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JAK selectivity and the implications for clinical inhibition of pharmacodynamic cytokine signalling by filgotinib, upadacitinib, tofacitinib and baricitinib

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ABSTRACT

Objective Janus kinase inhibitors (JAKinibs) are efficacious in rheumatoid arthritis (RA) with variable reported rates of adverse events, potentially related to differential JAK family member selectivity. Filgotinib was compared with baricitinib, tofacitinib and upadacitinib to elucidate the pharmacological basis underlying its clinical efficacy and safety.

Methods In vitro JAKinib inhibition of signal transducer and activator of transcription phosphorylation (pSTAT) was measured by flow cytometry in peripheral blood mononuclear cells and whole blood from healthy donors and patients with RA following cytokine stimulation of distinct JAK/STAT pathways. The average daily pSTAT and time above 50% inhibition were calculated at clinical plasma drug exposures in immune cells. The translation of these measures was evaluated in ex vivo-stimulated assays in phase 1 healthy volunteers.

Results JAKinib potencies depended on cytokine stimulus, pSTAT readout and cell type. JAK1-dependent pathways (interferon (IFN) α /pSTAT5, interleukin (IL)-6/pSTAT1) were among the most potentially inhibited by all JAKinibs in healthy and RA blood, with filgotinib exhibiting the greatest selectivity for JAK1 pathways. Filgotinib (200 mg once daily) had calculated average daily target inhibition for IFN α /pSTAT5 and IL-6/pSTAT1 that was equivalent to tofacitinib (5 mg two times per day), upadacitinib (15 mg once daily) and baricitinib (4 mg once daily), with the least average daily inhibition for the JAK2-dependent and JAK3-dependent pathways including IL-2, IL-15, IL-4 (JAK1/JAK3), IFN γ (JAK1/JAK2), granulocyte colony stimulating factor, IL-12, IL-23 (JAK2/tyrosine kinase 2) and granulocyte-macrophage colony-stimulating factor (JAK2/JAK2). Ex vivo pharmacodynamic data from phase 1 healthy volunteers clinically confirmed JAK1 selectivity of filgotinib.

Conclusion Filgotinib inhibited JAK1-mediated signalling similarly to other JAKinibs, but with less inhibition of JAK2-dependent and JAK3-dependent pathways, providing a mechanistic rationale for its apparently differentiated efficacy/safety profile.

INTRODUCTION

The Janus kinase (JAK) and signal transducers and activators of transcription (STAT) proteins constitute the JAK-STAT pathways, which are essential to immune regulation. Signalling through the pathways is initiated when a cytokine binds its cell-surface receptor, activating receptor-associated JAKs and phosphorylating STAT proteins. The

Key messages

What is already known about this subject?

- Janus kinase (JAK) inhibitors (JAKinibs) have emerged as an important new class of oral therapy for the treatment of rheumatoid arthritis (RA).
- Despite showing similar clinical efficacy, the reported incidence rates of some adverse events of special interest vary among the JAKinibs.
- The relationship between JAK isoform selectivity and inhibition of distinct cytokine responses at clinical plasma exposures of these JAKinibs could provide a mechanistic basis for their relative efficacy and safety profiles.

What does this study add?

- This study is the first to combine in vitro inhibition of cytokine responses in whole blood with clinical pharmacokinetics of filgotinib, baricitinib, tofacitinib and upadacitinib to model daily cytokine-mediated pharmacodynamic profiles in healthy individuals and patients with RA.
- These data demonstrate that, compared with upadacitinib, tofacitinib and baricitinib, filgotinib had similar calculated daily average inhibition of JAK1-dependent pathways activated by interleukin 6 and interferon α , but the least inhibition of JAK2-dependent and JAK3-dependent signalling.
- The observed inhibition of JAKinibs on cytokine signalling was highly nuanced, and it was observed to be dependent on cytokine stimulus, STAT (signal transducers and activators of transcription) substrate and cell type, indicating that the differential cytokine profiles may provide a mechanistic rationale for reported efficacy and safety.

STATs dimerise and migrate to the nucleus to induce and/or maintain immune responses via transcriptional regulation.

The JAK family consists of JAK1, JAK2, JAK3 and tyrosine kinase 2 (TYK2). They bind intracellularly to dimeric cytokine receptor chains in pairs that vary by receptor.^{1 2} Depending on which JAK is activated, the functional effects vary. Specific JAK pairs are implicated in diverse functions that regulate inflammation,³ haematopoiesis^{2 4–8} and

Key messages

How might this impact on clinical practice or future developments?

- This study demonstrates that JAKinibs have unique, differential effects on specific cytokine signalling pathways and supports that preferential JAK1 activity is sufficient to drive RA efficacy, while JAK2 and JAK3 inhibition may increase the incidence of adverse events of special interest.
- These data can be used to build mechanistic correlations between cytokine inhibition and rates of adverse events observed with real-world JAKinib use.

immune homeostasis (figure 1).^{9–14} The JAK-STAT pathways also play essential roles in immune-mediated pathology, including rheumatoid arthritis (RA).^{6 15} Uncontrolled cytokine expression drives chronic inflammation; inadequately treated, this leads to systemic illness, joint destruction and deformity that characterise RA.

JAK inhibitors (JAKinibs) have emerged as an important new class of oral therapy in RA.^{1 6} Baricitinib,¹⁶ tofacitinib^{17 18} and upadacitinib¹⁹ are currently approved in the USA, European Union, Japan and other countries. Filgotinib is a novel JAKinib and has recently been approved in the European Union and Japan.²⁰ Filgotinib forms an active human metabolite, GS-829845/G254445,²¹ that contributes to its pharmacological activity.²² Studies suggest that JAK1 inhibition might be largely responsible for the efficacy of JAKinibs in RA.^{23 24} Biochemically, all these JAKinibs show the greatest potency at inhibiting JAK1, with varying levels of selectivity for other JAK isoforms.^{21 25–29} While these inhibitors have similar efficacy in patients with RA, reported rates of adverse events (AEs) differ, including: increased incidences of herpes zoster (HZ), serious infections,

venous thromboembolism, decreased natural killer (NK) cell numbers, thrombocytopenia and anaemia (online supplemental table 1).^{16 18–20} Genetic evidence has linked JAK2 to erythropoiesis and thrombopoiesis, JAK3 to lymphocyte proliferation and immune homeostasis, and TYK2 to antiviral responses.^{6 30 31} Therefore, differences in JAKinib selectivity for cytokine signalling via distinct JAK pairs may provide a mechanistic rationale for reported differences in safety profiles.

The objective of this study was to calculate the daily clinical pharmacodynamic (PD) inhibition profiles of cytokine signalling for filgotinib, baricitinib, tofacitinib and upadacitinib to compare their inhibition and selectivity. To accomplish this, we used a combination of in vitro cellular cytokine assays in human peripheral blood mononuclear cells (PBMCs) and whole blood (WB), coupling these results with the clinical RA plasma exposures of each JAKinib. Furthermore, we confirmed these results through ex vivo PD data obtained from blood samples from phase 1 healthy volunteers administered filgotinib.

METHODS

Detailed experimental procedures are depicted in figure 2 and described in the online supplemental materials.

RESULTS

JAK isoforms differentially contribute to JAK-STAT pathway activity

JAKinibs showed dose-dependent inhibition of cytokine-stimulated phosphorylated STAT (pSTAT) levels in CD4+ and CD8+ T-cells, monocytes, NK cells, neutrophils and B-cells in human WB (tables 1 and 2, online supplemental figures 1–4) and PBMCs (online supplemental tables 2 and 3). For a given cell type, STAT substrate and cytokine stimulus, potency differences were observed between JAKinibs. The potencies measured in WB were consistently weaker than those assessed in PBMCs (tables 1

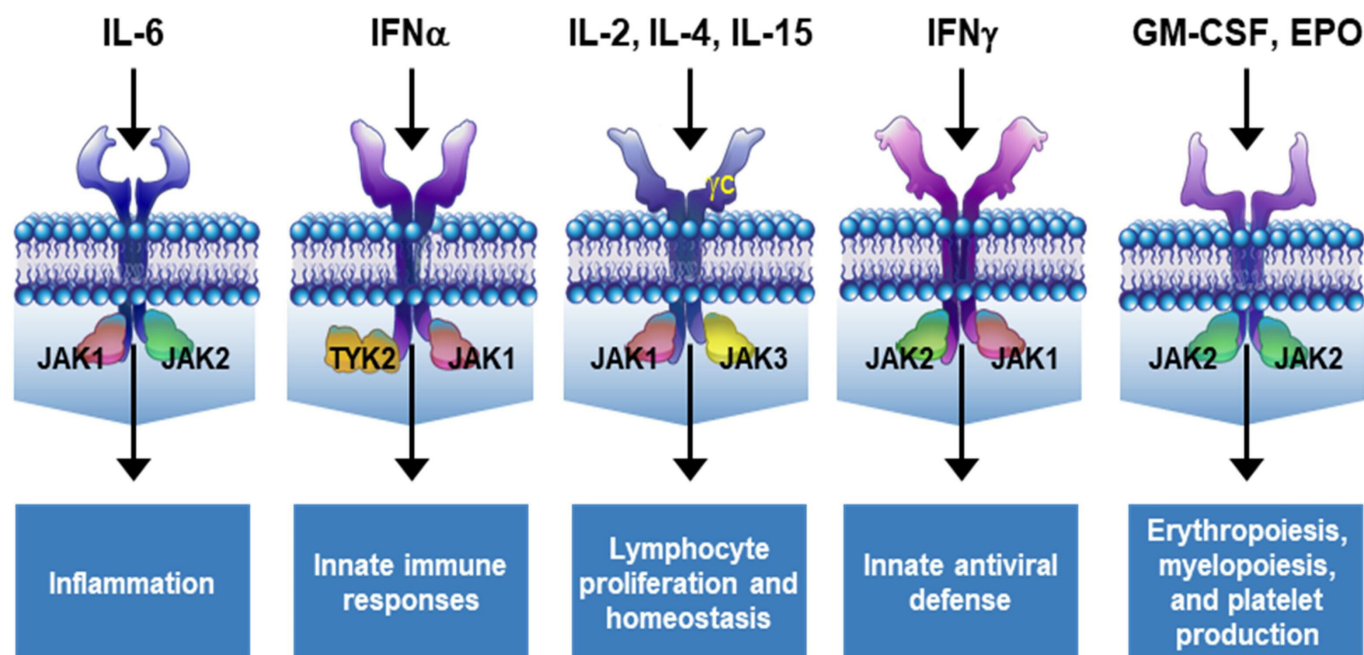
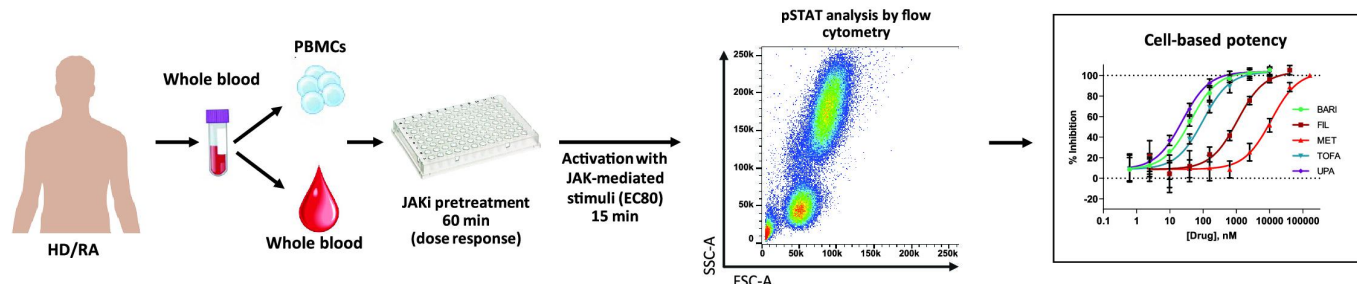


Figure 1 Cytokine receptors are associated with distinct JAK pairing patterns. The JAK isoforms involved in each pathway vary according to the specific cytokine receptor and dictate downstream outcomes. Figure adapted from Winthrop.⁴⁹ EPO, erythropoietin; GM-CSF, granulocyte-macrophage colony-stimulating factor; IFN, interferon; IL, interleukin; JAK, Janus kinase; TYK2, tyrosine kinase 2.

A. Experimental procedure



B. Data analysis procedure

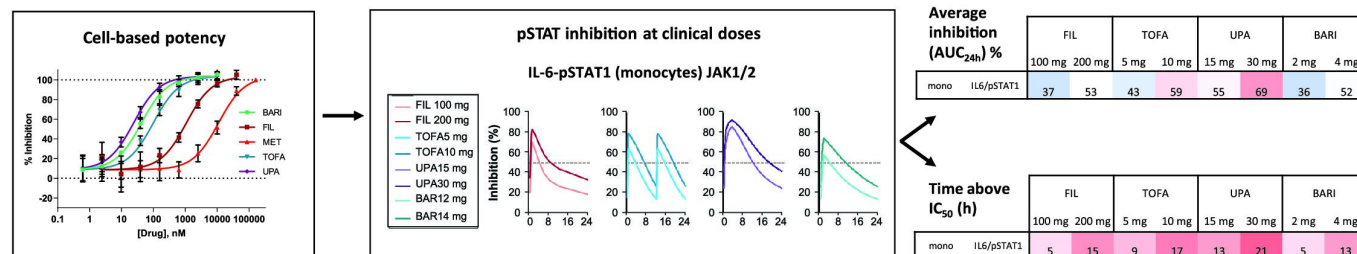


Figure 2 (A) Experimental design and (B) data analysis procedures. Detailed methods are included in online supplemental materials. AUC_{24h}, area under the curve 0–24 hours; BARI, baricitinib; FIL, filgotinib; FSC-A, forward scatter area; HD, healthy donors; IC₅₀, half maximum inhibitory concentration; IL, interleukin; JAK, Janus kinase; JAKi, JAK inhibitor; MET, major metabolite of filgotinib (GS-829845); mono, monocytes; PBMCs, peripheral blood mononuclear cells; pSTAT, phosphorylated signal transducer and activator of transcription; RA, rheumatoid arthritis; SSC-A, side scatter area; TOFA, tofacitinib; UPA, upadacitinib.

and 2, online supplemental tables 2 and 3). This discrepancy was accounted for by compound plasma binding (online supplemental figure 5).

