Autoantibodies binding to citrullinated telopeptide of type II collagen and to cyclic citrullinated peptides predict synergistically the development of seropositive rheumatoid arthritis

Short running title: Autoantibodies to citrullinated collagens and anti-CCPs in RA patients

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ABSTRACT

Objectives: To find out whether autoantibodies to citrullinated telopeptides of type I and II collagens and to cyclic citrullinated peptides (CCPs) predict the development of rheumatoid arthritis (RA).

Methods: A case-control study (matched for sex, age and municipality) was nested within a Finnish cohort of 19072 adults who had neither arthritis nor a history of it at the baseline examination during 1973-77. 124 subjects developed RA by late 1989 and of them 89 were positive for rheumatoid factor (RF). Preillness serum specimens were analysed for autoantibodies against arginine (A) or citrulline (C) containing synthetic telopeptides using a chemiluminescence method and for anti-CCPs Mark2 with an ELISA method.

Results: The mean levels of autoantibodies to citrulline-containing telopeptides and the C/A ratios of type I and II collagens and to CCP were higher in subjects who later developed RF-positive RA. In the highest tertiles of C/A (I), C/A (II) ratios and anti-CCPs levels the relative risk of RF-positive RA was significantly increased. In the multifactorial model only anti-CCPs retained its statistical significance. However, the interaction term of C/A (II) ratio and anti-CCPs proved statistically significant (p = 0.02). The subjects ranked into the highest tertiles of both C/A (II) ratio and anti-CCPs had an odds ratio of 20.06 (95% confidence interval, 4.37-92.06) of developing RF-positive RA compared with those in the lowest tertiles of these antibodies. None of the autoantibodies predicted RF-negative RA.

Conclusion: Autoantibodies to citrullinated telopeptides of type I and II collagen and to CCPs exert a synergistic effect on the risk of seropositive RA.

Key Terms: Antibodies to cyclic citrullinated peptides (anti-CCPs)
Antibodies to citrullinated type I and II collagens
Rheumatoid arthritis (RA)
INTRODUCTION

Rheumatoid arthritis (RA) is a systemic autoimmune disease of unknown aetiology, characterized by chronic joint inflammation that often leads to joint destruction. Both environmental and genetic factors play a role in the etiopathogenesis of the disease (1). It was recently reported that one environmental factor, smoking, can trigger immune reactions characteristic of RA (2) and the appearance of antibodies to citrullinated proteins (3). Citrullination of proteins is a normal post-translational process occurring late during the protein lifespan, e.g. in filaggrin and vimentin during their apoptotic degradation. Several autoantibodies found in RA are directed to epitopes in citrullinated proteins (4, 5). Citrulline is formed from arginine residues in peptides or proteins by the enzyme peptidylarginine deiminases (PADs). There are at least 5 genetically different forms of PADs of which two isotypes, PAD2 and PAD4, have been found in monocytes and macrophages of inflamed synovium. There is also polymorphism related to rheumatoid arthritis (6-8). It has been hypothesized that during rheumatic synovitis PAD is released into the extracellular space inducing locally citrullination of arginine residues in several proteins, e.g. in vimentin, fibrin and fibrinogen, fibronectin, α-enolase, keratin, initial factor 4G1 and finally type I and II collagens (4, 5, 9-15).

Several autoantibodies, such as RF and filaggrin related antibodies (antikeratin antibodies, antiperinuclear factor, and antibodies to human filaggrin) as well as anti-CCPs have been shown to precede the onset of clinical RA, often by many years (16-21). The frequency of positive reactions is dependent on the time elapsed from taking the serum specimen to the onset of clinical disease (17, 21).

Anti-CCPs antibodies are currently the most specific markers of RA and have been widely used for diagnostic purposes (20, 22). According to the recent review of Avouac et al (23) the mean (+ SD) sensitivity value for the second generation anti-CCP assays was 68 (+15) % in 58 articles published in 1999-2006. The sensitivity of anti-CCP2 is close to that of RF whereas the specificity in differentiating RA from other rheumatic diseases is strikingly high, 96 (+3) %.

