EXTENDED REPORT

Nerve growth factor inhibition with tanezumab influences weight-bearing and subsequent cartilage damage in the rat medial meniscal tear model

Timothy P LaBranche, Alison M Bendele, Brian C Omura, Kathryn E Gropp, Susan I Hurst, Cedo M Bagi, Thomas R Cummings, Lonnie E Grantham II, David L Shelton, Mark A Zorbas

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ABSTRACT

Objective To investigate whether the effects of nerve growth factor (NGF) inhibition with tanezumab on rats with medial meniscal tear (MMT) effectively model rapidly progressive osteoarthritis (RPOA) observed in clinical trials.

Methods Male Lewis rats underwent MMT surgery and were treated weekly with tanezumab (0.1, 1 or 10 mg/kg), isotype control or vehicle for 7, 14 or 28 days. Gait deficiency was measured to assess weight-bearing on the operated limb. Joint damage was assessed via histopathology. A second arm, delayed onset of treatment (starting 3–8 weeks after MMT surgery) was used to control for analgesia early in the disease process. A third arm, mid-tibial amputation, evaluated the dependency of the model on weight-bearing.

Results Gait deficiency in untreated rats was present 3–7 days after MMT surgery, with a return to normal weight-bearing by days 14–28. Prophylactic treatment with tanezumab prevented gait deficiency and resulted in more severe cartilage damage. When onset of treatment with tanezumab was delayed to 3–8 weeks after MMT surgery, there was no increase in cartilage damage. Mid-tibial amputation completely prevented cartilage damage in untreated MMT rats.

Conclusions These data suggest that analgesia due to NGF inhibition during the acute injury phase is responsible for increased voluntary weight-bearing and subsequent cartilage damage in the rat MMT model. This model failed to replicate the hypotrophic bone response observed in tanezumab-treated patients with RPOA.

INTRODUCTION

Knee osteoarthritis (OA) is a condition characterised by pain, inflammation and functional disability. OA pain is complex and involves both inflammatory and neuropathic components mediated through persistent tissue injury and release of inflammatory mediators. Pain treatment for OA is problematic because many standard therapies provide minimal pain relief and do not address underlying mechanisms driving disease pathophysiology. The neurotrophin, nerve growth factor (NGF), is considered a key modulator of pain perception in several chronic pain conditions, including OA. Tanezumab, a humanised monoclonal antibody, binds NGF and prevents interaction with its receptors (high-affinity transmembrane tyrosine kinase receptor (TrkA) and the low-affinity NGF receptor (p75)). Tanezumab provided significant improvement in pain, physical function and patients’ global assessments in a number of chronic pain conditions.

Investigator reports of adverse events initially described as osteonecrosis leading to total joint replacement during the conduct of phase III clinical OA studies led the US Food and Drug Administration to put trials of all NGF inhibitors on partial clinical hold. Blinded adjudication of the results showed that there was no increase in osteonecrosis, nor frequency of total joint replacement with tanezumab monotherapy. Tanezumab treatment was however associated with increased incidence of rapidly progressive osteoarthritis (RPOA). A retrospective analysis of the data suggested ways to mitigate this risk, and based on these data the clinical hold was lifted to allow further trials to test these risk mitigation approaches.

The increased frequency of RPOA was unexpected, as no issues with bone or joints were seen in non-clinical studies of anti-NGF therapy using large multiples of the clinical dose. Also, no evidence of abnormal bone or joint phenotypes exists in humans with TrkA or p75 null mutations other than that observed in congenital pain insensitivity mutations. Last, in an experimental fracture model, anti-NGF therapy was shown to ameliorate fracture pain without impacting bone healing. Meniscal injury and acute meniscectomy are known to increase the risk of knee OA. Up to 80% of patients with knee OA, as well as a high percentage of age-matched controls, have evidence of meniscal injury at the time of diagnosis yet abnormal load-bearing and joint instability stemming from meniscal injury resulting in substantial OA lesions may take years. Attempts to model this in animals have produced varied results. The medial meniscal tear (MMT)-induced joint damage model in rats has many features attractive for an animal model. These animals show joint instability and tibial cartilage damage in as little as 7–14 days after surgery. MMT-induced joint damage lesions are highly reproducible and include articular cartilage proteoglycan loss, chondrocyte...
degeneration and loss of matrix. Although erosion of cartilage is a feature of this model, rarely does it progress to ulceration within 14 or 28 days. Subchondral bone sclerosis and osteophyte formation, which are compensatory responses to altered mechanical loading and joint instability are present in this model. 