Cell type impacted the measured JAKinib potency to inhibit cytokine signalling. For example, comparing WB CD4+ cells and monocytes, there was an approximately threefold difference for the JAK1/TYK2-dependent interferon (IFN) α -stimulated pSTAT1 for each JAKinib ($p < 0.001$), whereas for

B and NK cells, the potencies were comparable (tables 1 and 2, online supplemental table 4). For each cytokine evaluated, cell type affected JAKinib potencies with varying magnitudes of significance.

JAKinib potencies were dependent on the STAT substrate phosphorylated in response to a given stimulus. In healthy donors, inhibition was consistently 7-fold to 11-fold greater for JAK1/JAK2-driven interleukin (IL)-6/pSTAT1 than for

Table 1 JAKinib IC₅₀ values in CD4+ T-cells, monocytes and NK cells from whole blood assays

Stimulation/pSTAT	CD4+ T-cells					Monocytes					NK cells				
	BARI	FIL	MET	TOFA	UPA	BARI	FIL	MET	TOFA	UPA	BARI	FIL	MET	TOFA	UPA
IC ₅₀ , nM															
JAK2/JAK2 or JAK2/TYK2-dependent cytokines															
G-CSF/pSTAT3			NS			81	4977	50 215	292	81			NS		
GM-CSF/pSTAT5			NS			127	9916	102 910	510	74			NS		
IL-12/pSTAT4			NS			NS					269	10 351	221 777	1216	364
JAK1/JAK2/TYK2-dependent cytokines															
IFN α /pSTAT1	50	1096	17 161	98	30	192	4560	91 078	393	83	131	2440	41 161	256	117
IFN α /pSTAT3	39	871	12 644	80	24	40	991	15 793	86	17	30	675	8620	60	25
IFN α /pSTAT5	28	638	9587	49	17	25	613	9518	51	11	34	507	7484	50	26
IFN γ /pSTAT1			NS			74	4138	62 374	228	58			NS		
IL-6/pSTAT1	29	783	5637	63	27	39	1011	10 019	84	22			NS		
IL-6/pSTAT3	274	5435	62 680	644	225	161	3527	49 109	368	100			NS		
JAK1/JAK3-dependent cytokines															
IL-2/pSTAT5	40	988	14 079	40	21			NS			80	2153	23 824	87	56
IL-4/pSTAT6	73	1458	39 420	77	36	53	1337	36 537	116	32	40	869	20 984	45	25
IL-15/pSTAT5	38	967	14 326	39	21			NS			83	2044	27 540	93	58

G-CSF, GM-CSF, IFN α , IFN γ , IL-6, IL-2, IL-4 and IL-15 reported IC₅₀ values are based on the average of duplicates from 7–10 healthy volunteer whole blood.

BARI, baricitinib; FIL, filgotinib; G-CSF, granulocyte colony stimulating factor; GM-CSF, granulocyte-macrophage colony stimulating factor; IC₅₀, half maximum inhibitory concentration; IFN, interferon; IL, interleukin; JAK, Janus kinase; JAKinib, Janus kinase inhibitor; MET, major metabolite of filgotinib (GS-829845); NK, natural killer cell; NS, not sampled; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; TYK2, tyrosine kinase 2; UPA, upadacitinib.

Table 2 JAKinib IC₅₀ values in B-cells, neutrophils and CD8+ T-cells from whole blood assays

	B-cells					Neutrophils					CD8+ T-cells				
	BARI	FIL	MET	TOFA	UPA	BARI	FIL	MET	TOFA	UPA	BARI	FIL	MET	TOFA	UPA
Stimulation/pSTAT	IC ₅₀ , nM														
JAK2/JAK2 or JAK2/TYK2-dependent cytokines															
G-CSF/pSTAT3			NS			404	16 717	158 111	1245	369			NS		
GM-CSF/pSTAT5			NS			66	3436	40 925	143	21			NS		
IL-23/pSTAT3*			NS			NS					210	15 040	138 638	970	368
JAK1/JAK2/TYK2-dependent cytokines															
IFN α /pSTAT1	156	2957	54 052	322	112			NS			80	1809	27 730	163	56
IFN α /pSTAT3	28	588	9777	62	20	47	833	13 438	66	18	35	795	10 617	73	23
IFN α /pSTAT5	23	436	7524	44	16			NS			27	607	8231	46	17
IFN γ /pSTAT1	22	900	12 820	61	21	78	3137	42 649	152	46			NS		
JAK1/JAK3-dependent cytokines															
IL-2/pSTAT5			NS					NS			32	809	11 046	33	21
IL-4/pSTAT6	295	6426	164 309	356	162	71	1200	37 069	106	37	47	1022	25 900	51	26
IL-15/pSTAT5			NS					NS			56	1459	19 958	56	32

G-CSF, GM-CSF, IFN α , IFN γ , IL-2, IL-4 and IL-15 reported IC₅₀ values are based on the average of duplicates from 6–10 healthy volunteer whole blood.

*Memory CD8+ T-cells.

BARI, baricitinib; FIL, filgotinib; G-CSF, granulocyte colony stimulating factor; GM-CSF, granulocyte-macrophage colony-stimulating factor; IC₅₀, half maximum inhibitory concentration; IFN, interferon; IL, interleukin; JAK, Janus kinase; JAKinib, Janus kinase inhibitor; MET, major metabolite of filgotinib (GS-829845); NS, not sampled; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; TYK2, tyrosine kinase 2; UPA, upadacitinib.

IL-6/pSTAT3 ($p < 0.001$), potentially implicating that JAK1 is predominantly mediating phosphorylation of STAT1 and JAK2 is mediating STAT3 (tables 1 and 2, online supplemental table 5). Similarly, JAKinib inhibition of JAK1/TYK2-mediated IFN α -driven pSTAT5 and pSTAT3 was more potent than pSTAT1, again potentially demonstrating the reported reliance on TYK2 for regulating STAT1 phosphorylation.³²

JAKinibs showed differences in inhibition of different cytokines using the same JAK pair. Filgotinib showed an approximately fourfold potency shift for JAK1/JAK2-dependent IL-6/pSTAT1 compared with IFN γ /pSTAT1 in monocytes ($p < 0.001$) (table 1, online supplemental table 6). For other JAKinibs, potency differences were observed, although with less magnitude. These data highlight that cytokine receptors using identical JAK pairs may have differential reliance on a JAK isoform for mediating signalling.

JAKinibs demonstrate distinct cellular selectivity

Selectivity between JAKinibs was analysed by measuring inhibition of cytokine signalling via each JAK pair in WB monocytes or NK cells (table 1). JAKinibs most potently inhibited the JAK1/TYK2-dependent IFN α /pSTAT5. Measured half maximum inhibitory concentration (IC₅₀) values for other cytokine responses were normalised to this value to control for intrinsic JAKinib potency differences (figure 3). All JAKinibs showed selectivity within approximately fivefold on JAK1-dependent pathways including IL-6 (JAK1/JAK2) and IL-15 (JAK1/JAK3). Conversely, JAKinibs showed differential selectivity against JAK2-mediated pathways. Filgotinib, GS-829845, tofacitinib and upadacitinib showed more than fivefold selectivity versus JAK2-dependent granulocyte colony stimulating factor (G-CSF)-driven or granulocyte-macrophage colony-stimulating factor (GM-CSF)-driven signalling, with filgotinib demonstrating the greatest selectivity against JAK2 compared with JAK1 (IFN α). Baricitinib showed lower JAK1 selectivity (≤ 5.1 -fold for JAK1 versus non-JAK1 pathways; figure 3).

JAKinibs show differentiated JAK1-selective pharmacological profiles at clinical plasma exposures

Representative cytokine/pSTAT pathway inhibition profiles were modelled over a 24-hour period at clinically relevant drug

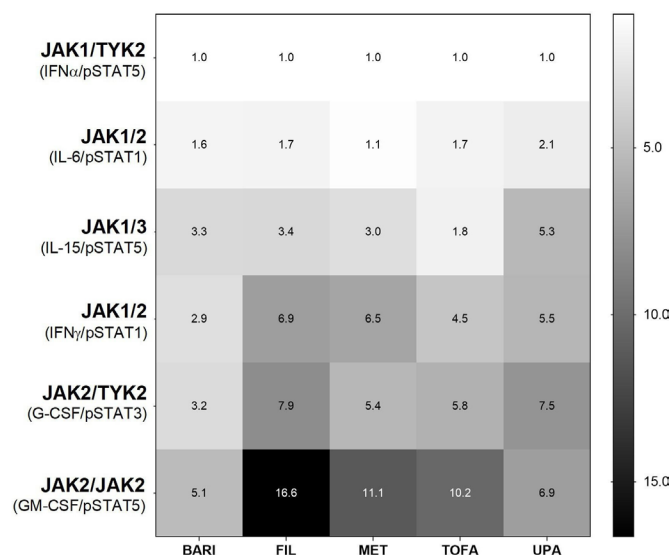


Figure 3 Distinct JAKinib cellular selectivity for JAK heterodimeric cytokine signalling. Mean fold selectivity for each JAK-dimer pair normalised to inhibition of JAK1/TYK2 pathway (IFN α /pSTAT5 in monocytes). Cytokine/pSTAT pairs: JAK1/TYK2 (IFN α /pSTAT5 in monocytes); JAK1/2 (IL-6/pSTAT1 in monocytes); JAK1/3 (IL-15/pSTAT5 in NK cells); JAK1/2 (IFN γ /pSTAT1 in monocytes); JAK2/TYK2 (G-CSF/pSTAT3 in monocytes); and JAK2/2 (GM-CSF/pSTAT5 in monocytes). BARI, baricitinib; FIL, filgotinib; G-CSF, granulocyte-colony stimulating factor; GM-CSF, granulocyte macrophage colony stimulating factor; IFN, interferon; IL, interleukin; JAK, Janus kinase; JAKinib, JAK inhibitor; MET, major metabolite of filgotinib (GS-829845); NK, natural killer; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; TYK2, tyrosine kinase 2; UPA, upadacitinib.

plasma concentrations in patients with RA at low and high JAKinib doses evaluated in the phase 3 clinical programmes (figure 4).^{33–35}

The predicted average inhibition and time above 50% inhibition (IC₅₀ coverage) of the JAK1/JAK2-mediated IL-6/pSTAT1 (figure 4A) and JAK1/TYK2-mediated IFN α /pSTAT5 (figure 4B) are similar among JAKinibs at doses reported to have similar overall American College of Rheumatology efficacy responses (filgotinib 200 mg once daily; baricitinib 4 mg once daily; tofacitinib 5 mg two times per day; and upadacitinib 15 mg once daily) (online supplemental table 1). For IL-6/pSTAT1, all JAKinibs showed similar inhibition at therapeutic doses. For IFN α /pSTAT5, filgotinib had comparable daily inhibition as baricitinib and tofacitinib, and slightly less than upadacitinib. Some differences were noted in the time above 50% inhibition among JAKinibs. Thus, globally, the four JAKinibs demonstrate comparable IC₅₀ coverage and percentage of pSTAT inhibition at therapeutic doses for these 2 JAK1-driven pathways.

On other JAK1/JAK2-dependent signalling pathways (IFN- γ -induced pSTAT1) in representative cell types, filgotinib 200 mg displayed comparable or lower inhibition and time above 50% inhibition compared with baricitinib 4 mg, tofacitinib 5 mg and upadacitinib 15 mg (figure 4C,D). At these same doses, the average daily inhibition of JAK1/JAK3-dependent IL-4/pSTAT6 with filgotinib and baricitinib was comparable, but significantly lower than with tofacitinib and upadacitinib ($p < 0.001$). Time above 50% inhibition was ≥ 3 times longer for tofacitinib and upadacitinib compared with filgotinib 200 mg ($p < 0.001$) (figure 4D). Similarly, the average daily inhibition and time above 50% inhibition of IFN γ /pSTAT1 were lowest for filgotinib and tofacitinib, compared with baricitinib and upadacitinib ($p < 0.001$) (figure 4C).

For JAK1-independent pathways, filgotinib 200 mg showed similar average daily inhibition of JAK2/TYK2-dependent G-CSF/pSTAT3 signalling as tofacitinib, but lower than baricitinib or upadacitinib (figure 4E). On the JAK2/JAK2-dependent GM-CSF/pSTAT5 pathway, upadacitinib and baricitinib showed ≥ 3 -fold greater inhibition than filgotinib 200 mg ($p < 0.001$), with only upadacitinib achieving $> 50\%$ inhibition during the dose interval ($p < 0.001$) (figure 4F). At clinical doses of upadacitinib, there were greater inhibition and time above 50% inhibition for JAK2-dependent and JAK3-dependent pathways signalling via G-CSF, GM-CSF, IFN γ and IL-4 compared with filgotinib 200 mg ($p < 0.001$) (figure 4C–F).

Taken together, these data demonstrate that filgotinib preferentially inhibits JAK1 pathways at therapeutic doses, with upadacitinib showing the least selectivity at clinical doses. As reported, baricitinib displays JAK1/JAK2 selectivity²⁶ and tofacitinib mainly inhibits JAK1 and JAK3.

In vitro study predictability and translation to clinical PD effects

The in vitro predicted inhibition of IL-6/pSTAT1 and GM-CSF/pSTAT5 in healthy donor blood was confirmed in ex vivo blood measurements from healthy volunteers orally dosed with filgotinib 200 mg. Average inhibitions of IL-6/pSTAT1 and GM-CSF/pSTAT5 with filgotinib were 78% and 36%, respectively (figure 5A). Subtracting the placebo response, there was a good concordance between the measured IL-6 and GM-CSF daily inhibition (68% and 15%, respectively) to the in vitro predicted inhibitions (64% and 6%, respectively; figure 5A), directly demonstrating the clinical relevance of the WB cellular modelling calculations.