The inciting stimuli for the production of antibodies to citrullinated peptide and protein epitopes are not known, nor is it known whether these antibodies participate in the rheumatic inflammation. On the other hand, it has long been supposed that autoimmunity against collagens might be involved in the pathogenesis of RA. In our laboratory the enzyme-linked immunosorbent assays (ELISAs) were first used to the existence of autoantibodies reacting against citrullinated peptides derived from type I and type II collagens in patients with RA (15). Secondly we developed more sensitive methods for these autoantibodies which were used in this study. In these automatic CLIA methods 38% of early RA patients showed increased binding of synthetic citrullinated carboxytelopeptide derived from α1 chain of type I collagen. For corresponding carboxytelopeptide from the α1 chain of type II collagen, 35 patients’ sera out of 120 early RA patients bound the citrullinated peptide more strongly than the arginine peptide (24).

Many antigens will be citrullinated and many antibodies are formed against these antigens. The anti-CCP2 is an artificial antigen of unknown origin that is kept secret for commercial reasons. Now we want to test whether citrullinated collagen peptides are also predictive for development of RA and add to the predictive power of anti-CCP2.

SUBJECTS AND METHODS

Patients and controls

During 1973-1977, the Social Insurance Institution’s Mobile Clinic Unit carried out multiphasic health examinations in 12 municipalities in four regions on Finland (25-27). In each of the four regions, all inhabitants or a random sample of inhabitants of one rural municipality and one urban or semi urban municipality as well as the employees of one factory were invited to attend the examination (25). A total of 19518 subjects who were 20 years of age or older (83% of those
invited) participated in the examinations. Of them 446 had a previous history of inflammatory arthritis and were excluded from the current study. The mean age of the subjects was 45 years (range 20-98).

The population at risk (19072 subjects) consisted of participants who had no previous history of arthritis or other rheumatic disease, according to the Social Insurance Institution’s population register or to a questionnaire administrated at the baseline examination. Morbidity and mortality of all examinees were followed up continuously. Participants in the survey who later developed chronic arthritis were identified by linking the survey data with the Social Insurance Institution’s population register up to the end of 1989, using the unique identification code assigned to each Finnish citizen. In Finland, reimbursed drugs are provided for certain rheumatic diseases, including chronic inflammatory rheumatic diseases. Eligibility requires a comprehensive medical certificate written by the attending physician and approved by an expert adviser (specially trained physician) on behalf of the sickness insurance system. Nearly all certificates are based on examinations and follow-up experience at special rheumatology clinics. These written documents were carefully reviewed to identify incident cases of RA on the basis of the following criteria: a) The patient had been granted the right to specially reimbursed medication because of a physician’s diagnosis of clinical RA; b) there was no history of rheumatoid disease at the health examination; and c) according to the doctor’s certificate, the onset of the disease occurred after the health examination. Information on the RF status was obtained from the certificates. Testing had been performed in several different laboratories. Sufficient information concerning diagnosis and onset of disease was obtained in 124 cases, of which 89 were RF positive by the time of the diagnosis (16).

The certificates are written to provide evidence that a patient has the disease and needs treatment, but they are not keyed to any set of criteria. Most patients with RA entitled to reimbursement for drug treatment meet, however, the revised criteria of the American Rheumatism Association, and the sensitivity of the sickness insurance data exceeds 90% for seropositive RA (28, 29). Half of the RF-negative cases have been reported as having radiographic changes typical for RA (30).

A nested case-control design was applied to study anti-CCPs for its prediction of clinical RA. Three controls per case were selected by individual matching, using sex, age, and municipality as matching factors. Exact matching was performed for sex and municipality, whereas nearest available matching was used for age within the sex-municipality strata. Matching for municipality also controlled for both the time of the baseline examination and for the duration of storage of serum samples (16).