The objective of this study was to characterise the impact of NGF inhibition with tanezumab on voluntary weight-bearing and subsequent articular cartilage damage in MMT rats to see if this model would be useful for investigating potential mechanisms underlying the clinical findings of increased RPOA in patients treated with tanezumab.

**METHODS**

**Animals and tanezumab administration**

For animal care and use, see online supplementary text S1.

All animals underwent MMT or sham surgery on study day 0. In MMT surgery rats, the medial meniscus in the right hind limb was cut through the full thickness to simulate a complete tear. In sham surgery animals, the knee was opened and the medial meniscus was touched only (not cut). Sham surgery rats received vehicle, and animals that underwent MMT surgery were treated with either isotype control (a non-specific antibody of the same type (immunoglobulin G) and class (2a) as tanezumab), vehicle (10 mM trehalose buffer) or tanezumab (0.1, 1 or 10 mg/kg) subcutaneously once a week for 7, 14 or 28 days.

In the first arm of the study, treatment began on the day of surgery (day 0, figure 1A). To examine the impact of analgesia on weight-bearing and lesion severity, a second arm evaluated delaying the onset of tanezumab treatment (0.1 mg/kg weekly) until gait deficiency was no longer evident (day 23); half the animals started receiving treatment on day 23, while the other half were delayed another month (day 57) before treatment was initiated (figure 1B). In both arms of this study, anti-drug antibodies in plasma were measured, as well as concentration of parent compound in serum to evaluate the potential for clearance of tanezumab via immunologic response.

To understand the role of weight-bearing in the progression of knee joint damage in the rat MMT model, a third arm of the study evaluated the impact of mid-tibial amputation on MMT lesion progression, eliminating weight-bearing as a variable. The knees were collected 14 days after MMT for micro-CT imaging, followed by histopathology (figure 1C). In the first two study arms, the impact of NGF inhibition on subchondral bone, osteophyte size and growth plate thickness was evaluated in MMT rats to assess secondary effects on the overall joint.

**Gait deficiency**

Difference in dynamic weight-bearing between the ipsilateral (MMT operated; right side) limb and contralateral (control; left side) limb is an established approach to measuring gait in rat and mouse models of OA. Voluntary weight-bearing by non-amputated MMT rats was assessed on days −3, 3, 7, 14 and 28 in the first arm of the study and additional days 37, 42, 56, 63 and 71 in the delayed-treatment study to confirm that there was no pain relapse in any animals (see online supplementary text S2).

**Histopathology and quantitative image analysis**

Microscopy-based semiquantitative scoring of cartilage and bone changes is an established endpoint for describing response to compounds in MMT rats. Cohorts of animals were sacrificed 7, 14 or 28 days after surgery or 14 days after treatment onset (day 23 or 57). Knees were collected at necropsy and evaluated via light microscopy by a veterinary pathologist as described (see online supplementary text S3). 

**Radiography and µCT imaging**

Knees were X-rayed with a MX20 specimen scanner (Faxitron Bioptics, Tucson, Arizona, USA) using recommended settings; exposure 12–18 s at 31–35 kV. Radiographs were used to assess gross anatomy of the region of interest (ROI) evaluated by µCT and to inspect bone samples for presence of abnormalities.

µCT was conducted on the right tibial epiphysis and metaphysis using a MicroCT 100 system (Scanco Medical, Bassersdorf, Switzerland) with the following parameters: 800 slices, 10 µm resolution, total scanned area of 8.0 mm² and source energy of 70 kVp, 115 µA at 8 W to capture the entire proximal tibia
section. Cortical and cancellous bone of the entire epiphysis were scanned and analysed in an ROI on 100 consecutive slices with 1.0 mm thickness that best represented the central segment of the epiphysis. Parameters included tissue volume (TV), bone volume (BV), BV/TV ratio and bone mineral density (BMD). Cortical and cancellous bone of the proximal metaphysis were scanned and analysed in an ROI on 100 consecutive slices 1 mm below the growth plate with a thickness of 1.0 mm that best represented the central metaphysis segment. Parameters included TV, BV, BV/TV, trabecular number, trabecular thickness, trabecular separation and BMD.

