C-reactive protein and serological indices of disease activity in systemic lupus erythematosus

J. V. BERTOUCHE,¹ P. J. ROBERTS-THOMPSON,¹ P. H. FENG,²
AND J. BRADLEY¹

From the ¹Department of Clinical Immunology, Flinders Medical Centre, South Australia, and ²Singapore General Hospital, Singapore

SUMMARY The concentration of C-reactive protein (CRP) in sera from 70 patients with systemic lupus erythematosus (SLE) showed no correlation with commonly accepted laboratory indices of disease activity. Most patients had detectable serum CRP, but in some patients CRP was not found despite repeated testing. This absence of a CRP response did not appear to be related to medication. In some patients high levels of CRP were seen in the absence of infection. Measurement of serum CRP in SLE is unlikely to be useful in the laboratory diagnosis of disease activity.

C-reactive protein (CRP) is a major acute-phase reactant whose concentration may rise with tissue inflammation or necrosis, infectious diseases, and malignancy. CRP levels have been used to monitor disease activity and response to therapy in a number of rheumatic diseases, including rheumatoid arthritis,¹ vasculitic syndromes,² and ankylosing spondylitis.³ In systemic lupus erythematosus (SLE) the role of CRP is less clear, and there have been a number of conflicting reports as to the incidence and significance of elevated CRP levels. Early studies⁴⁻⁶ indicated that patients with SLE rarely have positive CRP sera but that a positive result suggests superimposed infection. These studies used a semiquantitative capillary precipitation technique to detect CRP, and false positive results can occur in the presence of IgM antibodies such as rheumatoid factor.⁷

Subsequent studies⁸⁻⁹ using radial immunodiffusion¹⁰ suggested that CRP elevation in the course of SLE is most frequently associated with exacerbations of clinical disease activity and does not necessarily indicate the presence of infection. Becker *et al.*,¹¹ using an electroimmunoassay technique to measure CRP, found that higher concentrations were seen in clinically active disease but that levels greater than 60 mg/l suggested intercurrent infection. All of these papers have correlated CRP levels with clinical manifestation of disease activity. A number of immunological investigations including double-standard DNA antibodies (DNA Abs), circulating

immune complexes (CICs), and C3 and C4 are considered to be useful in laboratory monitoring of disease activity in SLE.

The purpose of this study was to determine whether CRP levels could be correlated with any of these serological indices.

Patients and methods

Seventy patients fulfilling the American Rheumatology Association criteria for the classification of SLE were included in the study. The group consisted of subjects of Chinese and Indian descent resident in Singapore who attended the LE Clinic at Singapore General Hospital. None of the patients had clinical evidence of any infective process at the time of sampling. All patients were seen by one of us (J.B.), and clinical details and serum samples were collected and transported at -20°C to Adelaide, where they were stored at this temperature until required.

CRP was measured by means of radial immunodiffusion.¹⁰ The diameters of precipitin rings were measured with a magnifying (×7) calibrated eyepiece, and the smallest diameter ring that could be measured confidently corresponded to a concentration of 6 µg/ml (6 mg/l). The CRP standard was obtained from Behringwerke AG, West Germany, and the anti-CRP antiserum was supplied by Dako Immunoglobulins, Denmark. DNA Abs were measured by the Pincus modification of the Farr technique.¹² C3, C4 and alpha-1-antitrypsin (α1 AT) were measured by a nephelometric technique with a Technicon Autoanalyser. CICs were measured by the fluid phase Clq binding assay.¹³

Accepted for publication 10 January 1983.

Correspondence to Professor J. Bradley, Department of Clinical Immunology, Flinders Medical Centre, Bedford Park, South Australia 5042.

Laboratory results regarded as abnormal were: CRP $>6 \mu\text{g/ml}$ (6 mg/l) (normal <6 as obtained from 100 blood donors), DNA Abs $>30\%$, C3 $<55 \text{ mg/100 ml}$ (0.55 g/l) (normal range $55\text{--}120 \text{ mg/100 ml}$) ($0.55\text{--}1.20 \text{ g/l}$), C4 $<20 \text{ mg/100 ml}$ (0.2 g/l) (normal range $20\text{--}50 \text{ mg/100 ml}$) ($0.2\text{--}0.5 \text{ g/l}$), α_1 AT $<0.85 \text{ mg/ml}$ (0.85 g/l) (normal range $0.85\text{--}2.13 \text{ mg/ml}$ (or g/l)), and CICs >3 units.