Assays for detecting autoantibodies against collagen telopeptides

Blood samples were drawn at the mobile clinic and sera stored frozen at -20°C until used for testing. The principle of assays for detecting autoantibodies against collagen telopeptides has been described previously (24). In brief, measurements were carried out with two-site chemiluminescence immunoassays, which detect IgG antibodies against the synthetic C-telopeptides of the α1 chain of human type I and II collagens (EKAGDGGRYYXA and EKGPDPLQYMXA, where X stands for arginine or citrulline. Peptides are from NeoMPS, Strasbourg, France). The serum samples were diluted 1:10 and incubated with the above biotintylated peptide and magnetic particles. After subsequent washing, acridinium-labeled anti-human IgG antibodies were added to the reaction mixture. The wells containing the washed magnetic particles with antigen/antibody/complex were transferred to the system luminometer, which automatically injected triggers, initiating the chemiluminescence reaction. Light was quantified by a luminometer and expressed as relative luminescence units (RLU). The system background was measured daily by running the System Performance Controls provided by Nichols Institute Diagnostics (San Clemente, CA, USA). To assure the validity of the results, controls with three different concentrations were measured. These controls included negative, low, and high titres...
and their coefficient of variance were between 2 – 15 % (see Table 2 in reference 24). We could not analyze A (II) from five sera, A (I) and C (I) from 44 sera due to problems with the instrument. This means that results regarding A/C (I) has become more conservative and the statistical power is less than for A/C (II).

Enzyme immunoassays for anti-CCP2 antibodies

Anti-CCPs antibodies were measured in duplicates in the sera using the Immunoscan RA (Mark 2) kits from Euro Diagnostica (Malmö, Sweden) according to the manufacture’s instructions. Cut-off limit for positive result in the assay was 25 U/ml. The standard curve was linear to the level of the provided negative serum which gave in 12 successive assays an average (+ SD) of 12.8 (+ 0.84) U/ml. The interassay variation (reproducibility) for the negative serum was 6.6% and that for the lowest standard (25 U/ml) 3.2%. Thus, although the absolute levels below the lowest standard may be inaccurate, the ranking is correct for fractile divisions.

Statistical analysis

Associations between different autoantibodies, antibody ratios, sex and age were assessed with general linear model. The significance of the differences in mean levels of serum antibodies between RA cases and their individually matched controls was tested by the paired t-test. The conditional logistic model (31) was used to estimate the associations between serum antibody levels and the risk of RA. Antibody levels were also entered as potential confounding factors into the model. The relative risks, estimated as odds ratios with 95% confidence intervals, were computed for tertiles of serum anti-CCP2s levels (<15.0; 15.0-16.2; ≥16.3 units), C/A (I) ratios (<0.86; 0.86-0.94; ≥0.95) and C/A (II) ratios (<0.85; 0.86-0.94; ≥0.95). The tertile divisions were based on the antibody distributions among controls. Effect-modification of each antibody was tested by entering interaction terms “tertile A * tertile B” in addition to tertiles A and B into the model. Statistical significance of interaction was tested with the likelihood ratio test based on the model and expressed as exact p-value.

Subgroup analyses were carried out among those whose time span from the baseline examination to the onset of RA was >5 years, and whose baseline anti-CCPs was within normal range (<25 U/ml), to control possible effects of early but still un-diagnosable RA on the results.

RESULTS

Baseline levels of different autoantibodies and antibody ratios in future cases with RA and their matched controls are presented in Table 1. The mean levels of antibodies to C (I), C (II) and CCPs, and C/A (I) and C/A (II) ratios were higher in cases than controls for RF-positive RA and RA in total, whereas antibodies to A (I) or A (II) showed no case-control difference in any comparison. No antibody level or C/A ratio differed between cases with RF-negative RA and their matched controls.

Among the future cases of any RA, men had significantly higher levels of anti-CCPs and a higher C/A (I) ratio than had women and individuals with high C/A (I) ratios had a significantly higher level of anti-CCPs and an almost significantly higher C/A (II) ratio (Table 2). Among the control subjects the correlations between sex, age and the antibodies were much weaker (Table 2).

In the highest tertiles of C/A (I) and C/A (II) ratios and, in particular, in the highest tertile of serum anti-CCPs the relative risk of RF-positive RA was significantly increased (Table 3). The three antibody predictors, however, tended to confound the effects of each other, and after entering of all the three into the multifactorial model only anti-CCPs retained its statistical significance.