**Pharmacokinetics and anti-drug antibody**

To evaluate the potential for clearance of compound via immunologic response, we measured anti-drug antibodies in plasma as well as concentration of tanezumab in serum pharmacokinetic (PK) samples. Blood samples were collected on days 3, 7, 14 and 21. An additional PK sample was taken 4 days prior to termination. In the delayed-treatment study, blood samples for PK and anti-drug antibodies were collected on days 16, 30, 57 and 64. PK plasma concentrations of tanezumab were measured using a validated ELISA at ICON Development Solutions (Whitesboro, New York, USA). The lower limit of quantitation in Lewis rat plasma was 50 ng/mL. Concentrations below the limit of quantitation were treated as 0 ng/mL in the PK calculations. Anti-drug antibodies concentrations in serum were measured using a validated ELISA at ICON Development Solutions. Final assessment of induction of an anti-drug antibodies response was based on comparison of predose and postdose results for each animal.
**Statistical analyses**

Body weight change, weight-bearing (gait) deficiency at each time point and histopathology outcomes were analysed. Vehicle and sham surgery groups were compared with the isotype group using a two-sided t test (with a Welch correction for unequal variances if necessary) or Mann-Whitney U test. Each tanezumab-treated group was compared with the isotype group using a two-sided Dunnett or Dunn multiple comparisons test. Significance for all tests was set at \( p \leq 0.05 \). Gait deficiency was assessed by Dunnett and t-tests.

**RESULTS**

**Tanezumab treatment influences weight-bearing and subsequent cartilage damage in the rat MMT model**

Although MMT rats with intact hind limbs appeared to ambulate normally after surgery, they experienced a measurable weight-bearing (gait) deficiency on the operated limb. Gait deficiency in MMT rats administered vehicle or isotype control peaked 3 days after surgery (11.8% and 12.2% decrease, respectively), continued through day 7 and resolved by days 14–28 (figure 2A). In contrast, tanezumab-treated MMT rats exhibited no gait deficiency at any point, indicating that tanezumab-treated MMT rats were bearing full weight on the operated limbs throughout the study (figure 2). Tanezumab-treated rats exhibited more severe total tibial cartilage substantial degeneration width than either vehicle or isotype controls on days 7, 14 and 28 (figure 2B). No significant differences in body weight gain were observed over the course of the study. Focal areas of alopecia along the mouth/muzzle began developing on day 14 and were seen in most animals in the 1 mg/kg tanezumab group by day 28. Gait analysis was also performed as part of the delayed-treatment study (see online supplementary table S1).

There was evidence of a robust, compensatory (hypertrophic) response to the partially destabilised joint. Namely, a time-dependent increase in subchondral bone sclerosis was observed in untreated MMT rats (table 1), which was exacerbated by tanezumab (statistically significant greater subchondral sclerosis compared with isotype controls on days 7, 14 and 28). Larger osteophyte size was also observed in tanezumab-treated MMT rats compared with isotype controls on days 7 (tanezumab 10 mg/kg, \( p < 0.05 \)), 14 (tanezumab 1 mg/kg, \( p < 0.01 \)) and 28 (tanezumab 0.1, 1 and 10 mg/kg, \( p < 0.001 \) for all; table 1), indicating a time–dose relationship with increasing effect. No osteophytes were observed in sham surgery animals. Last, a significant focal increase in growth plate thickness was also observed in tanezumab-treated MMT rats compared with isotype control-treated rats (1 and 10 mg/kg on days 14 and 28, \( p \leq 0.05 \), table 1).

**Delaying onset of tanezumab treatment protects against cartilage damage in MMT rats**

Gait deficiency began shortly after MMT surgery and appeared to resolve by days 14–28. To confirm whether changes in weight-bearing during this early postoperative period was the principal driver of increased joint damage in the tanezumab-treated MMT rats, we delayed the onset of treatment from day 0 to day 23 (cohort 1) or day 57 (cohort 2). The second cohort in particular initiated treatment well past the time that gait abnormalities normalised. Tanezumab-treated MMT rats had significantly greater cartilage degeneration width than MMT controls when treatment began on day 23 (\( p = 0.035 \), figure 3). However, delaying treatment until weight-bearing fully returned to normal (ie, day 57 but not day 23) resulted in complete protection from worsening of joint damage when compared with control treatments (figure 3).