To determine if inhibition of cytokine responses in blood of healthy individuals was an appropriate surrogate for RA responses, we directly compared inhibition of cytokine pathways in a small number of blood samples of healthy and RA donors. Using these measured values, the predicted daily cytokine inhibition profile for IFN α /pSTAT5 in CD4+ cells by JAKinibs was comparable in healthy and RA blood (figure 5B). Across pathways, generally comparable average daily inhibitions were predicted for JAKinibs in healthy and RA blood (figure 5C), suggesting that measurements in healthy blood could be translated to predictive inhibitions in patients with RA.

Pharmacological profiles of JAKinibs across cell populations and cytokine stimuli at clinical plasma exposures

Clinical PD inhibition curves of cytokine signalling mediated by distinct JAK pairs were modelled (online supplemental figures 6–9). Average inhibition and time above 50% inhibition are summarised at clinical doses (online supplemental table 7).

For JAK1/TYK2, IFN α /pSTAT5 signalling showed average inhibitions $> 50\%$ across all cell populations tested (figure 6A,B). Comparable levels of average daily inhibition were generally observed across cell populations, with upadacitinib showing greater inhibition in monocytes and CD4+ cells versus filgotinib ($p < 0.05$). The magnitude of inhibition was lower for IFN α -driven pSTAT3 and pSTAT1, but all JAKinibs showed similar potency shifts. The weaker inhibition of IFN α /pSTAT1 ($< 50\%$) by all JAKinibs potentially indicates more TYK2 dependence for STAT1 phosphorylation.³²

For JAK1/JAK2, IL-6/pSTAT1 signalling was inhibited by all JAKinibs, whereas there was half the inhibition of IL-6/pSTAT3 (figure 6A,C). Differences in average daily inhibition by JAKinibs between cell types were observed, but overall filgotinib showed comparable or greater inhibition of IL-6/pSTAT1, while tofacitinib showed the least ($p < 0.001$). In contrast, for the IFN γ /pSTAT1 pathway (also mediated by JAK1/JAK2), upadacitinib and baricitinib showed greater inhibition of this pathway compared with filgotinib ($p < 0.001$). There was a cell type-dependent difference in the magnitude of inhibition, with the greatest daily inhibition observed in B-cells and roughly half as much in neutrophils and monocytes, indicating a potentially greater reliance on JAK2 relative to JAK1 in mediating IFN γ inhibition in specific cells.

For JAK1/JAK3, filgotinib consistently showed the least inhibition of common γ -chain cytokines (IL-2, IL-15 and IL-4) (figure 6A,D). Greater daily inhibition of IL-2-mediated and IL-15-mediated pSTAT5 followed tofacitinib $>$ upadacitinib $>$ baricitinib $>$ filgotinib. There were significant differences between JAKinib daily inhibition that were dependent on cell type, but tofacitinib and upadacitinib reproducibly showed greater inhibition in CD4+ and CD8+ cells than filgotinib ($p < 0.001$). Tofacitinib, upadacitinib and baricitinib showed greater inhibition of IL-4/pSTAT5 than filgotinib in all cell types evaluated ($p < 0.05$).

For JAK2/TYK2, the JAKinibs had a reduced effect on JAK2-mediated signalling, with $< 40\%$ average inhibition of G-CSF/pSTAT3 in monocytes and neutrophils and $< 20\%$ average inhibition of IL-12/pSTAT4 in NK cells and IL-23/pSTAT3 in CD8+ memory T-cells, compared with the JAK1-mediated pathways (figure 6A,E). Upadacitinib and baricitinib showed significantly greater inhibition in monocytes compared with filgotinib. In the remaining cell types, inhibition by JAKinibs was approximately 25%–50% of that in monocytes, and a trend for lower inhibition by filgotinib compared with other JAKinibs was observed.

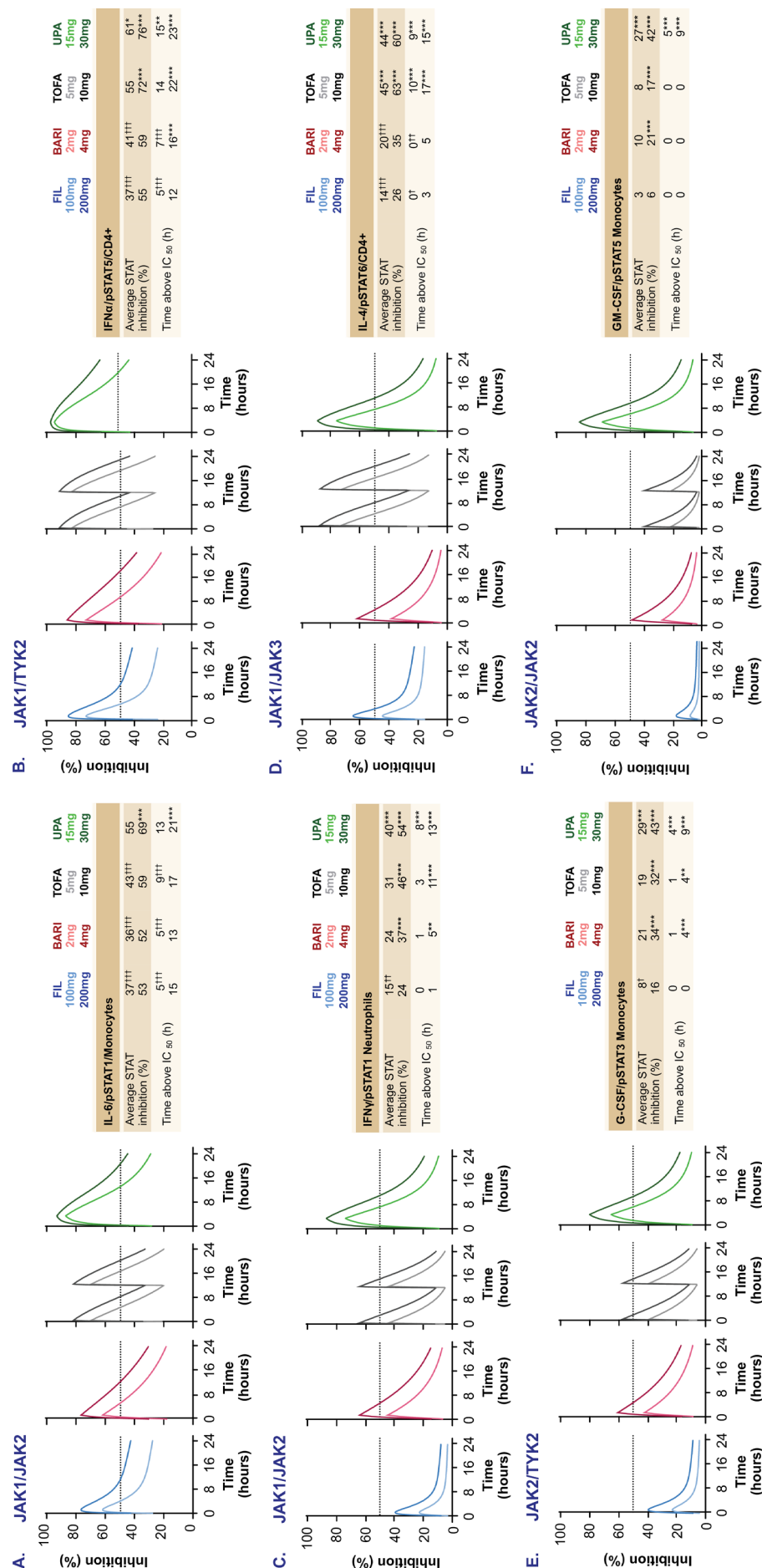


Figure 4 Predicted cytokine inhibition profiles of JAKinibs at clinical doses. Pharmacodynamic pSTAT inhibition for JAKinibs at rheumatoid arthritis clinical doses over a 24-hour dose interval at steady state for selected cytokine stimulations based on in vitro blood measurements (n=7–10). (A) JAK1/JAK2 (IL-6/pSTAT1 in monocytes); (B) JAK1/TYK2 (IFN α /pSTAT5 in CD4+ T-cells); (C) JAK1/JAK2 (IFN γ /pSTAT1 in neutrophils); (D) JAK1/JAK3 (IL-4/pSTAT6 in CD4+ T-cells); (E) JAK2/TYK2 (G-CSF/pSTAT3 in monocytes); (F) JAK2/JAK2 (GM-CSF/pSTAT5 in monocytes). Dashed lines show 50% target inhibition. The average daily STAT inhibition and the number of hours per day JAKinib concentrations above 50% inhibition are indicated. FIL values include contribution from the major metabolite of FIL. *P<0.05, **P<0.01, ***P<0.001 higher vs FIL (200mg); †P<0.05, ††P<0.01, †††P<0.001 lower vs FIL (200mg). BARI, baricitinib; FIL, filgotinib; G-CSF, granulocyte-colony stimulating factor; IFN, interferon; IL, interleukin; JAK, Janus kinase; JAKinib, JAK inhibitor; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; TYK2, tyrosine kinase 2; UPA, upadacitinib.

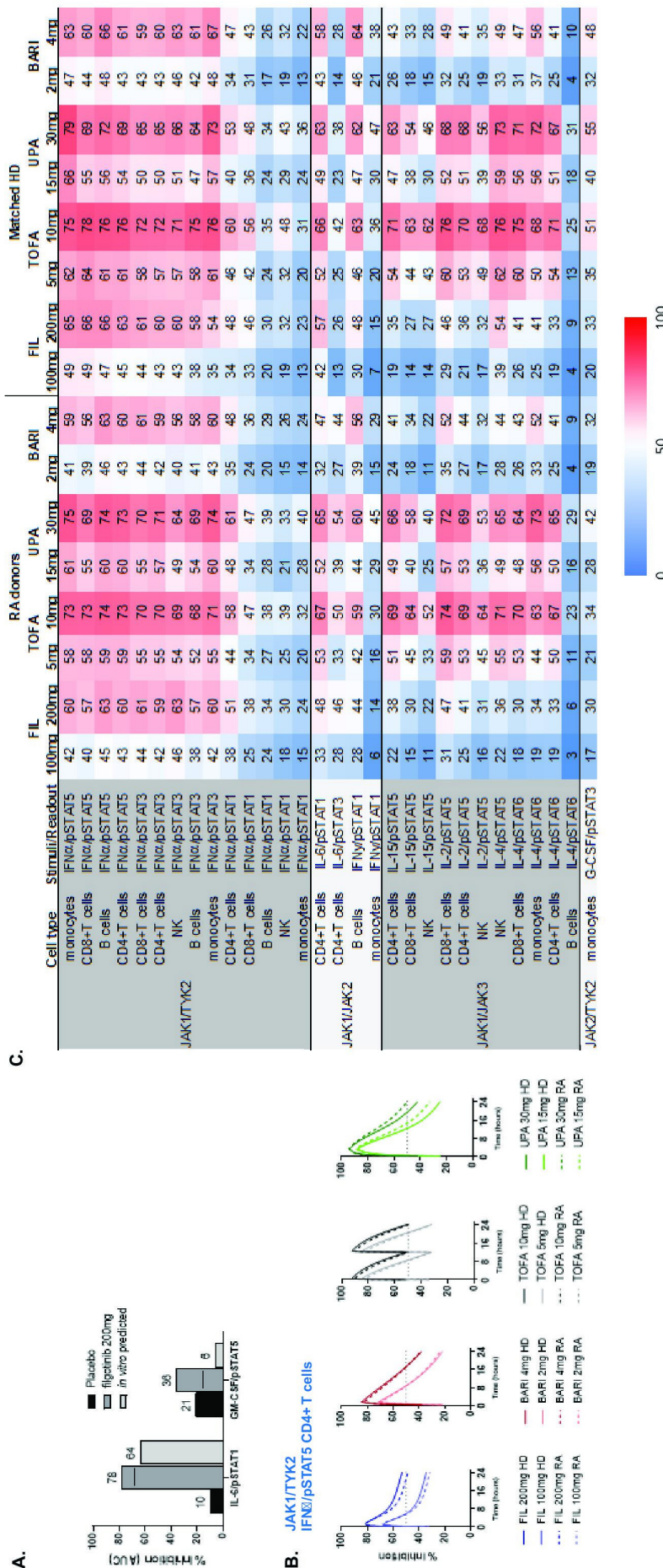
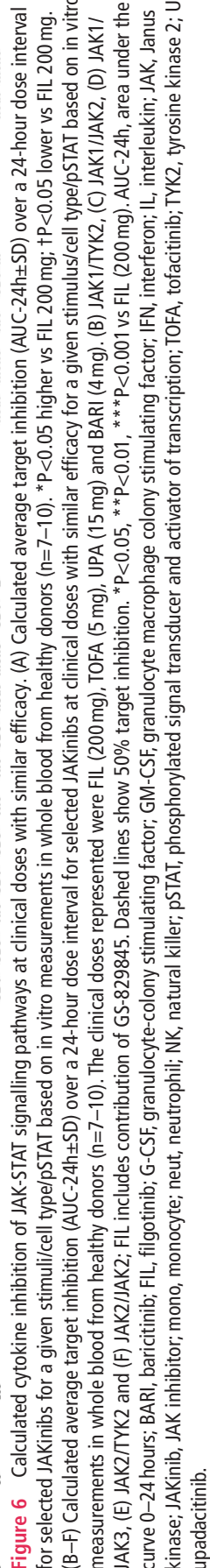


Figure 5 Cytokine-induced pSTAT inhibition in the samples of patients with RA and in ex vivo stimulated blood from phase 1 studies of healthy volunteers. (A) Measured average inhibition of ex vivo stimulated pSTAT1 (IL-6/CD4+ T-cells) and pSTAT5 (GM-CSF/monocytes) over a 24-hour period in healthy volunteers receiving FIL (200 mg once daily) or placebo or from in vitro calculated values. The black bar indicates placebo-adjusted inhibition. FIL values include contribution of GS-829845. (B) Calculated pSTAT5 inhibition for JAKinibs at RA clinical doses over a 24-hour dose interval at steady state for IFN α /pSTAT5 in CD4+ T-cells in HDs (n=2) and patients with RA (n=3) based on in vitro measurements. (C) Heatmap of average daily percent STAT inhibition (area under the curve (AUC)—24 hours) by JAKinibs at clinical doses in patients with RA and matched HDs (based on in vitro whole blood measurements); GM-CSF; granulocyte macrophage colony stimulating factor; HDs, healthy donors; IFN, interferon; JAK, Janus kinase; JAKinib, JAK inhibitor; NK, natural killer; pSTAT, phosphorylated signal transducer and activator of transcription; RA, rheumatoid arthritis; SD, standard deviation; TOFA, tofacitinib; TYK2, tyrosine kinase 2; UPA, upadacitinib.