An examination of possible effect-modification by sex and age on the association between each antibody level and the risk of RF-positive RA revealed no significant interaction (data not shown). However, the interaction term between the C/A (II) ratio and anti-CCPs levels proved statistically significant (p = 0.02). The subjects ranked into the highest tertiles of both the C/A (II) ratio and anti-CCPs had a relative risk of 20.06 (95% confidence interval, 4.37-92.06) for developing RF-
positive RA compared with those in the lowest tertiles of those antibodies. A synergistic effect-modification also seemed to exist for the C/A (I) ratio and anti-CCPs, but their interaction term was not statistically significant (Table 4). No effect-modification was suggested between the C/A (I) and (II) ratios (data not shown).

Data analysis was also restricted to time span of ≥5 years from baseline examination until the diagnosis of RA. The relative risks for follow-up times over five years did not notably differ from those for the entire follow-up (data not shown). The analysis was also carried out among those whose anti-CCPs levels were within normal limits (<25 U/ml). Again, no substantial alteration in the main results was observed. Odds ratios (95% confidence intervals) of seropositive RA for anti-CCPs concentrations of <15.0, 15.0-16.2 and 16.3-24.9 U/ml were 1.00 (reference category), 1.71 (0.67-4.36) and 3.08 (1.11-8.47), respectively.

Smoking neither showed any association with the levels of anti-CCP or C/A (I) or C/A (II); nor any confounding effect on the results. The interaction term between the C/A (II) ratio and anti-CCP levels remained statistically significant (p = 0.02), when smoking history (never smoked; quit; <20 cigarettes/day; ≥20 cigarettes/day) was entered into the model as a potential confounding factor.
Table 1. Average levels of antibodies to arginine (A) and citrulline (C) containing telopeptides of type I and II collagens, the corresponding C/A ratios and anti-CCP antibodies in future patients with rheumatoid arthritis (RA) and their controls.

<table>
<thead>
<tr>
<th>Baseline variable</th>
<th>RF-positive RA</th>
<th>RF-negative RA</th>
<th>RA in total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Case mean (SD)</td>
<td>Control mean (SD)</td>
<td>p</td>
</tr>
<tr>
<td>Number of females (%)</td>
<td>58 (65.2)</td>
<td>27 (77.1)</td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>45.6 (13.1)</td>
<td>45.6 (13.0)</td>
<td>0.41</td>
</tr>
<tr>
<td>A (I) (unit)</td>
<td>10.4 (6.6)</td>
<td>9.8 (3.4)</td>
<td>0.43</td>
</tr>
<tr>
<td>C (I) (unit)</td>
<td>19.3 (42.1)</td>
<td>8.8 (2.8)</td>
<td>0.03</td>
</tr>
<tr>
<td>C/A (I) (ratio)</td>
<td>1.9 (4.4)</td>
<td>0.9 (0.1)</td>
<td>0.04</td>
</tr>
<tr>
<td>A (II) (unit)</td>
<td>10.4 (5.3)</td>
<td>10.0 (2.9)</td>
<td>0.47</td>
</tr>
<tr>
<td>C (II) (unit)</td>
<td>15.2 (28.0)</td>
<td>9.0 (2.5)</td>
<td>0.04</td>
</tr>
<tr>
<td>C/A (II) (ratio)</td>
<td>1.6 (3.3)</td>
<td>0.9 (0.1)</td>
<td>0.06</td>
</tr>
<tr>
<td>Anti-CCP (unit)</td>
<td>219.1 (489.1)</td>
<td>16.0 (1.5)</td>
<td>0.0002</td>
</tr>
</tbody>
</table>
Table 2. Age and sex-adjusted mean values ($\overline{x}$) of anti-CCP antibodies and of antibody ratios to citrulline/arginine containing (C/A) (I) and (II) C-telopeptides by sex and tertiles of age and each antibody among future cases of rheumatoid arthritis and their controls. $r =$ partial correlation coefficient based on continuous variables and $p =$ statistical significance for its difference from zero.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Category</th>
<th>Cases of RA</th>
<th>Controls</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Category Tertile *</td>
<td>Anti-CCP (unit)</td>
<td>C/A (I) ratio</td>
</tr>
<tr>
<td></td>
<td></td>
<td>$\overline{x}$ $r$ $p$</td>
<td>$\overline{x}$ $r$ $p$</td>
</tr>
<tr>
<td>Sex †</td>
<td>Men</td>
<td>311.7 108.9 0.23 0.01</td>
<td>2.8 1.2 0.19 0.04</td>
</tr>
<tr>
<td></td>
<td>Women</td>
<td>108.9 131.7 0.23 0.01</td>
<td>1.3 1.4 0.01 0.88</td>
</tr>
<tr>
<td>Age§ (years)</td>
<td>&lt; 41</td>
<td>139.8 183.3 0.06 0.83</td>
<td>1.0 2.6 0.20 0.11</td>
</tr>
<tr>
<td></td>
<td>41 – 50</td>
<td>183.3 139.8 0.06 0.83</td>
<td>1.3 1.0 1.0 1.0</td>
</tr>
<tr>
<td></td>
<td>≥ 51</td>
<td>195.0 183.3 0.06 0.83</td>
<td>1.3 1.0 1.0 1.0</td>
</tr>
<tr>
<td>Anti-CCP (unit)</td>
<td>Lower</td>
<td>1.3 1.1 0.11 0.49</td>
<td>1.1 0.9 0.12 0.40</td>
</tr>
<tr>
<td></td>
<td>Medium</td>
<td>1.3 1.1 0.11 0.49</td>
<td>1.1 0.9 0.12 0.40</td>
</tr>
<tr>
<td></td>
<td>Higher</td>
<td>1.3 1.1 0.11 0.49</td>
<td>1.1 0.9 0.12 0.40</td>
</tr>
<tr>
<td>C/A (I) ratio</td>
<td>Lower</td>
<td>99.4 39.5 0.29 0.01</td>
<td>1.0 0.8 0.23 0.06</td>
</tr>
<tr>
<td></td>
<td>Medium</td>
<td>99.4 39.5 0.29 0.01</td>
<td>1.0 0.8 0.23 0.06</td>
</tr>
<tr>
<td></td>
<td>Higher</td>
<td>312.6 312.6 0.29 0.01</td>
<td>1.0 0.8 0.23 0.06</td>
</tr>
<tr>
<td>C/A (II) ratio</td>
<td>Lower</td>
<td>82.4 164.7 0.15 0.25</td>
<td>1.1 0.8 0.22 0.07</td>
</tr>
<tr>
<td></td>
<td>Medium</td>
<td>236.8 236.8 0.15 0.25</td>
<td>1.1 0.8 0.22 0.07</td>
</tr>
</tbody>
</table>