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**Table 1  Histopathology summary, n=10/group; mean (SE)**

<table>
<thead>
<tr>
<th>Parameter/day</th>
<th>Isotype control</th>
<th>Vehicle control</th>
<th>Sham surgery</th>
<th>Tanezumab 0.1 mg/kg/week</th>
<th>Tanezumab 1 mg/kg/week</th>
<th>Tanezumab 10 mg/kg/week</th>
</tr>
</thead>
<tbody>
<tr>
<td>Growth plate difference (medial-lateral), μm</td>
<td>41.33 (5.43)</td>
<td>33.33 (2.22)</td>
<td>5.33 (2.95)***</td>
<td>50.67 (10.29)</td>
<td>60.00 (8.94)</td>
<td>70.67 (11.60)</td>
</tr>
<tr>
<td>Medial collateral ligament, μm</td>
<td>865.33 (26.70)</td>
<td>893.33 (13.91)</td>
<td>870.67 (35.84)</td>
<td>1068.00 (33.46)***</td>
<td>1041.33 (48.42)***</td>
<td>1128.00 (60.65)***</td>
</tr>
<tr>
<td>Bone damage score†</td>
<td>2.00 (0.15)</td>
<td>2.00 (0.00)</td>
<td>0.00 (0.00)***</td>
<td>2.40 (0.16)</td>
<td>2.20 (0.13)</td>
<td>2.20 (0.13)</td>
</tr>
<tr>
<td>Bone sclerosis score†</td>
<td>3.00 (0.26)</td>
<td>3.40 (0.16)</td>
<td>0.00 (0.00)***</td>
<td>4.60 (0.16)**</td>
<td>4.40 (0.24)**</td>
<td>4.20 (0.29)*</td>
</tr>
<tr>
<td>Tibial osteophyte size, μm</td>
<td>119.33 (26.79)</td>
<td>121.33 (26.88)</td>
<td>0 (0.00)***</td>
<td>167.33 (21.79)</td>
<td>150.33 (29.65)</td>
<td>223.67 (14.59)*</td>
</tr>
</tbody>
</table>

*\( p < 0.05 \) versus isotype control; **\( p < 0.01 \) versus isotype control; ***\( p < 0.001 \) versus isotype control.

†Damage to calcified cartilage and subchondral bone was scored on a 6-point numerical scale (0=no changes to 5=increased basophilia; marked to severe fragmentation of calcified cartilage, mesenchymal change in marrow involves up to three-fourths of the total area and articular cartilage has collapsed into the epiphysis to a depth of >250μm from tidemark).

‡Medial tibial subchondral/epiphysial bone sclerosis was scored on a 6-point numerical scale (0=no changes to 5=76%–100% increase in subchondral or epiphyseal trabecular bone thickness in medial versus lateral; very little marrow space remains in medial tibia).
Limb amputation protects against joint damage in untreated MMT rats
As reported,26 as well as in this study (figure 4) signs of tibial articular cartilage damage appear in normal weight-bearing MMT rats in as early as 7 days after surgery. In this study, those changes were prevented by tibial amputation (figure 4). MMT rats with tibial amputations are not placing weight on their operated limbs, as evidenced by less cortical and cancellous bone in amputated rats than unamputated rats, regardless of whether they had MMT surgery performed (table 2 and figure 4C).

PK and anti-drug antibody levels
For PK and anti-drug antibodies results, see online supplementary text S4.

DISCUSSION
The serious joint adverse events observed in placebo-controlled clinical trials of anti-NGF antibodies have been adjudicated as either normal or RPOA, the latter characterised by atrophic bone with subchondral insufficiency and/or occasional foci of secondary osteonecrosis.33 This clinical finding was unexpected, since extensive non-clinical safety studies failed to find adverse events of bone or joint. The current study was undertaken to see if a commonly used and relevant arthritis model, the rat MMT, would be useful for further understanding the finding.

