used for the comparison of active disease and remission. Mann-Whitney tests were used to compare remission and controls. Spearman rank tests were used to calculate the correlations between the different acquisition and analysis techniques. The plot of the average and the difference of two digital image analysis procedures and the calculation of the correlation were performed as suggested by Bland and Altman.¹²

Results

PATIENT DEMOGRAPHICS

The mean age of the 10 patients with RA (six male, four female) was 72 years (range 59–77). Five patients had erosive disease and five patients non-erosive disease; average disease duration was 54 months (range 1–216). The mean age of the control subjects (six male, four female) was 39 years (range 28–54). None of the 10 patients with RA had taken disease modifying antirheumatic drugs at the time of the first biopsy procedure.

RESULTS OF SQA AND DIA

As shown in table 1, SQA of the sections stained for IL1 β showed a significant difference between active disease and remission ($p < 0.005$), whereas the scores for IL1 β were similar for patients with RA in remission and controls. DIA showed a significant difference between active disease and remission for 60 HPF ($p = 0.005$), 20 HPF ($p = 0.005$), and six HPF ($p < 0.01$). Comparison of the results of DIA between patients with RA in remission and controls yielded no significant differences. SQA of the sections stained for VCAM-1 showed no significant difference between patients with active disease and patients in remission; in controls the scores were signifi-

cantly lower ($p < 0.05$). DIA showed significant differences between active disease and remission for 60 HPF ($p < 0.05$), 20 HPF ($p = 0.01$), and six HPF ($p < 0.05$). Comparison of the results of DIA between remission and controls also showed significant differences for 60 HPF ($p = 0.01$), 20 HPF ($p = 0.007$), and six HPF ($p = 0.005$).

CORRELATIONS BETWEEN SQA AND DIFFERENT DIA METHODS

Figures 1A and B show that for IL1 β , SQA was highly significantly correlated with DIA of 20 HPF ($r_s = 0.942$, $p < 0.0001$), and 20 HPF with 60 HPF ($r_s = 0.967$, $p < 0.0001$), respectively. A strong correlation was found between 60 HPF and 6 HPF ($r_s = 0.844$, $p < 0.0001$) as well. To investigate the effect of the limitation of the number of HPF in DIA we calculated the correlation between methods of measurement.¹² The calculated correlation between 60 HPF and 20 HPF was -0.08 , suggesting equal variance. Correlation between 60 HPF and 6 HPF was -0.834 , suggesting a marked difference in the variability between subjects for the two methods despite the strong correlation. Figure 1C shows the plot of the difference against the average of both 60 HPF and 20 HPF. Similar results were found for VCAM-1.

Discussion

In this study we show that DIA and SQA give similar results for the analysis of both IL1 β and VCAM expression in ST sections. Overall, DIA proved to be slightly more sensitive, especially in comparing remission with controls. Moreover, even a limited analysis of 20 HPF (or 2.5 mm² of ST) produced representative results.

To our knowledge this is the first paper comparing SQA with DIA for the analysis of cytokine and adhesion molecule expression in the synovial tissue of patients with RA. We chose to investigate the expression of IL1 β in synovial tissue because this cytokine plays a part in both inflammation and destruction. Furthermore, it has been documented that joint destruction associated with RA can be found in patients in clinical remission and we expected this cytokine to be of special interest in our study group.¹³ DIA identified differences between controls and patients with RA in remission, but these changes were not statistically significant.

We investigated VCAM-1 based upon the role of the VCAM-1/VLA-4 ligand pair in the preferential recruitment of activated lymphocytes and monocytes from the circulation into the RA ST.¹⁴ Of interest, we noted an

increased expression of VCAM-1 in patients with RA in remission. This observation is consistent with previous observations showing signs of activation in clinically unaffected joints.¹⁵

We conclude that the ability of DIA to quantify the expression of a marker offers a sensitive tool to determine protein expression in synovial tissue. Because it can be expected that DIA will be increasingly used for the evaluation of synovial tissue, further standardisation of the methodology is mandatory.

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