The results were set out in 2×2 contingency tables and analysed by the chi-squared test. Analysis of DNA Abs, C3, and C4 results was performed by the Kendall coefficient of concordance.¹⁴

Results

CRP was detectable in 55 patients. Thirteen of them had levels greater than $6 \mu\text{g/ml}$ with a range from $6.8 \mu\text{g/ml}$ to 99.5 g/ml . The remaining 42 patients in

this group had concentrations less than $6 \mu\text{g/ml}$ but a visible precipitation ring. (SI conversion = $\mu\text{g/ml} = \text{mg/l}$). DNA Abs were elevated in 42 patients. C3 was reduced in 39 patients and C4 reduced in 27 patients. α_1 AT was in the normal range in 50 patients and elevated in 20 patients. CICs were elevated in 20 patients. The complete results for patients with an elevated CRP are shown in Table 1. Results for those patients with no detectable CRP are shown in Table 2. In the overall group of 70 patients there was no significant correlation between CRP levels and DNA Abs, CICs, C3, C4, or α_1 AT ($p > 0.05$, chi-squared test). By contrast there was a highly significant correlation between elevated DNA Abs and reduced levels of C3 and C4 ($W = 0.7$, Kendall coefficient of concordance). The correlation between elevated CICs, elevated DNA Abs, reduced C3, and reduced C4 was not significant.

Table 1 Summary of serological indices in those patients with an elevated CRP

Patient	CRP ($\mu\text{g/ml}$)	DNA Abs (%)	C3 (g/l)	C4 (g/l)	CICs (units)	α_1 AT (g/l)
1	25.8	41	0.63	0.37	<3	1.74
2	7.5	36	0.40	0.25	<3	2.72
3	20	15	0.46	0.22	14	1.2
4	61.8	32	0.91	0.46	<3	3.42
5	19.9	97	0.20	0.17	<3	2.49
6	99.9	28	0.40	0.18	<3	3.93
7	12.5	17	0.20	0.20	<3	1.86
8	48.5	25	0.82	0.45	<3	2.13
9	14	34	0.74	0.37	5.3	1.52
10	18.5	15	0.76	0.39	48	3.75
11	60	85	0.32	0.24	5.3	2.62
12	6.8	24	0.60	0.31	<3	1.83
13	50.6	16	0.44	0.22	12	2.71

SI conversion: CRP $\mu\text{g/ml} \times 1 = \text{mg/l}$.

Table 2 Summary of serological indices in those patients with no detectable CRP

Patient	CRP ($\mu\text{g/ml}$)	DNA Abs (%)	C3 (g/l)	C4 (g/l)	CICs (units)	α_1 AT (mg/ml)
14	—	94	0.11	0.5	<3	1.44
15	—	12	0.37	0.21	<3	1.38
16	—	48	0.89	0.33	<3	1.71
17	—	31	0.63	0.18	<3	2.3
18	—	14	0.53	0.29	<3	1.68
19	—	84	0.22	0.14	<3	1.44
20	—	47	0.47	0.15	<3	1.92
21	—	16	0.79	0.22	<3	1.92
22	—	44	0.23	0.09	9.4	1.74
23	—	47	0.48	0.17	<3	1.86
24	—	96	0.29	0.17	23	2.1
25	—	12	0.55	0.28	<3	1.08
26	—	78	0.26	0.21	3.5	2.04
27	—	66	0.34	0.9	<3	1.68
82	—	48	0.34	0.2	63	1.8

SI conversion: CRP $\mu\text{g/ml} \times 1 = \text{mg/l}$.

Discussion

We have shown that CRP levels do not correlate with commonly accepted laboratory indices of disease activity in SLE. This is in contrast with rheumatoid arthritis, where CRP levels correlate significantly with clinical¹⁵ radiological¹⁶ and serological¹⁷ indices of disease activity. The present study used the Mancini radial immunodiffusion technique for CRP measurement. The results show that 42 patients had visible precipitation rings which could not be measured accurately because of their small diameter. More sensitive measurement techniques not available to us such as radioimmunoassay would have allowed quantitation of these results. Nevertheless CRP was elevated in only 13 patients, and none had clinical evidence of infection. Of these, 6 patients were studied on presentation and in the absence of any treatment and 7 were receiving corticosteroids. Fifteen patients had no visible precipitation ring. Further serum samples from 5 of these patients were measured subsequently, and still no precipitation ring was visible. Four of these patients were sampled on presentation and without treatment, and the remaining 11 were receiving corticosteroids. Seven of these 11 patients had active disease both clinically and serologically. It is known that corticosteroids do not directly influence the CRP response unless underlying disease activity is modified,¹⁸ and it has been previously shown in SLE that there is no correlation between CRP levels and prednisolone dose rate.¹⁹ Our findings are in accord with these reports, as those patients with absent CRP had both active disease and were receiving corticosteroids at the time of serum collection.

The absence of CRP in some patients has been previously noted by Morrow *et al.*,⁹ and it has previously been suggested by Becker *et al.*¹¹ that hereditary effects on production of CRP in response to certain stimuli may account for this unresponsiveness. It was further suggested by Becker *et al.*¹¹ that, as CRP interacts with complement, then impairment of the acute-phase response on a genetic basis may be important in the pathogenesis of inflammatory disease. It should be emphasised that single serum samples only were obtained from each patient, and previous reports^{5,9} have shown that serial CRP levels from the same patient may show quite marked fluctuations.