Treating MMT rats with tanezumab did not recapitulate the atrophic bone destruction associated with RPOA in clinical trials. Although anti-NGF treatment early in the MMT disease process influenced weight-bearing and drove additional cartilage damage, secondary changes in subchondral bone were instead hypertrophic (subchondral bone thickening, with osteophytes present). There were however notable findings related to weight-bearing and its impact on joint damage, highlighting that this model is ideal for evaluating the relationship between joint instability, changes in weight/load-bearing and health of articular cartilage damage.24 27 Cartilage damage and subsequent effects on bony tissues (ie, subchondral bone sclerosis and osteophyte formation) appear to be exquisitely sensitive to weight-bearing early in the rat MMT disease process (ie, 0–14 days after meniscal tear). In contrast, treatment in the chronic phase after gait deficiency had already normalised, similar to the clinical trial experience, had no adverse effects on the joint.

Mid-tibial amputation protected against MMT-induced joint damage, further suggesting that dynamic weight-bearing is the key driver for joint damage in this model. For instance, there was more severe decreased bone volume in the proximal tibial metaphysis of amputated rats versus normal rats. The idea that weight-bearing drives joint damage in the MMT rat is also supported by observations that MMT rats given intra-articular injections of irritating substances leading to joint swelling and pain actually result in decreased cartilage damage and bone sclerosis scores, largely because the rats are unwilling to bear weight on the joint (A. Bendele, unpublished observations).30

In this study, MMT rats exhibited gait deficiency on the operated limb shortly after surgery, which returned to baseline by days 14–28. Transection of the meniscus leads to this gait abnormality, not general surgery trauma, as animals undergoing sham surgery exhibited no change in weight-bearing at any point. Treatment with tanezumab at ≥0.1 mg/kg resulted in MMT rats behaving similarly to sham surgery rats, since they did not exhibit altered weight-bearing. These results, combined with other work investigating the role ofNGF in a meniscal transection rat model of OA, suggest that analgesia can significantly influence voluntary weight-bearing.

Figure 4  The rat MMT model is dependent on dynamic weight-bearing for cartilage damage. Rats underwent either MMT or sham surgery ± amputation of the lower tibia. (A) Tibial cartilage substantial degeneration width (SE), as determined by histopathology at day 14 after MMT surgery was observed in MMT rats (no amputation). (B) Representative photomicrographs from each group (16× magnification). (C) Top row depicts X-ray images of the right knee. Second row shows representative two-dimensional μCT images of the proximal tibia (arrows indicate less cancellous bone at metaphysis). Third row shows the image of tibial epiphysis (arrow indicates thicker subchondral bone at medial tibial plateau and arrowhead points at osteophyte). Bottom row shows proximal tibial metaphysis (square indicates areas of secondary spongiosa where cancellous bone has been evaluated). MMT, medial meniscal tear; μCT, micro-CT.
Results suggest that at least two separable phases of pain behaviours occur in the rat MMT model: (1) an early phase, in which gait abnormality is present and dynamic weight-bearing significantly alters the future trajectory of joint damage, and (2) a later phase, in which there is no detectable gait abnormality and tissue destruction is no longer sensitive to NGF inhibition. From this, we propose that increased joint damage seen in tanezumab-treated MMT rats was due to an increase in voluntary weight-bearing early in the disease process and not due to direct toxic effect of tanezumab on cartilage or bone. To support this, an additional arm of the study was conducted delaying onset of tanezumab treatment from day 0 to day 23 or 57 after MMT surgery when gait deficiency was no longer present. Overall, the results presented here support the conclusion that analgesia may exacerbate joint damage when it is present during the early phase. This is likely due to mechanical overloading shortly after meniscal injury. The relevance of this model to RPOA reported in human clinical trials should be considered. Specifically, results from this study may help guide future clinical trial design by suggesting exclusion criteria (i.e., patients deemed sensitive to mechanical overloading/radiographic evidence of bone or joint insufficiency).

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Contributors TPL, KEG, TRC, DLS and MAZ designed the study. TPL, KEG, TRC, DLS and MAZ were involved in data interpretation and writing and reviewing the manuscript. TPL, KEG, TRC, DLS and MAZ also acknowledge Drs Larry Whiteley, Nasir Khan and Lloyd Dethloff (Pfizer) for their guidance and critical review of this work. Medical writing support was provided by Joseph Oleynek at Engage Scientific Solutions and was funded by Eli Lilly & Co. and Pfizer.

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Competing interests KEG, SIH, CM, TRL and MAZ are employees of Pfizer and own stock and/or stock options in Pfizer. TPL and DLS were employees of Pfizer at the time of these studies and own stock and/or stock options in Pfizer. LEG is a contracted resource of and owns stock in Pfizer.