* The tertiles are based on the distributions among controls.
† Adjusted for age only
§ Adjusted for sex only
Table 3. Relative odds (OR’s), with 95% confidence intervals (CI’s), of rheumatoid factor positive rheumatoid arthritis between tertiles* of anti-CCP antibodies and of ratios of antibodies to citrulline/arginine containing (C/A) (I) and (II) C-telopeptides.

<table>
<thead>
<tr>
<th>Antibody</th>
<th>Tertile*</th>
<th>Number of cases</th>
<th>Number of controls</th>
<th>OR</th>
<th>(95% CI)</th>
<th>OR</th>
<th>(95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Anti-CCP (unit)</td>
<td>Lower</td>
<td>15</td>
<td>83</td>
<td>1.00</td>
<td></td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Medium</td>
<td>18</td>
<td>88</td>
<td>1.84</td>
<td>(0.78 – 4.34)</td>
<td>1.91</td>
<td>(0.76 – 4.80)</td>
</tr>
<tr>
<td></td>
<td>Higher</td>
<td>56</td>
<td>90</td>
<td>7.25</td>
<td>(3.06 – 17.20)</td>
<td>6.12</td>
<td>(2.44 – 15.35)</td>
</tr>
<tr>
<td>C/A (I) (ratio)</td>
<td>Lower</td>
<td>15</td>
<td>71</td>
<td>1.00</td>
<td></td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Medium</td>
<td>26</td>
<td>80</td>
<td>2.41</td>
<td>(0.95 – 6.10)</td>
<td>2.03</td>
<td>(0.79 – 5.23)</td>
</tr>
<tr>
<td></td>
<td>Higher</td>
<td>41</td>
<td>88</td>
<td>3.78</td>
<td>(1.47 – 9.68)</td>
<td>2.54</td>
<td>(0.98 – 6.60)</td>
</tr>
<tr>
<td>C/A (II) (ratio)</td>
<td>Lower</td>
<td>25</td>
<td>86</td>
<td>1.00</td>
<td></td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Medium</td>
<td>22</td>
<td>88</td>
<td>0.98</td>
<td>(0.48 – 2.00)</td>
<td>1.13</td>
<td>(0.49 – 2.60)</td>
</tr>
<tr>
<td></td>
<td>Higher</td>
<td>42</td>
<td>86</td>
<td>2.44</td>
<td>(1.11 – 5.37)</td>
<td>2.31</td>
<td>(0.93 – 5.75)</td>
</tr>
</tbody>
</table>