For JAK2/JAK2, inhibition of the GM-CSF/pSTAT5 pathway was significantly greater with tofacitinib, upadacitinib and baricitinib in neutrophils ($p < 0.01$) and for upadacitinib and baricitinib in monocytes ($p < 0.001$) compared with filgotinib (figure 6F). These data are consistent with the observation that upadacitinib and baricitinib inhibit JAK2 signalling at clinically relevant exposures, as seen for IFN γ /pSTAT1 (figure 6A,C) and G-CSF/pSTAT3 (figure 6A,E).

DISCUSSION

JAKinibs have similar efficacy in RA with distinct safety profiles, potentially reflecting differential JAK isoform selectivity. Enzymatically, JAKinibs have the greatest potency on JAK1, an isoform predominantly involved in inflammatory and innate immune responses.^{21 22 26–29} Given that cytokine signalling involves JAK pairs, consideration solely of JAK1 enzymatic inhibition is insufficient to capture the differential effects of heterogeneous cytokine activation. Inhibition of JAK2 and JAK3 may contribute to AEs, given their involvement in regulating immune cell proliferation and homeostasis. In this study, we demonstrated that cytokine signals were differentially inhibited by filgotinib, tofacitinib, upadacitinib and baricitinib in human PBMCs and WB that translated to differential PD inhibition clinically, revealing unique JAKinib pharmacological selectivity profiles. Our key finding is that filgotinib inhibits JAK1-mediated IFN α and IL-6 similar to other JAKinibs at doses demonstrating similar efficacy, but exhibits reduced inhibition of JAK2-dependent and JAK3-dependent pathways, potentially explaining the pharmacological basis for the reported differentiated safety profile.

In cellular assays, JAKinibs showed the greatest potency at inhibiting JAK1-dependent IFN α and IL-6 pathways and are calculated to have comparable average daily inhibition clinically, highlighting a shared JAK1 PD profile. These results are consistent with previously reported data for JAKinibs in PBMCs and blood.^{25 27} To our knowledge, this is the first comparison of JAKinib cytokine signalling inhibition of blood from healthy subjects and patients with RA, revealing an overall similar potency and predicted inhibition clinically across a panel of cytokines. Interestingly, IL-6/pSTAT3 inhibition in healthy donors was roughly half that of IL-6/pSTAT1, but in RA donors the predicted daily inhibition was comparable, indicating that some JAKinib responses in RA may not be adequately characterised using healthy donors as a surrogate. Additional studies would be required to verify these initial observations. As IL-6 and IFN α are well-established drivers of autoimmunity,^{36–38} and IL-6 is a clinically validated target in RA,³⁹ the observation that all JAKinibs strongly suppress IL-6 and IFN α signalling with comparable potency implicates these pathways as primary drivers of efficacy in RA.^{25 27}

In contrast to the comparable IFN α and IL-6 inhibition, JAKinibs showed significant differences in other JAK1-dependent pathways, reflecting the interplay between cell type, JAK pairing and STAT substrate downstream of different cytokine stimuli. Nuances in cytokine inhibitory profiles for JAKinibs emerged. For example, filgotinib, baricitinib and upadacitinib showed more than threefold weaker inhibition of JAK1/JAK3-dependent IL-15/pSTAT5 pathway relative to IFN α /pSTAT5, whereas tofacitinib showed only a 1.8-fold reduction. Filgotinib showed 6.9-fold weaker activity on JAK1/JAK2-dependent IFN γ /pSTAT1 relative to IFN α /pSTAT5. At clinical exposures, these intrinsic potency differences were amplified. Consistently, there were lower average inhibition and less time above 50% inhibition for filgotinib and baricitinib compared with tofacitinib and

upadacitinib on JAK1/JAK3-dependent pathways. On the JAK1/JAK2-dependent IFN γ /pSTAT1 signalling, filgotinib showed lower inhibition than other JAKinibs in all cell types evaluated. This observation contrasts with the comparable inhibition on JAK1/JAK2-dependent IL-6/pSTAT1, indicating a stronger dependence of JAK1 in mediating IL-6/pSTAT1, as previously reported.^{6 40} Conversely, IFN γ /pSTAT1 may be more reliant on JAK2, suggesting that, at clinical exposures, upadacitinib and baricitinib inhibit JAK2 more than filgotinib and tofacitinib. The lower inhibition of IFN γ /pSTAT1 compared with IL-6/pSTAT1 in CD14+ monocytes for JAKinibs is consistent with data reported by Dowty *et al*;²⁵ however, in that study filgotinib showed comparable inhibition with other JAKinibs. This may be related to the use of a single average concentration to determine target coverage, as opposed to an integrated PD area under the curve approach, or use of saturating levels of cytokine stimulus.²⁵

JAK2-dependent cytokine signalling pathways were less inhibited than JAK1-dependent pathways for all JAKinibs, with filgotinib showing the least inhibition, including in neutrophils. The low level of inhibition of G-CSF/pSTAT3 in neutrophils is consistent with that observed by others in granulocytes.²⁵ JAKinibs are associated with serious infections,^{16 18–20} so minimising inhibition in neutrophils may maintain antipathogenic function. These data are consistent with the observation that filgotinib is the least potent compound for inhibition of IFN γ /pSTAT1, which is dependent on JAK2.

The data for JAKinib inhibition of JAK2-dependent and JAK3-dependent cytokine responses corroborate previous findings that JAKinibs selectively inhibit cytokines that signal via JAK1 versus JAK2, and tofacitinib and upadacitinib have greater relative inhibition of JAK3-dependent common γ -chain cytokine receptor pathways compared with baricitinib or filgotinib at clinical doses.^{24 25 27} Furthermore, our data demonstrate that filgotinib has greater selectivity for cytokine signalling via JAK1 versus JAK2 than baricitinib and upadacitinib. The relatively low clinical inhibition of JAK2-dependent GM-CSF, IL-12, IL-23 and G-CSF is consistent with previous findings that filgotinib did not inhibit these pathways.²⁵ Despite these shared findings, there were notable differences in reported results. Dowty *et al*²⁵ claim ‘limited differentiation’ of JAKinib selectivity; however, it was not statistically powered for comparison, nor did they provide a clinical PD profile of cytokine inhibition. Their approach used an average clinical JAKinib concentration at similarly efficacious therapeutic doses.²⁵ As inhibitory activity is not linear over a concentration range, methodology using a single average concentration may minimise PD differences, potentially obscuring clinical safety and efficacy implications. Currently, it is unknown what level of target inhibition or time above 50% inhibition could have clinical impact on safety, and subtle differences may be meaningful.

Our data indicate that JAKinibs have clinically differentiated PD profiles that mechanistically may underlie incidence rates of presumed class-effect AEs.^{41–48} Selectivity for JAK1 can drive RA efficacy, whereas increasing inhibition of JAK2-dependent and JAK3-dependent pathways may elicit safety liabilities. This is predicted from genetic deletion and clinical data, showing that JAK2 is essential for erythropoiesis, myelopoiesis and platelet production, and JAK3 is critical for lymphocyte proliferation and homeostasis.⁴⁹ Compared with other JAKinibs, filgotinib had less inhibition of JAK1-mediated IFN γ (JAK1/JAK2) and IL-2, IL-15 and IL-4 (JAK1/JAK3), JAK2-mediated G-CSF, IL-12 and IL-23 (JAK2/TYK2), and GM-CSF (JAK2/JAK2). The reduced inhibition of JAK2-dependent and JAK3-dependent

cytokine signalling by filgotinib may underlie its lower reported impact on homeostatic immune functions that control NK cells, platelet numbers, anaemia, lymphocyte numbers, infection and HZ (online supplemental table 1).^{49–51}

This study has limitations. We established that JAKinib effects are context-dependent and influenced by parameters including cytokine stimulus, STAT substrate and cell type. The interdependence of these parameters needs to be considered when extrapolating these data. We evaluated cytokines that signal via distinct JAK pairs and generalised the results to other cytokines using the same JAK pairs. Our results, however, indicate that the inhibitory effect of each JAKinib is dependent on the specific cytokine stimulus, STAT substrate and cell type, so the association of pathway inhibition with clinical impact would require cell-specific cytokine evaluation. To validate whether the findings in healthy controls would predict cellular effects at clinical exposures, we measured ex vivo effects of IL-6/pSTAT1 and GM-CSF/pSTAT5 in healthy volunteers dosed with filgotinib and showed good agreement with the in vitro-derived extent of inhibition. Although we studied JAKinib cytokine inhibition in blood from patients with RA, the number of donors and the breadth of the cytokine panel were limited. The validity of extrapolating these findings to a broad spectrum of cytokine responses in patients with RA requires further evaluation. While this study provides a potential mechanistic basis for the differential JAKinib rates of AEs from meta-analyses, confirmation would involve head-to-head clinical evaluations.

In conclusion, we demonstrated that, based on in vitro cellular assays and clinical pharmacokinetics, filgotinib is predicted to have a differentiated cytokine PD profile in the clinical setting compared with other JAKinibs. JAK1-dependent IL-6 and IFN α pathways were comparably inhibited by JAKinib doses that have similar efficacy in RA, but filgotinib demonstrated reduced JAK2 and JAK3 activity, providing a potential mechanistic basis for the reported differences in its safety profiles.

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Competing interests PGT, BM, FC, AM and JAD are employees of Gilead Sciences, Inc. RG is an employee of Galapagos SASU.

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Ethics approval The study was conducted in accordance with the ethical principles of the Declaration of Helsinki and Good Clinical Practice guidelines, and was approved by the Independent Ethics Committee or Institutional Review Board. The study was performed in compliance with the ethical principles of good clinical practice and according to the ICH Harmonised Tripartite Guideline. Patients provided written informed consent to participate in the study and had the right to withdraw at any time.

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Data availability statement All data relevant to the study are included in the article or uploaded as supplementary information. Anonymised individual patient data will be available upon request to qualified external researchers 6 months after FDA and European Medicines Agency approval per Gilead's Clinical Trial Disclosure and Data Transparency Policy as posted at <https://www.gilead.com/research/disclosure-and-transparency>.

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Supplemental Materials to:

Traves PG, Murray B, Campigotto F, et al.

JAK Selectivity and the Implications for Clinical Inhibition of Pharmacodynamic Cytokine Signaling by Filgotinib, Upadacitinib, Tofacitinib, and Baricitinib

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Detailed Methods

PBMC and WB Preparation and Experimental Design

Figure 2 illustrates the experimental and data analysis procedures. Whole blood samples from healthy donors (N=10) were collected by venipuncture in sodium heparin and processed within 24 hours. PBMCs were harvested from matched whole blood (200mL) by Ficoll gradient. PBMCs or WB were plated into 96-well flat bottom plates and incubated with each JAKinib. Filgotinib (GS-6034) and its active metabolite (MET, GS-829845) were synthesized at Gilead Sciences, Inc. (Foster City, CA). Baricitinib (GS-623452) was purchased from AstaTech, Inc. (Bristol, PA), tofacitinib (GS-493645) was synthesized by Advanced Chemblocks, Inc. (Burlingame, CA), and upadacitinib (GS-942618) was synthesized by MedChemExpress (Monmouth Junction, NJ). Baricitinib, tofacitinib, and upadacitinib were prepared as 10mM stocks and filgotinib and MET as 40mM or 160mM stocks in dimethyl sulfoxide (DMSO). JAKinibs were applied using 8-point dose-titrations ranging from 0.6–10,000nM (baricitinib, tofacitinib, and upadacitinib) or either 2.4–40,000nM or 9.7–160,000nM (filgotinib and MET) (four-fold dilutions) in duplicate, for 60 min prior to cytokine stimulation for 15 min at 37°C.

Whole blood samples from rheumatoid arthritis donors (n=3; patient details are provided in **Appendix 1**) and matched healthy donors (n=2) were collected by venipuncture in sodium heparin, shipped ON at 4C and processed within 36 hours. RA and HD samples were gender and aged matched. RA blood samples were obtained from BioIVT (Hicksville, NY) for experimental use. The inclusion criteria for RA patients included patients able to provide written informed consent, with confirmed diagnosis of RA, with active moderate to severe RA, who have not taken an investigational drug in the last year and who have not been treated with any JAK inhibitor.

Cytokine EC₈₀ Determination

Cytokine concentrations were optimized in PBMCs and WB (N=3) to achieve 80% maximal pSTAT signals in each cell population. Final concentrations were: PBMCs; GM-CSF (9pg/ml), G-CSF (4ng/mL), IFN-γ (0.2ng/ml), IFN-α (6ng/ml), IL-2 (5ng/ml), IL-4 (0.2ng/ml), IL-6 (4ng/ml), and IL-15 (0.4ng/ml). Blood; GM-CSF (9pg/ml), G-CSF [10-25ng/mL (10ng/ml donors 1 and 2, 25ng/ml donors 4-6)], IFN-γ (8ng/ml), IFN-α (6ng/ml), IL-2 (5ng/ml), IL-4 (0.3ng/ml), IL-6 (10ng/ml), IL-12 (10ng/ml), IL-23 (100ng/ml) and IL-15 (1ng/ml).

Flow Cytometry

Cells were lyse/fixed, permeabilized and frozen at -80°C. STAT phosphorylation (pSTAT1, pSTAT3, pSTAT4, pSTAT5 and pSTAT6) was detected and quantified by multicolor flow cytometry in the phenotypically gated leukocyte subpopulations defined as CD19+ (B cells), CD3+CD4+ (CD4+ T cells), CD3+CD8+ (CD8+ T cells), CD3+CD8+CD45RA- (CD8+ memory T cells), CD3-CD56+ (NK cells), CD15+ (neutrophils), and CD14+ (monocytes) (See **Appendix 2** for reagents and compounds and **Appendix 3** for gating strategies). The mean fluorescent intensity (MFI) for the unstimulated versus stimulated conditions was analyzed and reported when >1.4 and >200 fluorescent units. The raw flow cytometry data are shown in **Appendix 4**.