Although we did not attempt to examine fully the acute-phase proteins in this study, it is of interest that all patients had either normal or elevated α_1 AT levels. A report on serum amyloid-A protein (SAA), another acute-phase protein, in a number of inflammatory diseases, including SLE, has recently been published.²⁰ SAA levels were found to be lower in

SLE than rheumatoid arthritis and juvenile chronic arthritis and tended to parallel CRP concentrations in these diseases. There are many other acute-phase proteins, and a more comprehensive study of their behaviour in SLE would be particularly interesting.

There was a significant correlation between elevated DNA Abs, low C3, and low C4, confirming the impression that these indices tend to parallel each other. The lack of correlation with CICs is undoubtedly due to the fluid phase assay used. It is well recognised that the solid phase Clq binding assay is more specific for the CICs found in SLE.²¹

In conclusion, CRP levels do not correlate with some of the commonly accepted indices of disease activity in SLE and are not likely to be helpful in the laboratory assessment of disease activity.

References

- Hedlund P. Clinical experimental studies on C-reactive protein (acute phase protein). *Acta Med Scand* 1961; **361** (suppl): 35-71.
- Parish W E. Studies on vasculitis. VII. C-reactive proteins as a substance perpetuating chronic vasculitis. Occurrence in lesions and concentrations in sera. *Clin Allergy* 1976; **6**: 543-50.
- Cowling P, Ebringer R, Cawdell D, Ishii M, Ebringer A. C-reactive protein, ESR, and klebsiella in ankylosing spondylitis. *Ann Rheum Dis* 1980; **39**: 45-9.
- Hill A G S. C-reactive protein in the chronic rheumatic diseases. *Lancet* 1951; **ii**: 807-12.
- Honig S, Gorevic P, Weissman G. C-reactive protein in systemic lupus erythematosus. *Arthritis Rheum* 1977; **20**: 1065-9.
- Bravo M G, Alarcon-Segovia D. C-reactive protein in the differential diagnosis between infection and disease reactivation in SLE. *J Rheumatol* 1981; **8**: 291-4.
- Deyo R A, Pope R M, Persellin R H. Interference by rheumatoid factor with the detection of C-reactive protein by the latex agglutination method. *J Rheumatol* 1980; **7**: 279-87.
- Zein N, Ganuza C, Kushner I. Significance of serum C-reactive protein elevation in patients with systemic lupus erythematosus. *Arthritis Rheum* 1979; **22**: 7-12.
- Morrow W J W, Isenberg D A, Parry H F, Snaith M L. C-reactive protein in sera from patients with systemic lupus erythematosus. *J Rheumatol* 1981; **8**: 599-604.
- Mancini G, Carbonara A O, Hermans J F. Immunochemical quantification of antigens by single radial immunodiffusion. *Immunochemistry* 1965; **2**: 235-54.
- Becker G J, Waldburger M, Hughes G R V, Pepys M B. Value of C-reactive protein measurement in the investigation of fever in systemic lupus erythematosus. *Ann Rheum Dis* 1980; **39**: 50-2.
- Pincus T. Immunochemical conditions affecting the measurement of DNA antibodies using ammonium sulphate precipitation. *Arthritis Rheum* 1971; **14**: 623-30.
- Nydegger U E, Lambert P H, Gerber H, Miescher P A. Circulating immune complexes in the serum in systemic lupus erythematosus and in carriers of hepatitis B antigen: quantitation by binding to radiolabelled Clq. *J Clin Invest* 1974; **54**: 297-309.
- Siegel S. *Non-parametric statistics for the behavioural sciences*. New York: McGraw-Hill, 1956.
- McConkey B, Crookson R A, Crookson A P. The assessment of rheumatoid arthritis. *Q J Med* 1972; **41**: 115-25.
- Amos R S, Constable T J, Crookson R A, Crookson A P,

- McConkey B. Rheumatoid arthritis: relation of serum C-reactive protein and erythrocyte sedimentation rates to radiographic changes. *Br Med J* 1977; **1**: 195-7.
- 17 Pepys M B. C-reactive protein fifty years on. *Lancet* 1981; **1**: 653-7.
- 18 Lambert P H, Dixon F J, Zubler R H, *et al.* A WHO collaborative study for the evaluation of eighteen methods for detecting immune complexes in serum. *J Clin Lab Immunol* 1978; **32**: 1253-88.
- 19 Pereira da Silva J A, Elkon K B, Hughes G R V, Dyck R F, Pepys M B. C-reactive protein levels in systemic lupus erythematosus: a classification criterion? *Arthritis Rheum* 1980; **23**: 770-1.
- 20 de Beer F C, Fagan E A, Hughes G R V, Mallya R K, Lanham J G, Pepys M B. Serum amyloid-A protein concentration in inflammatory diseases and its relationship to the incidence of reactive systemic amyloidosis. *Lancet* 1982; **ii**: 231-234.
- 21 Mallya R K, Vergani D, Tee D E H, *et al.* Correlation in rheumatoid arthritis of concentrations of plasma C3d, serum rheumatoid factor, immune complexes and C-reactive protein with each other and with clinical features of disease activity. *Clin Exp Immunol* 1982; **48**: 747-53.