Ethics approval Oversight by local institutional animal care and use committees.

Table 2 Bone parameters evaluated ex vivo by μ-CT at proximal tibial epiphysis and metaphysis

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Normal rats</th>
<th>MMT surgery</th>
<th>MMT plus amputation</th>
<th>Sham plus amputation</th>
</tr>
</thead>
<tbody>
<tr>
<td>TV, mm³</td>
<td>1.63±0.11</td>
<td>1.87±0.14</td>
<td>1.83±0.07</td>
<td>1.67±0.11</td>
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<td>BV, mm³</td>
<td>0.76±0.10</td>
<td>0.74±0.09</td>
<td>0.67±0.04±1,4</td>
<td>0.64±0.05±1,4</td>
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<tr>
<td>BV/T, ratio</td>
<td>0.46±0.03</td>
<td>0.43±0.02</td>
<td>0.37±0.03±1,4</td>
<td>0.38±0.02±1,4</td>
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<tr>
<td>BMD, g/cm²</td>
<td>985.14±16.44</td>
<td>990.85±14.89</td>
<td>983.37±14.97</td>
<td>1005.05±11.05</td>
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Proximal tibial metaphysis (cortical and cancellous bone)

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Normal rats</th>
<th>MMT surgery</th>
<th>MMT plus amputation</th>
<th>Sham plus amputation</th>
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<tbody>
<tr>
<td>TV, mm³</td>
<td>2.81±0.05</td>
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<tr>
<td>BV, mm³</td>
<td>0.73±0.06</td>
<td>0.57±0.06††</td>
<td>0.24±0.07†††,†††</td>
<td>0.26±0.05†††,†††</td>
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<tr>
<td>BV/T, ratio</td>
<td>0.26±0.02</td>
<td>0.20±0.02†</td>
<td>0.08±0.02†††,†††</td>
<td>0.09±0.02†††,†††</td>
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<tr>
<td>BMD, g/cm²</td>
<td>929.74±32.06</td>
<td>922.53±57.59</td>
<td>848.15±40.35</td>
<td>943.72±68.42</td>
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Proximal tibial epiphysis (cortical and cancellous bone)

<table>
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<tr>
<th>Parameter</th>
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<th>MMT surgery</th>
<th>MMT plus amputation</th>
<th>Sham plus amputation</th>
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<tr>
<td>TV, mm³</td>
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<td>BV, mm³</td>
<td>0.15±0.02</td>
<td>0.07±0.01††</td>
<td>0.01±0.01†††,†††</td>
<td>0.03±0.02†††,†††</td>
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<tr>
<td>BV/T, ratio</td>
<td>0.16±0.02</td>
<td>0.08±0.01††</td>
<td>0.01±0.01†††,†††</td>
<td>0.03±0.02†††,†††</td>
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<tr>
<td>TbN, 1/mm³</td>
<td>2.82±0.35</td>
<td>1.65±0.26††</td>
<td>0.32±0.19†††,†††</td>
<td>0.63±0.32†††,†††</td>
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<tr>
<td>TbT, μm³</td>
<td>0.056±0.002</td>
<td>0.048±0.005†</td>
<td>0.039±0.007†††,†††</td>
<td>0.041±0.010†††</td>
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<tr>
<td>TbS, μm³</td>
<td>0.30±0.04</td>
<td>0.57±0.09††</td>
<td>4.94±6.64†††,†††</td>
<td>2.01±0.23†‡‡,†‡‡</td>
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<tr>
<td>BMD, g/cm²</td>
<td>918.43±19.86</td>
<td>923.92±37.10</td>
<td>864.96±41.79</td>
<td>921.60±45.66</td>
</tr>
</tbody>
</table>

1p<0.05 versus normal; 2p<0.05 versus MMT; 3p<0.01 versus normal; 4p<0.01 versus MMT; 5p<0.001 versus normal and 6p<0.001 versus MMT.

BMD, bone mineral density; BV, bone volume; MMT, medial meniscal tear; TbN, trabecular number; TbS, trabecular separation; TbT, trabecular thickness; TV, tissue volume; μ-CT, micro-CT.

Provenance and peer review Not commissioned; externally peer reviewed.

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Basic and translational research

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