Curve-Fitting Parameters for Concentration Response Curves

For a given response [cell source (PBMC or WB), stimulation (IL-2, IL-15, IL-4, IL-12, IL-23, IFNα, IFNγ, IL-6, G-CSF, and GM-CSF), cell type (B, CD4+, CD8+, CD8+ memory T cells, NK, monocytes, and neutrophils), and pSTAT (pSTAT1, 3, 4, 5, 6)] the half maximal (IC₅₀) values for each JAKinib was determined by inhibition of MFI. A 4-parameter logistic curve fit was reported as % pSTAT inhibition using GraphPad Prism 8.1.2. The IC₅₀ value from a curve was accepted if the following conditions were met: (1) the R² was >0.7, and (2) the standard error

of the slope was <6 . The arithmetic mean IC_{50} value for each case was estimated in PBMCs ($N=3-5$) and WB ($N=6-10$) and compared among JAKinibs.

Pharmacokinetic (PK) Profiles

Human steady-state population PK curves in RA subjects for baricitinib once-daily dosing at 2 or 4mg,[1] TOFA with twice-daily 5 and 10mg,[2], and UPA with once-daily dosing at 15 and 30mg,[3] were used as previously reported. The PK profile of FIL was simulated based on a 2-compartment model, including a first-order absorption with lag time. For the metabolite, the PK was obtained based on a 1-compartment model, including a zero-order input, followed by first-order absorption model.

Estimation and Comparison of Time Above IC_{50} and Daily Percent Inhibition

The mean IC_{50} values obtained from duplicate dose-response curves and human population PK curves were used to model time above IC_{50} and average daily percent inhibition for each cytokine/STAT pair in individual cell types. In vitro IC_{50} value in PBMCs were adjusted for protein binding by dividing the IC_{50} value for each donor by the experimentally assessed unbound fraction ($N=3$). Measured protein binding were: baricitinib 53%, filgotinib 64%, GS-829845 69%, tofacitinib 67%, and upadacitinib 50% (FDA Center for Drug Evaluation and Research NDA 211675). No adjustments were made to whole blood values. The time above IC_{50} was defined as the total time the PK concentration was above the IC_{50} values. The average daily percent inhibition was obtained by entering the steady state PK concentrations into the concentration response curves, computing the area under the curve (AUC), and dividing it by 24 hours.

Statistical Analysis

Comparisons between filgotinib and the other treatment groups were performed using a two-way mixed effect Analysis of Variance (ANOVA) to model the time above IC_{50} and average daily percent inhibition with fixed effect for compound for each stimulation, cell type, and pSTAT combination and random effect for donor with Dunnett's multiplicity correction for multiple comparison adjustments. Filgotinib 200mg was considered the reference group for the comparisons. The same mixed effect model was used to fit the $\log(IC_{50})$ values and estimate and compare mean IC_{50} values. Reported time above IC_{50} , average daily percent inhibition, and IC_{50} values corresponded to the least squares means of the compound effects from those models. Comparisons were considered significant at a two-sided alpha level of 0.05. Analyses were performed using GraphPad Prism version 8.1.2 and SAS version 9.4. Analysis between protein-adjusted PBMC values and WB were performed using Pearson's correlation. Sample size ($N=10$) was based on feasibility and internal pilot data.

Clinical study: Ex Vivo Stimulation of Phase 1 Samples

The phase 1 study in human volunteers was a randomized, placebo-controlled study: (study identifier: NCT01179581). Subjects received filgotinib 200mg once daily or placebo. Blood was collected from healthy volunteers ($N=10$) into lithium heparin vacutainer tubes, following their informed consent. Blood was equilibrated at $37^{\circ}C$ for 30 min and triggered with either recombinant human IL-6 (10 ng/mL; R&D Systems) or recombinant human GM-CSF (20 pg/mL; PeproTech) at $37^{\circ}C$ for 20 min and treated with prewarmed 1X lysis/fix buffer (BD Biosciences) to lyse RBCs and fix leukocytes. Cells were permeabilized with 100% methanol and incubated with anti-pSTAT1 (#612564) and anti-CD4 (#555349) or anti-pSTAT5 (#612567) and anti-CD33 (#345800) antibodies (BD Biosciences) at $4^{\circ}C$ for 30 min, washed once with PBS 1X, and analyzed on a FACSanto II flow cytometer. Full details have been published previously.[4]

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Appendix 1: Characteristics of RA Donors

	RA Donor 1	RA Donor 2	RA Donor 3
RA Diagnosis Date	1996	2014	2017
Gender	Female	Female	Female
Age	52 years	40 years	56 years
Ethnicity	Caucasian	Caucasian	Caucasian
Other Conditions	Celiac Disease, Lupus, Sjogren's Syndrome, Psoriasis	None	None
Treatment	Hydrocodone	Gabapentin, Hydroxychloroquine (Plaquenil), Meloxicam, Tramadol	Humira Injection
Disease Severity	Severe	Moderate	Moderate
Receiving Treatment for Other Pathologies?	Yes; Synthroid, Tizanidine, Prednisone, Doxycycline, Testosterone Cream	No	No
Pregnant or nursing?	No	No	No
Taking an investigational product in the last year?	No	No	No

Appendix 2: Reagents and Compounds**EC₈₀ Values**

Whole Blood Cytokine EC80s			
Cytokine	Vendor	Catalog No.	EC80
GMCSF	Peprotech	300-03	9pg/ml
GCSF*	Peprotech	300-23	Donor1-2: 10ng/ml Donor 4-6: 25ng/ml
IFN γ	Peprotech	300-02	8ng/ml
IFN α	Sigma	H6041	6ng/ml
IL-2	Gibco	PHC0021	5ng/ml
IL-4	Peprotech	200-04	0.3ng/ml
IL-6	Peprotech	200-06	10ng/ml
IL-15	Peprotech	200-15	1ng/ml
IL-12	R&D systems	219-IL	10ng/ml
IL-23	Proteintech	HZ-1254	100ng/ml
PBMC Cytokine EC80s			
Cytokine	Vendor	Catalog No.	EC80
GMCSF	Peprotech	300-03	9pg/ml
GCSF	Peprotech	300-23	4ng/ml
IFN γ	Peprotech	300-02	0.2ng/ml
IFN α	Sigma	H6041	6ng/ml
IL-2	Gibco	PHC0021	5ng/ml
IL-4	Peprotech	200-04	0.2ng/ml
IL-6	Peprotech	200-06	4ng/ml
IL-15	Peprotech	200-15	0.4ng/ml

*EC₈₀ value of GCSF on whole blood was increased by 2.5x after the first 2 donors were treated.

Staining Panels

Conjugated STAT Antibodies (Post MeOH Perm) All Donors					
Cytokines	Antibody	Vendor	Dilution	Catalog No.	Clone
IFN γ , IL-6, IFN α	STAT1	CST	50	8009S	58D6/P-STAT1 (Tyr701)
G-CSF, IL-6, IFN α , IL-23	STAT3	BD	20	562071	4/P-STAT3 (pY705)
GM-CSF, IL-2, IFN α , IL-15	STAT5	BD	20	562076	47/P-STAT5 (pY694)
IL-4	STAT6	BD	20	562079	18/P-STAT6 (pY641)
IL-12	STAT4	Miltenyi	20	130-114-492	REA855/P-STAT4 (pY693)

Panel 1: Donor 1 WB						
Antigen	Color	Titer	Catalog No.	Clone	Vendor	Pre vs Post
CD8	UV805	50	612889	SK1	BD	Pre-Perm
CD4	UV737	50	612748	SK3	BD	Pre-Perm
CD14	UV395	50	563561	MphiP9	BD	Pre-Perm
CD19	BV785	50	563325	SJ25C1	BD	Pre-Perm
CD56	BV711	100	318336	HCD56	Biolegend	Pre-Perm
CD16	BV711	100	302044	3G8	Biolegend	Pre-Perm
CD3	APC Fire	200	300470	UCHT1	Biolegend	Post-Perm

Panel 2: Donor 1 PBMC, Donor 2 WB+PBMC						
Antigen	Color	Titer	Catalog No.	Clone	Vendor	Pre vs Post
CD8	UV737	50	612754	SK1	BD	Pre-Perm
CD4	UV805	50	612887	SK3	BD	Pre-Perm
CD14	UV737	25	612763	M5E2	BD	Pre-Perm
CD19	BV785	50	563325	SJ25C1	BD	Pre-Perm
CD56	BV711	100	318336	HCD56	Biolegend	Pre-Perm
CD16	BV711	100	302044	3G8	Biolegend	Pre-Perm
CD3	PE	50	300441	UCHT1	Biolegend	Post-Perm
CD15	PECY7	100	323030	W6D3	Biolegend	Post-Perm

Panel 3: Donor 4 WB+PBMC						
Antigen	Color	Titer	Catalog No.	Clone	Vendor	Pre vs Post
CD8	UV737	50	612754	SK1	BD	Pre-Perm
CD4	UV805	50	612887	SK3	BD	Pre-Perm
CD14	UV395	25	563561	MphiP9	BD	Pre-Perm
CD19	BV785	50	563325	SJ25C1	BD	Pre-Perm
CD56	BV711	100	318336	HCD56	Biolegend	Pre-Perm
CD16	BV711	100	302044	3G8	Biolegend	Pre-Perm
CD3	PE	50	300441	UCHT1	Biolegend	Post-Perm
CD15	Percpcy5.5	200	323030	W6D3	Biolegend	Post-Perm

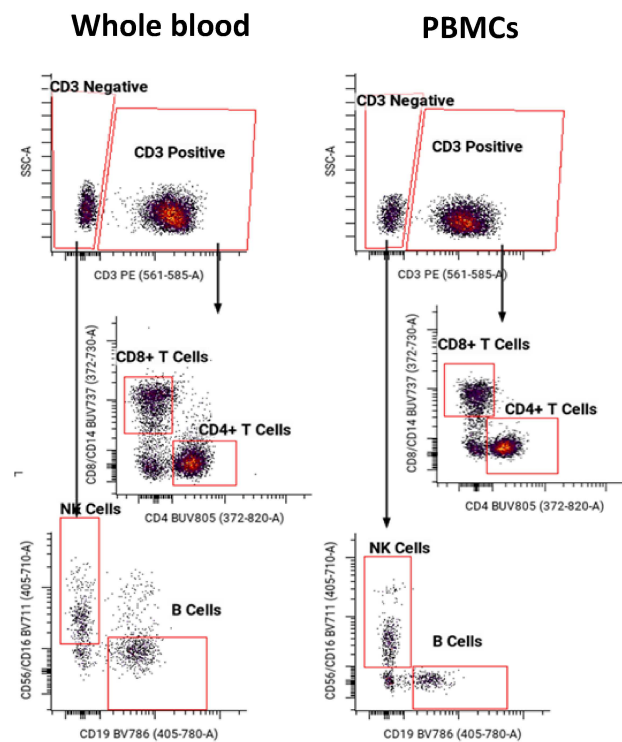
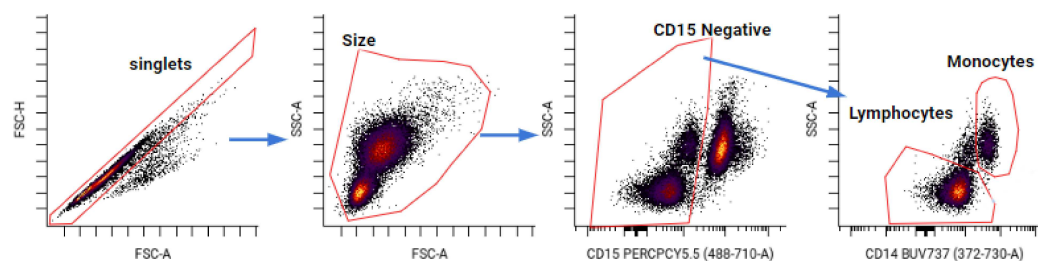
Panel 4: Donor 5 WB+PBMC, Donor 6 WB+PBMC						
Antigen	Color	Titer	Catalog No.	Clone	Vendor	Pre vs Post
CD8	AX700	100	557945	RPA-T8	BD	Pre-Perm
CD4	UV805	50	612887	SK3	BD	Pre-Perm
CD14	UV737	25	612763	M5E2	BD	Pre-Perm
CD19	BV785	50	563325	SJ25C1	BD	Pre-Perm
CD56	BV711	100	318336	HCD56	Biolegend	Pre-Perm
CD16	BV711	100	302044	3G8	Biolegend	Pre-Perm
CD3	BUV395	50	563546	UCHT1	BD	Pre-Perm
CD15	Percpcy5.5	200	323030	W6D3	Biolegend	Post-Perm

Panel 5: IL-12 and IL-23 Donors 1-10 WB						
Antigen	Color	Titer	Catalog No.	Clone	Vendor	Pre vs Post
CD8	BUV805	100	612889	SK1	BD	Pre-Perm
CD4	BUV737	50	612748	SK3	BD	Pre-Perm
CD45RA	PE-Cy7	25	560675	HI100	BD	Pre-Perm
CD19	BV786	50	563325	SJ25C1	BD	Pre-Perm
CD56	BV711	100	318336	HCD56	Biolegend	Pre-Perm
CD16	BV711	100	302044	3G8	Biolegend	Pre-Perm
CD3	PE	50	552127	SP34-2	BD	Post-Perm
CD15	PE-Cy7	200	323030	SSEA-1	Biolegend	Post-Perm

Appendix 3: Gating Strategies

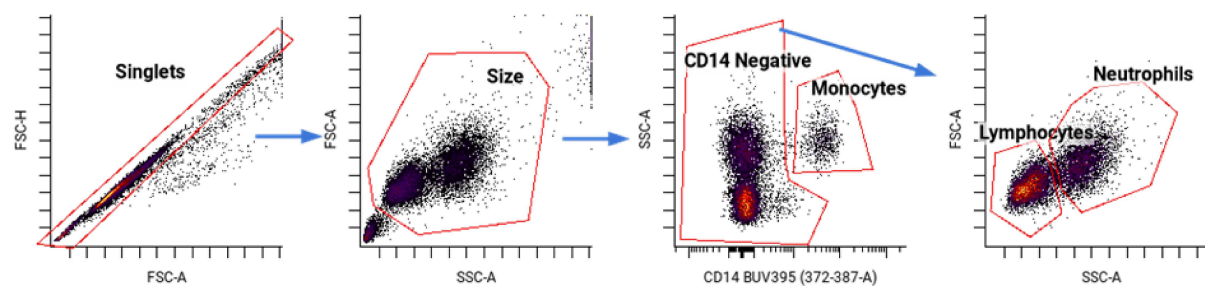
A) Gating strategy for common whole blood and PBMC cell populations (monocytes, CD4+, CD8+, NK cells, and B cells)

Initial phases of the gating strategy were common for whole blood and PBMCs

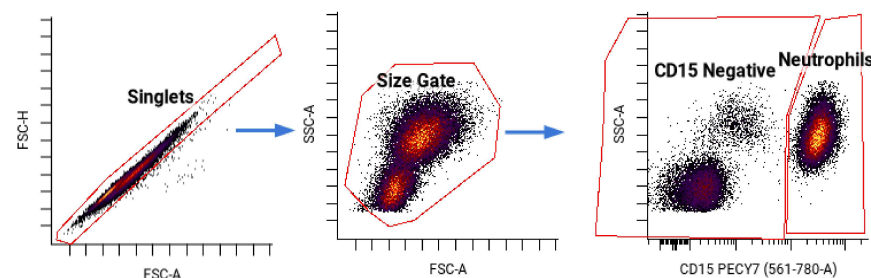


B) Gating strategy for neutrophils (whole blood). There were 3 different gating strategies depending on the antibody cocktail used.