* The tertiles are based on the distributions among controls.
† Adjusted for two other antibodies.
Table 4. Relative odds (OR’s), with 95% confidence intervals (CI’s), of rheumatoid factor positive rheumatoid arthritis between tertiles* of citrulline/arginine containing A. C/A (I) and B. C/A (II) C-telopeptide antibody ratios in tertiles* of anti-CCP antibodies.

<table>
<thead>
<tr>
<th></th>
<th>Anti-CCP</th>
<th>C/A (I) ratio</th>
<th></th>
<th>Anti-CCP</th>
<th>C/A (II) ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Lower</td>
<td>Medium</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lower</td>
<td>n †</td>
<td>OR (95% CI)</td>
<td>n †</td>
<td>OR (95% CI)</td>
<td>n †</td>
</tr>
<tr>
<td>Lower</td>
<td>4/25</td>
<td>1.00</td>
<td>3/25</td>
<td>1.05 (0.18 – 6.01)</td>
<td>8/21</td>
</tr>
<tr>
<td>Medium</td>
<td>3/26</td>
<td>0.79 (0.12 – 5.25)</td>
<td>8/17</td>
<td>5.58 (1.09 – 28.62)</td>
<td>15/37</td>
</tr>
<tr>
<td>Higher</td>
<td>8/27</td>
<td>2.26 (0.41 – 12.59)</td>
<td>6/33</td>
<td>2.58 (0.47 – 14.10)</td>
<td>27/28</td>
</tr>
</tbody>
</table>

p-value for interaction = 0.28

<table>
<thead>
<tr>
<th></th>
<th>Anti-CCP</th>
<th>C/A (II) ratio</th>
<th></th>
<th>Anti-CCP</th>
<th>C/A (II) ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Lower</td>
<td>Medium</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lower</td>
<td>n †</td>
<td>OR (95% CI)</td>
<td>n †</td>
<td>OR (95% CI)</td>
<td>n †</td>
</tr>
<tr>
<td>Lower</td>
<td>5/27</td>
<td>1.00</td>
<td>8/25</td>
<td>2.93 (0.68 – 12.67)</td>
<td>12/34</td>
</tr>
<tr>
<td>Medium</td>
<td>1/22</td>
<td>0.31 (0.03 – 3.26)</td>
<td>6/31</td>
<td>1.99 (0.39 – 10.08)</td>
<td>15/35</td>
</tr>
<tr>
<td>Higher</td>
<td>9/34</td>
<td>1.76 (0.36 – 8.69)</td>
<td>4/31</td>
<td>1.10 (0.18 – 6.69)</td>
<td>29/21</td>
</tr>
</tbody>
</table>

p-value for interaction = 0.02

* The tertiles are based on the distributions among controls.
† n = number of cases/controls
DISCUSSION
In the present study anti-CCPs levels already within the reference range (< 25 U/ml) predicted the development of RF-positive RA and simultaneous occurrence of autoantibodies to citrullinated telopeptide of type II collagen showed a synergistic effect. Citrulline to arginine telopeptide ratios and even the single assays for antibodies to citrulline containing telopeptides also predicted RF-positive RA significantly, but anti-CCPs confounded their effects. The study material consisted of preillness serum samples from subjects who were free from RA at the baseline examination.