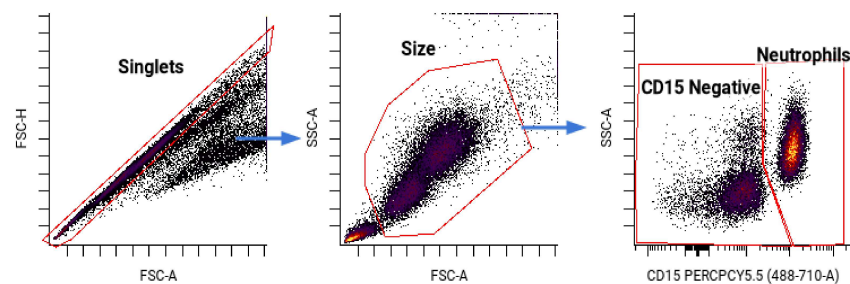
Strategy 1. No CD15 marker included.



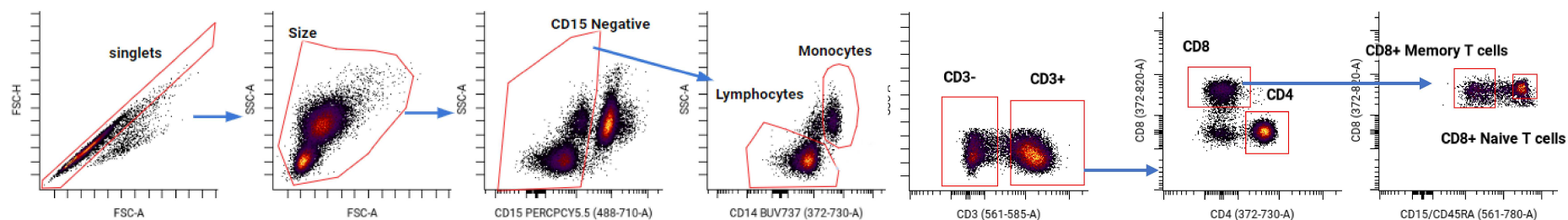
Strategy 2. CD15 PE-Cy7 marker.



Strategy 3. CD15 Percp-Cy5.5 marker.



C) Gating strategy for CD8+ memory T cells (IL-23 stimulation; Whole blood)



Appendix 4: Raw Flow Cytometry Data

Raw flow cytometry data is provided separately as an Excel document. Datasets are described below:

- Dataset 1: Donors 1-5. Whole blood and PBMCs data. Monocytes, B cells, CD4 T cells, CD8 T cells and NK cells Dataset 1-Neutrophils. Donors 1-5. Whole blood. Neutrophils
- Dataset 2: Donors 6-10. Whole blood data. Monocytes, B cells, CD4 T cells, CD8 T cells, NK cells and neutrophils
- Dataset 3: Donors 1-10. Whole blood data. IL-12 and IL-23 stimuli. NK cells and CD8+ memory T cells
- Dataset 4: Whole blood data. RA donors and matching healthy donors

Supplemental Table 1. Clinical Efficacy and Safety Findings From Phase 3 Studies of JAKinibs.

	Filgotinib	Tofacitinib	Upadacitinib	Baricitinib	
	200 mg	5 mg, bid	15 mg	2 mg	4 mg
MTX-IR ACR20/50/70/DAS28CR ^a	19/25/21/32 [1]	25/26/15/5 [2]	31/33/25/32 [3]	NR	37/32/22/33 ^b [4]
Biologic-IR ACR20/50/70/ DAS28CR ^c	35/28/15/14 [1]	17/18/12/4 [2]	37/22/5/20 [3]	22/12/11/9 ^d [4]	28/20/9/8 ^d [4]
Hemoglobin (g/dL)	+0.2 [5]	+0.08 [6]	-0.8 [7]	-0.28 [8]	-0.2 [8]
Herpes zoster (E/100PY)	0.1% (0.3% placebo) [1]	1-10% [2]	0.7% (0.2% placebo) [3]	1.4% (0.4% placebo) [4]	4.3% (1% placebo) [9]
Infections (E/100PY)	26.5 [1]	43.8 [2]	93.7 [3]	101 [4]	
Serious infection (E/100PY)	1.7 [1]	2.4 [2]	3.8 [3]	3.2 [4]	
Opportunistic infections (E/100PY)	0.1 [1]	<0.1 [2]	0.6 [3]	0 [9]	0.5 [9]
VTE (E/100PY)	0.2 [10]	0.27 [2]	0.5 [7]	0.6 [11]	0.8 [11]

^a MTX-IR response rates at week 24/26 (placebo-corrected); all patients on background MTX.

^b DAS28-hsCRP ≤ 3.2.

^c Biologic-IR response rates at week 12 (placebo corrected); DAS28 CR, complete remission is defined as DAS28<2.6; tofacitinib, baricitinib uses DAS28-ESR and other JAKinibs use DAS28-CRP, dose-dependent decrease; E/100 PY, events per 100 patient years; JAK, Janus kinase; JAKinib; Janus kinase inhibitor; NK, natural killer; NR, not reported; VTE, venous thromboembolism.

^d DAS28-ESR ≤ 3.2.

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Supplemental Table 2. JAKinib IC₅₀ Values in CD4+ T Cells, Monocytes, and NK Cells From PBMC Assays.

	CD4+ T Cells					Monocytes					NK Cells				
	BARI	FIL	MET	TOFA	UPA	BARI	FIL	MET	TOFA	UPA	BARI	FIL	MET	TOFA	UPA
Stimulation/pSTAT	IC ₅₀ , nM														
JAK2/2 or JAK2/TYK2-dependent cytokines															
G-CSF/pSTAT3	NS					13	985	11,630	67	13	NS				
GM-CSF/pSTAT5	NS					51	5012	59,031	240	24	NS				
JAK1/JAK2/TYK2-dependent cytokines															
IFN α /pSTAT1	20	618	9539	44	8	47	1440	34,045	108	19	53	1360	23,053	122	40
IFN α /pSTAT3	17	511	7720	39	7	8	275	4511	25	4	NS				
IFN α /pSTAT5	11	338	5227	23	5	8	224	3478	17	3	NS				
IFN γ /pSTAT1	NS					48	2785	41,945	165	26	NS				
IL-6/pSTAT1	18	512	5746	40	9	16	547	5457	46	6	NS				
IL-6/pSTAT3	NS					53	1556	20,908	157	25	NS				
JAK1/3-dependent cytokines															
IL-2/pSTAT5	16	447	7316	18	5	NS					27	875	11,473	31	13
IL-4/pSTAT6	29	744	20,257	38	11	20	604	17,497	52	9	19	388	10,974	29	8
IL-15/pSTAT5	14	453	7222	18	5	NS					29	978	13,072	34	14

G-CSF, GM-CSF, IFN α , IFN γ , IL-6, IL-2, IL-4, and IL-15, reported IC₅₀ values are based on the average of duplicate measures (N=3–5 healthy donors). Protein binding was not accounted for in the IC₅₀ calculations reported.

BARI, baricitinib; FIL: filgotinib; G-CSF, granulocyte colony stimulating factor; GM-CSF, granulocyte macrophage colony stimulating factor; IC₅₀, half maximum inhibitory concentration; IFN, interferon; IL, interleukin; JAK, Janus kinase; MET, GS-829845 (filgotinib metabolite); NK, natural killer; NS, not sampled; PBMC, peripheral blood mononuclear cells; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; TYK, tyrosine kinase; UPA, upadacitinib.

Supplemental Table 3. JAKinib IC₅₀ Values in B Cells, and CD8+ T Cells from PBMC Assays.

	B Cells					CD8+ T Cells				
	BARI	FIL	MET	TOFA	UPA	BARI	FIL	MET	TOFA	UPA
Stimulation/pSTAT	IC ₅₀ , nM									
JAK1/3-dependent cytokines										
IL-2/pSTAT5	NS					13	395	5377	16	5
IL-4/pSTAT6	106	2872	82,459	172	43	19	470	11,836	29	9
IL-15/pSTAT5	NS					20	685	10,370	25	8
JAK1/JAK2/TYK2-dependent cytokines										
IFNα/pSTAT1	56	1548	30,698	136	30	23	646	9465	45	10
IFNα/pSTAT3	13	380	6106	31	6	16	453	6628	36	8
IFNα/pSTAT5	9	255	4083	21	4	11	302	4236	21	5
IFNγ/pSTAT1	11	629	9303	38	9	NS				

IFN α , IFN γ , IL-2, IL-4, and IL-15, reported IC₅₀ values are based on the average of duplicate measures (N=3–5 healthy donors). Protein binding was not accounted for in the IC₅₀ calculations reported.

BARI, baricitinib; FIL, filgotinib; IC₅₀, half maximum inhibitory concentration; IFN, interferon; IL, interleukin; JAK, Janus kinase; MET, GS-829845 (filgotinib metabolite); NS, not sampled; PBMC, peripheral blood mononuclear cells; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; TYK2, tyrosine kinase 2; UPA, upadacitinib.

Supplemental Table 4. JAKinib Potency Differences Based on Cell Type.

Stimuli	pSTAT	Cell type A	Cell type B	BARI	FIL	MET	TOFA	UPA
G-CSF	STAT3	neutrophils	Monocytes	***	***	***	***	***
GM-CSF	STAT5	Monocytes	neutrophils	***	***	***	***	***
IFN α	STAT1	B cells	CD4+ cells	***	***	***	***	***
		B cells	CD8+ cells	***	***	***	***	***
		Monocytes	B cells		**	**		
		B cells	NK cells					
		CD8+ cells	CD4+ cells	*	*		*	**
		Monocytes	CD4+ cells	***	***	***	***	***
		NK cells	CD4+ cells	***	***	***	***	***
		Monocytes	CD8+ cells	***	***	***	***	***
		NK cells	CD8+ cells	***	***	***	***	***
		Monocytes	NK cells	*	***	***	*	*
	STAT3	CD4+ cells	B cells		**			
		CD8+ cells	B cells					
		Monocytes	B cells		***	**		
		NK cells	B cells					
		neutrophils	B cells	**	*	**		
		CD4+ cells	CD8+ cells					
		Monocytes	CD4+ cells					
		CD4+ cells	NK cells		**	*		
		neutrophils	CD4+ cells					
		Monocytes	CD8+ cells			*		
		CD8+ cells	NK cells					
		neutrophils	CD8+ cells			*		
		Monocytes	NK cells		***	***		*
		neutrophils	Monocytes					
		neutrophils	NK cells	*	*	***		*
	STAT5	CD4+ cells	B cells		**			
		CD8+ cells	B cells		*			
		Monocytes	B cells		*			*
		NK cells	B cells	*				**
		CD4+ cells	CD8+ cells					
		CD4+ cells	Monocytes					**
		NK cells	CD4+ cells					*
		CD8+ cells	Monocytes					**
		NK cells	CD8+ cells					*
		NK cells	Monocytes					***
IFN γ	STAT1	Monocytes	B cells	***	***	***	***	***
		neutrophils	B cells	***	***	***	***	***
		Monocytes	neutrophils				**	

IL-15	STAT5	CD8+ cells	CD4+ cells	**	***	*	**	**
		NK cells	CD4+ cells	***	***	***	***	***
		NK cells	CD8+ cells	*	*		**	***
IL-2	STAT5	CD4+ cells	CD8+ cells					
		NK cells	CD4+ cells	***	***	***	***	***
		NK cells	CD8+ cells	***	***	***	***	***
IL-4	STAT6	B cells	CD4+ cells	***	***	***	***	***
		B cells	CD8+ cells	***	***	***	***	***
		B cells	Monocytes	***	***	***	***	***
		B cells	NK cells	***	***	***	***	***
		B cells	neutrophils	***	***	***	***	***
		CD4+ cells	CD8+ cells	*		*	*	
		CD4+ cells	Monocytes				**	
		CD4+ cells	NK cells	***	***	***	**	
		CD4+ cells	neutrophils					
		Monocytes	CD8+ cells				***	
		CD8+ cells	NK cells					
		neutrophils	CD8+ cells				***	
		Monocytes	NK cells		*	***	***	
		neutrophils	Monocytes					
		neutrophils	NK cells	**		**	***	
IL-6	STAT1	Monocytes	CD4+ cells	*	*	***	*	
	STAT3	CD4+ cells	Monocytes	***	***	*	***	***

Comparisons between mean IC₅₀ values of different cell types in response to the same stimuli were performed using a 2-way mixed effect Analysis of Variance (ANOVA). Analysis based on in vitro whole blood measurements in healthy donors.

N=7–10 donors. **P*<0.05, ***P*<0.01, ****P*<0.001

BARI, baricitinib; FIL, filgotinib; IFN, interferon; IL, interleukin; JAK, Janus kinase; JAKinib; Janus kinase inhibitor; MET, GS-829845 (filgotinib metabolite); NK, natural killer; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; UPA, upadacitinib.

Supplemental Table 5. JAKinib Potency Differences Based on STAT Substrate.

Stimuli	Cell type	STAT A	STAT B	BARI	FIL	MET	TOFA	UPA
IFN α	B cells	STAT1	STAT3	***	***	***	***	***
	B cells	STAT1	STAT5	***	***	***	***	***
	B cells	STAT3	STAT5				*	
	CD4+	STAT1	STAT3					
	CD4+	STAT1	STAT5	**	**	**	***	**
	CD4+	STAT3	STAT5	*			**	*
	CD8+	STAT1	STAT3	***	**	**	**	***
	CD8+	STAT1	STAT5	***	***	***	***	***
	CD8+	STAT3	STAT5				**	*
	NK cells	STAT1	STAT3	***	***	***	***	***
	NK cells	STAT1	STAT5	***	***	***	***	***
	NK cells	STAT3	STAT5					
	monocytes	STAT1	STAT3	***	***	***	***	***
	monocytes	STAT1	STAT5	***	***	***	***	***
	monocytes	STAT3	STAT5	**	*	*	**	**
IL-6	CD4+	STAT3	STAT1	***	***	***	***	***
	monocytes	STAT3	STAT1	***	***	***	***	***

Comparisons between mean IC₅₀ values of different STATs activated in the same cell time and in response to the same stimuli were performed using a 2-way mixed effect ANOVA. Analysis based on in vitro whole blood measurements in healthy donors.

N=7–10 donors. **P*<0.05, ***P*<0.01, ****P*<0.001

BARI, baricitinib; FIL, filgotinib; IFN, interferon; IL, interleukin; JAK, Janus kinas; JAKinib; Janus kinase inhibitor; MET, GS-829845 (filgotinib metabolite); NK, natural killer; STAT, signal transducer and activator of transcription; TOFA, tofacitinib; UPA, upadacitinib.

Supplemental Table 6. JAKinib Potencies Impacted by Stimuli Utilizing the Same JAK Pair.

Cell type	STAT	Stimuli A	Stimuli B	BARI	FIL	MET	TOFA	UPA
Monocytes	STAT1	IFN γ	IL-6	**	***	***	***	***

Comparisons between mean IC₅₀ values observed for monocytes in response to 2 different stimuli signaling through the same JAK/STAT pair (JAK1/JAK2) were performed using a 2-way mixed effect ANOVA. Analysis based on in vitro whole blood measurements in healthy donors.

N=7–10 donors. ** P <0.01, *** P <0.001

BARI, baricitinib; FIL, filgotinib; IFN, interferon; IL, interleukin; JAK, Janus kinase; JAKinib, Janus kinase inhibitor; MET, GS-829845 (filgotinib metabolite); NK, natural killer; STAT, signal transducer and activator of transcription; TOFA, tofacitinib; UPA, upadacitinib.

Supplemental Table 7. Heatmap of Time Above 50% Inhibition at Clinical Doses of JAKinibs.

Signaling Pair	Cell Type	Stimuli Readout	Filgotinib ^M		Tofacitinib		Upadacitinib		Baricitinib	
			100mg	200mg	5mg	10mg	15mg	30mg	2mg	4mg
JAK1/TYK2	B cells	IFNα/pSTAT5	7 [†]	20	16 [†]	23	16 [†]	24*	9 [†]	19
	NK	IFNα/pSTAT5	7 [†]	20	14 [†]	22	12 [†]	18	5 [†]	13 [†]
	CD8+ T cells	IFNα/pSTAT5	6 [†]	17	15	22*	15	23*	8 [†]	16
	NK	IFNα/pSTAT3	6 [†]	17	13 [†]	20*	11 [†]	18	7 [†]	16
	monocytes	IFNα/pSTAT5	5 [†]	12	14	21*	21*	24*	8 [†]	18*
	CD4+ T cells	IFNα/pSTAT5	5 [†]	12	14*	22*	15*	23*	7 [†]	16*
	B cells	IFNα/pSTAT3	5 [†]	13	12	19*	13	21*	7 [†]	16*
	CD8+ T cells	IFNα/pSTAT3	4 [†]	10	10	17*	12	19*	5 [†]	13*
	neutrophils	IFNα/pSTAT3	4 [†]	8	12*	19*	15*	22*	3 [†]	10
	CD4+ T cells	IFNα/pSTAT3	3 [†]	7	9	16*	12*	19*	4 [†]	12*
	monocytes	IFNα/pSTAT3	3 [†]	7	10*	17*	16*	23*	4	12*
	CD4+ T cells	IFNα/pSTAT1	3 [†]	7	7	15*	11*	17*	2 [†]	9
	CD8+ T cells	IFNα/pSTAT1	2 [†]	6	5	12*	8	14*	1 [†]	7
	NK	IFNα/pSTAT1	0	1	0	2	1	3	0	0
	B cells	IFNα/pSTAT1	0	1	0	2	2	6*	0	0
	monocytes	IFNα/pSTAT1	0	0	0	1	4*	8*	0	0
JAK1/JAK2	CD4+ T cells	IL-6/pSTAT1	11 [†]	24	13 [†]	20 [†]	12 [†]	18 [†]	9 [†]	17 [†]
	monocytes	IL-6/pSTAT1	5 [†]	15	9 [†]	17	13	21*	5 [†]	13
	monocytes	IL-6/pSTAT3	0	0	0	2	3*	8*	0	0
	B cells	IFNγ/pSTAT1	4 [†]	9	13*	20*	13*	21*	12*	21*
	neutrophils	IFNγ/pSTAT1	0	1	3	11*	8*	14*	1	5*
	monocytes	IFNγ/pSTAT1	0	0	0	6*	6*	11*	0	5*
JAK1/JAK3	CD8+ T cells	IL-2/pSTAT5	2 [†]	5	14*	21*	11*	18*	2 [†]	19*
	CD4+ T cells	IL-2/pSTAT5	3 [†]	7	17*	23*	14*	21*	4 [†]	11*
	NK	IL-2/pSTAT5	0	2	8*	16*	6*	11*	0	4
	CD4+ T cells	IL-15/pSTAT5	3 [†]	9	19*	24*	15*	23*	5 [†]	13*
	CD8+ T cells	IL-15/pSTAT5	0 [†]	4	13*	21*	10*	16*	1 [†]	7*
	NK	IL-15/pSTAT5	0	2	8*	16*	6*	11*	0	3
	NK	IL-4/pSTAT6	3 [†]	6	15*	22*	12*	18*	4	12*
	CD8+ T cells	IL-4/pSTAT6	4 [†]	9	19*	24*	13*	21*	6 [†]	14*
	neutrophils	IL-4/pSTAT6	1	4	7*	15*	10*	16*	1	7
	monocytes	IL-4/pSTAT6	1 [†]	4	5	13*	10*	16*	1	8*
	CD4+ T cells	IL-4/pSTAT6	0 [†]	3	10*	17*	9*	15*	0 [†]	5
	B cells	IL-4/pSTAT6	0	0	1	3*	2	5*	0	1
JAK2/TYK2	monocytes	G-CSF/pSTAT3	0	0	1	4*	4*	9*	1	4*
	neutrophils	G-CSF/pSTAT3	0	0	0	0	0	1	0	0
	NK	IL-12/pSTAT4	0	0	0	0	0	0	0	0
	CD8+ m T cells	IL-23/pSTAT3	0	0	0	0	0	0	0	0
JAK2/JAK2	neutrophils	GM-CSF/pSTAT5	0	1	3	11*	14*	21*	0	6*
	monocytes	GM-CSF/pSTAT5	0	0	0	0	5*	9*	0	0

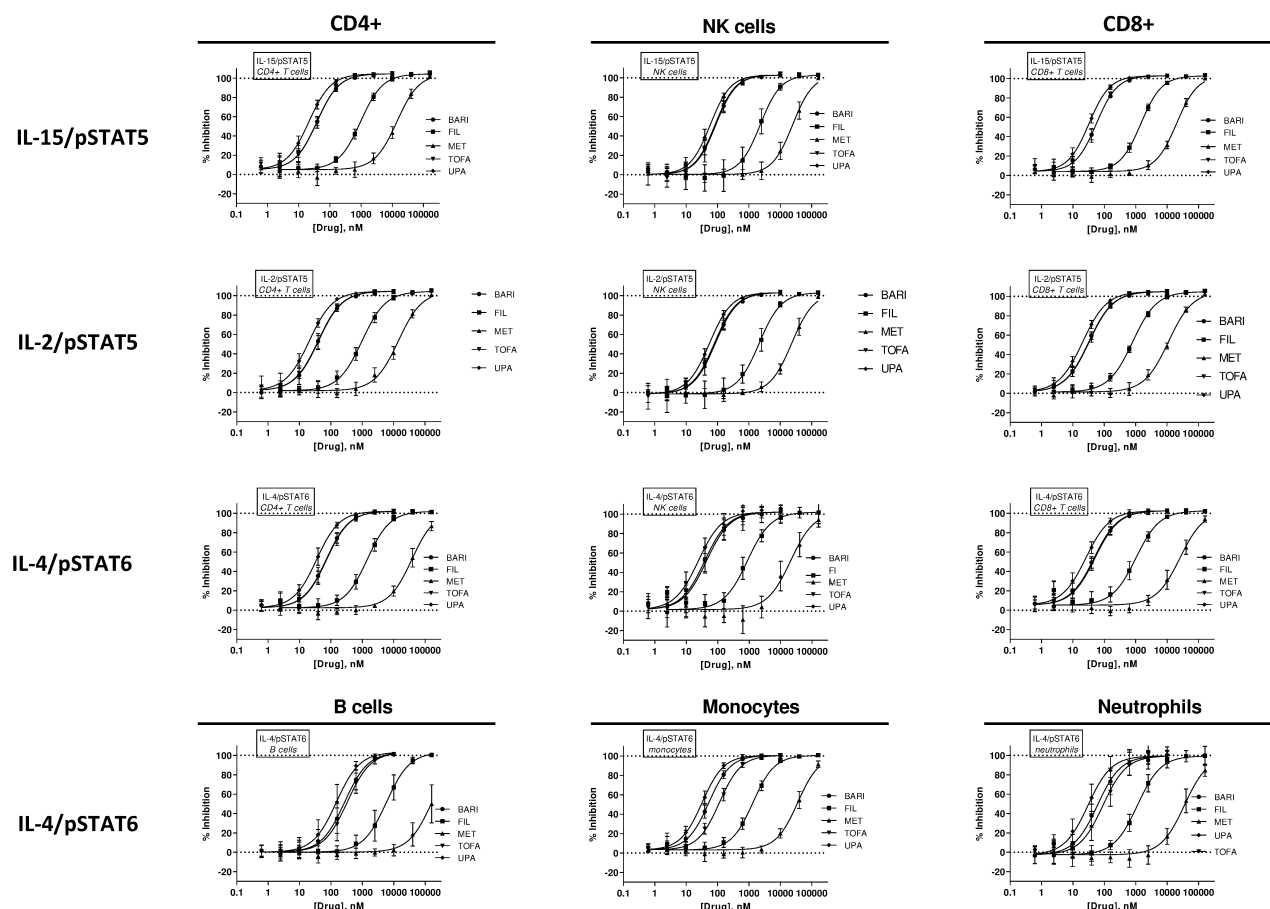


Heatmap represents time (hours) above 50% target inhibition by JAKinibs at clinical doses over a 24-hour dose period (based on in vitro WB measurements, N=6–10). FIL includes the metabolite exposure/activity calculation.

**P*<0.05 higher vs FIL 200 mg; †*P*<0.05 lower vs FIL 200 mg

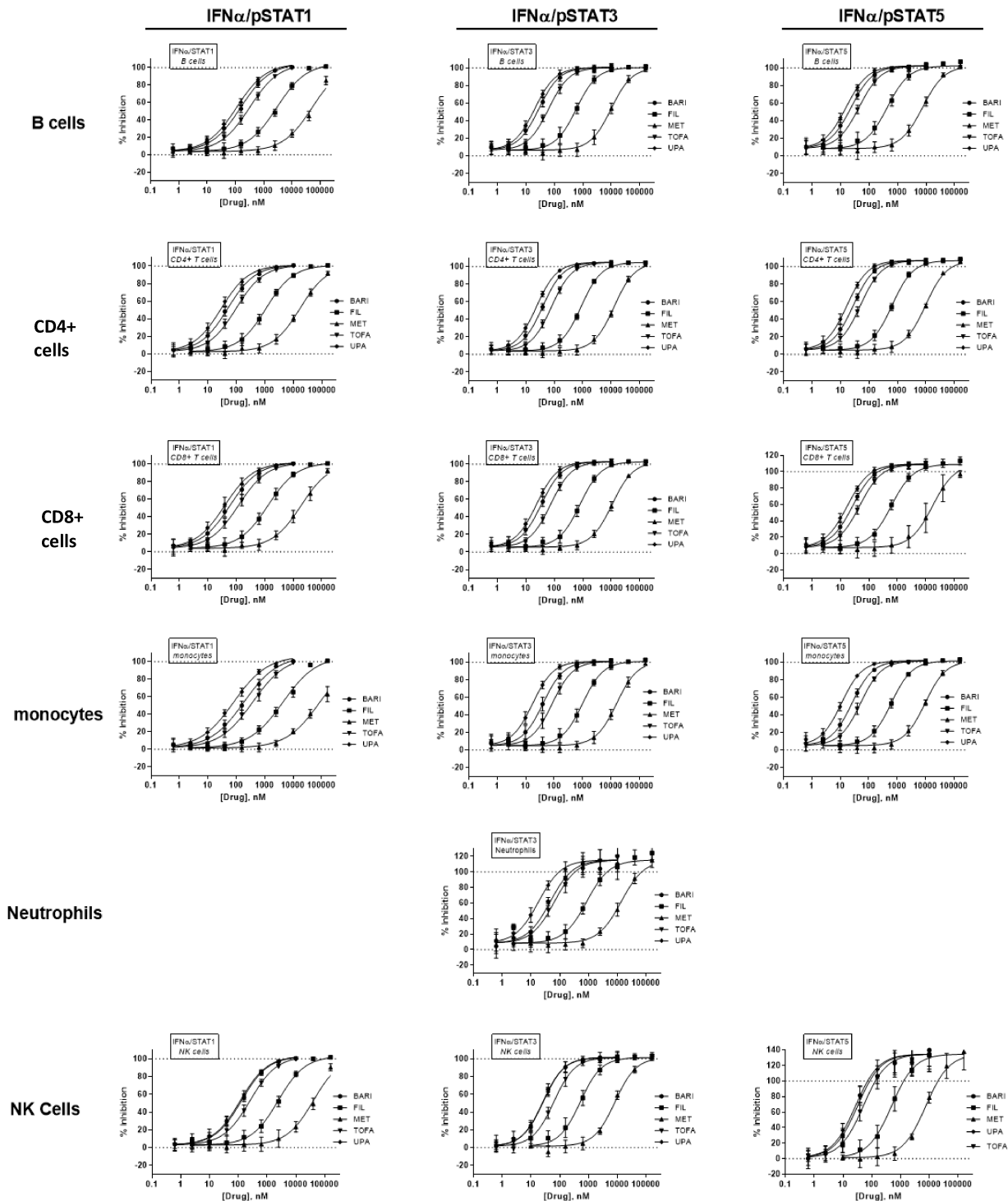
IFN, interferon; IL, interleukin; JAK, Janus kinase; JAKinib; Janus kinase inhibitor; NK, natural killer; pSTAT, phosphorylated signal transducer and activator of transcription; TYK2, tyrosine kinase 2.