The significant synergism we found between anti-CCPs and type II citrullinated telopeptide was unexpected. The previous anti-CCP1 was based on filaggrin-derived peptide, but the second-generation anti-CCP2s is totally new set of several antigens although both have surprisingly a similar brand name. The sequences of anti-CCP2 peptides have not yet been published (32). Also the number of different peptides in the CCP2 assay is secret, but anyhow it is sure that they are not present in human bodies (33). However, we have previously demonstrated that antibodies binding telopeptides are not recognising the anti-CCP2s antigens (15). Although there was no linear correlation between anti-CCPs results and type I and II telopeptide antibodies, both are increased in RA. Indeed, this synergism is a new finding that we could not predict beforehand. The difference between antibodies against citrullinated type I and II collagens might then be due to smaller number of samples of the former peptide which decreases the statistical power.

In our previous work we wanted to characterise sequential determinants in citrullinated fibrillar collagens and selected two pairs (arginine/citrulline) of linear synthetic peptides related to the triple-helical region and another two related to the carboxyterminal telopeptide of human type II collagen (15). About half of the RA sera tested bound some of the peptides, usually showing more binding to the arginine- than the citrulline-containing variant. For a peptide corresponding to the most terminal part of the telopeptide, however, significant binding to the citrullinated form was seen. Similar findings were made with peptides related to the telopeptides domains of the two different α-chains of type I collagen (15). Based on these results, we did not test other peptides than those located at the non-helical end of type I and II collagen molecules. One of the reasons was that there was not enough serum to test several collagenous peptides.

The process of the induction of autoantibodies against citrullinated proteins is not known. Synovial tissues contain several proteins that can be citrullinated (4). Possible candidates to provoke immune responses include citrullinated α and β chains of fibrin in synovial tissue (34) and the Sa antigen, identified as citrullinated vimentin (35). The presence of autoantibodies to citrullinated proteins, has recently been reported to predict the development of RA in of apparently healthy people (20, 21) and as recently reviewed (23) several studies have shown that anti-CCPs antibodies predict the progression of undifferentiated arthritis to RA. The results of the current study are concordant with our previous observation that also antifilaggrin antibodies within the normal range predict the development of RF-positive RA in a linear fashion (16). It may be noted that the RF and anti-citrullinated protein/peptide antibodies both have different associations with certain features of RA like extra-articular manifestations and that anti-TNF therapy appears to affect differently RF and anti-CCPs antibody levels (36).

Autoantibody positivity prior to the beginning of the clinical disease is a common finding in other autoimmune diseases, as systemic lupus erythematosus (37) and insulin-dependent diabetes mellitus (38). Thus, at least in theory it is today already possible to identify subjects at risk and even consider immunologic interventions to
prevent the onset of e.g. diabetes, although considerable difficulties are to be expected in such approaches (39). It is likely that there are in RA several antibodies against citrullinated proteins found either in intracellular or extracellular space, many of which could be innocent in nature. On the other hand, autoantibodies against citrullinated collagens could be harmful (40). They might participate in the destruction of both cartilage and periarticular bone. It is interesting that both type I and type II collagens have a similar sequence at the carboxytelopeptides and in RA both cartilage and periarticular bones are affected. Citrullination processes in joint collagens in RA is a demanding task to study, since the amount of citrulline could be very small and possibly subject to attack of autoantibodies against citrullinated collagens. In this respect new methods to analyze protein citrullination (41) might be helpful. At this time, however, it has been convincingly shown that both type I and type II collagen can be citrullinated in vitro (13-15) and that autoantibodies to these citrullinated antigens frequently occur in the sera of RA patients. Importantly, the binding of the autoantibodies to collagens could be specifically inhibited by citrulline-containing soluble peptides (42).

In conclusion, the present study shows that even low levels of anti-CCPs 2 antibodies (within the reference range, i.e., below 25 U/ml) predict in a dose-dependent manner the development of seropositive RA. Men had significantly higher levels of anti-CCPs and a higher C/A (I) ratio than had women. Simultaneous occurrence of antibodies to citrullinated telopeptide of type II collagen, expressed as C/A (II) ratio, exerts a synergistic effect.

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REFERENCES


Autoantibodies binding to citrullinated telopeptide of type II collagen and to cyclic citrullinated peptides predict synergistically the development of seropositive rheumatoid arthritis

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