Supplemental Figure 1.



JAKinib dose-response curves for STAT phosphorylation in response to JAK1/JAK3 dependent cytokines IL-2(pSTAT5), IL-4(pSTAT6) and IL-15(pSTAT5). Graphs represent percent pSTAT inhibition in whole blood from n=7–10 healthy volunteers. The percent inhibition values were calculated from duplicate wells of an 8-point dose response of each JAKinib and normalized to the difference in MFI measurements of cytokine stimulated minus unstimulated samples. Error bars represented SD. A four-parameter logistic fit model with shared upper and lower asymptotes and hillslope were used to fit the composite data.

Supplemental Figure 2.

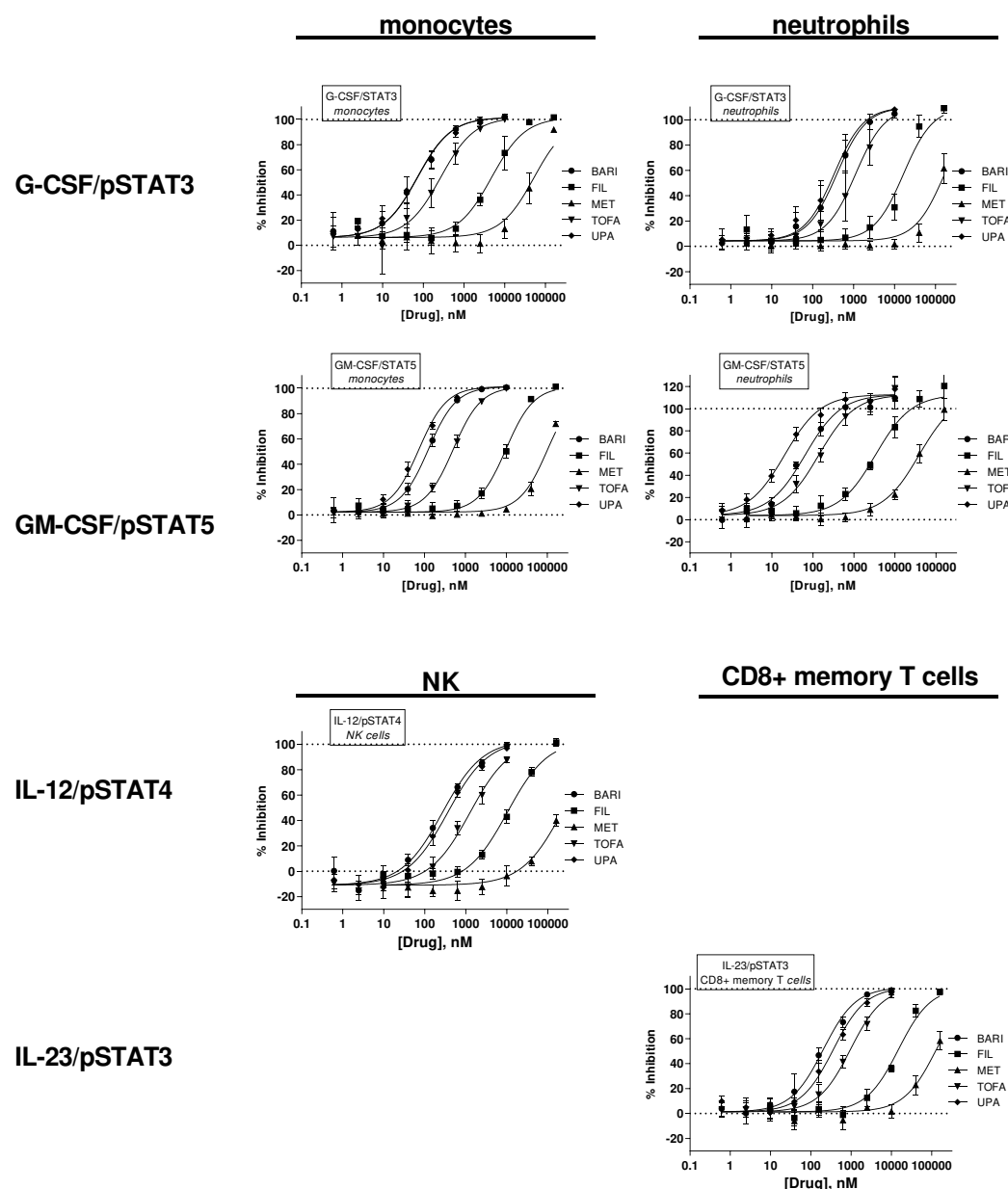


JAKinib dose-response curves for STAT (pSTAT1, pSTAT3, pSTAT5) phosphorylation in response to JAK1/TYK2 dependent cytokine IFNα. Graphs represent percent pSTAT inhibition in whole blood from n=9–10 healthy volunteers. In this figure, as well as

Supplemental Figures 3 and 4, the percent inhibition values were calculated from duplicate wells of an 8-point dose response of each JAKinib and normalized to the difference in MFI measurements of cytokine stimulated minus unstimulated samples. Error bars represented SD. A 4-parameter logistic fit model with shared upper and lower asymptotes and hillslope were used to fit the composite data.

BARI, baricitinib; FIL, filgotinib; IL, interleukin; JAK, Janus kinase; JAKinib, Janus kinase inhibitor; MET, GS-829845 (filgotinib metabolite); NK, natural killer; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; TYK2, tyrosine kinase 2; UPA, upadacitinib.

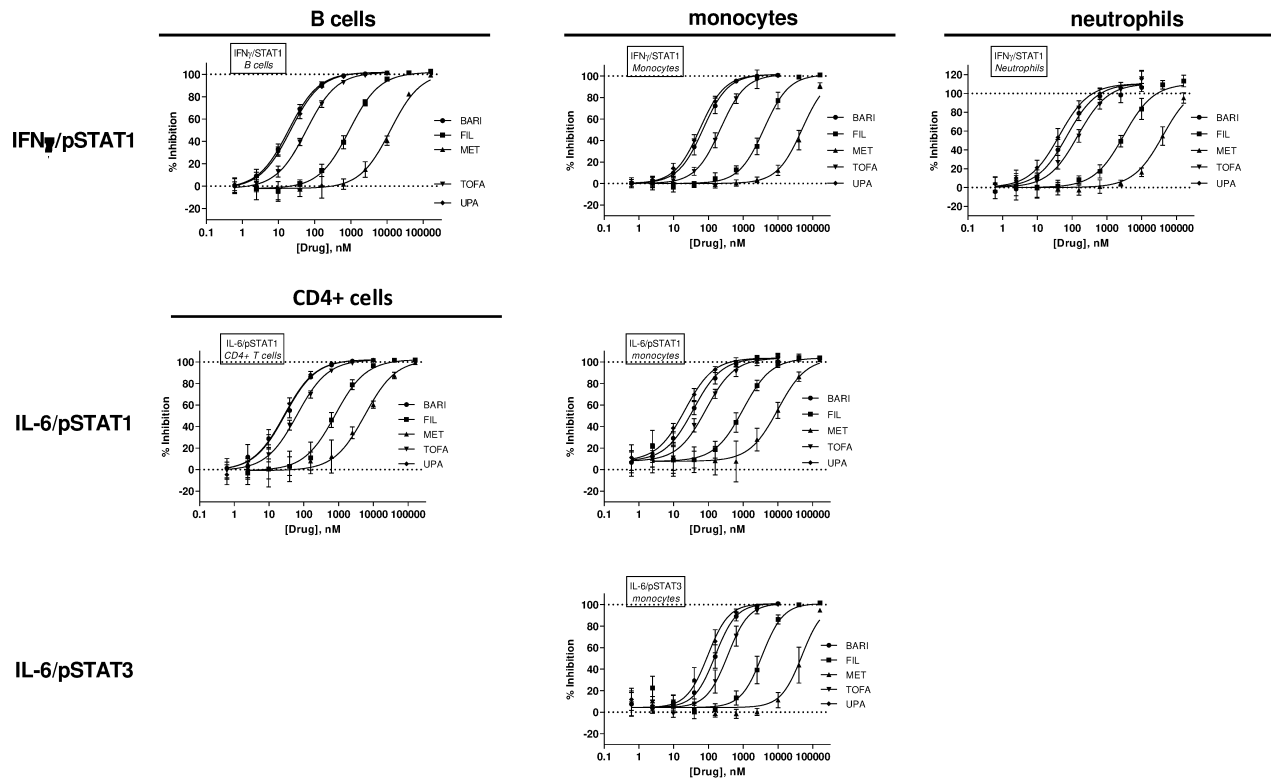
Supplemental Figure 3.



JAKinib dose-response curves for STAT phosphorylation in response to JAK2/TYK2-dependent cytokines G-CSF (pSTAT3), IL-12 (pSTAT4), and IL-23 (pSTAT3) and JAK2/JAK2-dependent GM-CSF (pSTAT5). Graphs represent percent pSTAT inhibition in whole blood from n=6–10 healthy volunteers.

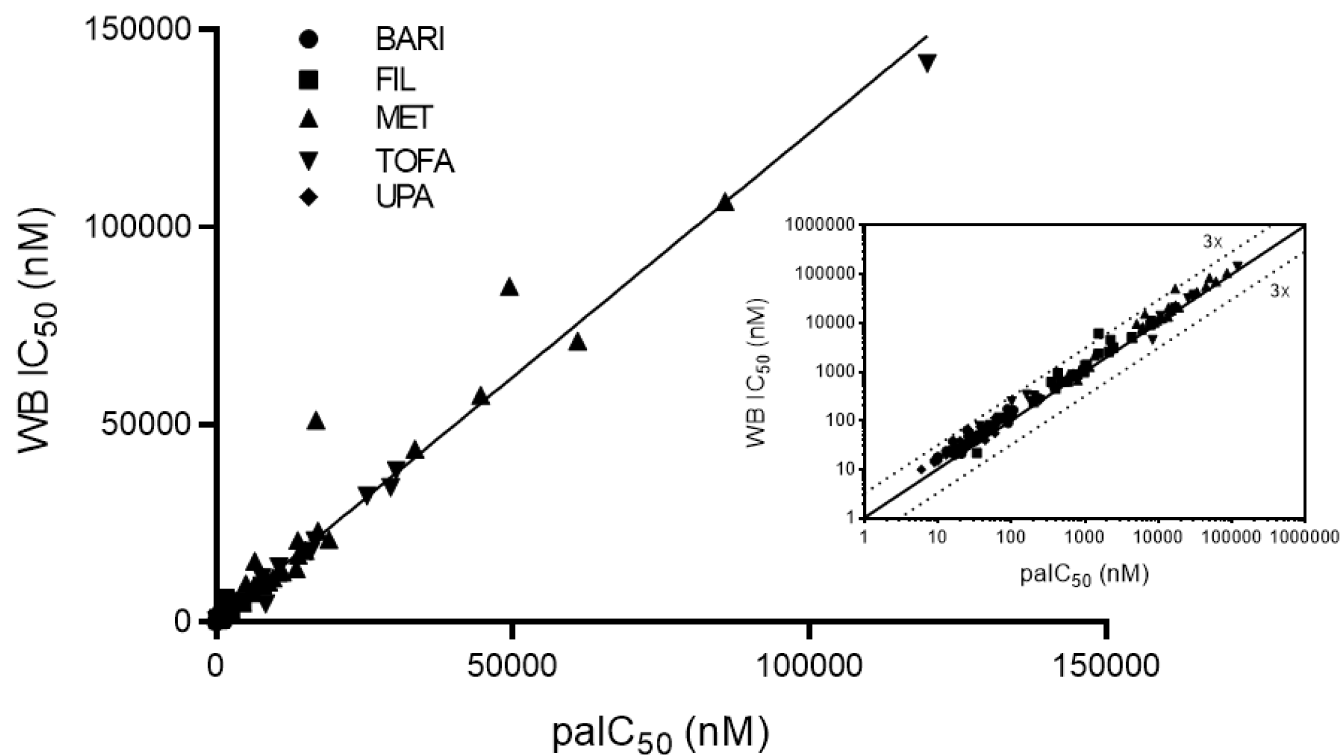
BARI, baricitinib; FIL, filgotinib; IL, interleukin; G-CSF, granulocyte-colony stimulating factor; GM-CSF, granulocyte macrophage-CSF; JAK, Janus kinase; JAKinib, Janus kinase inhibitor; MET, GS-829845 (filgotinib metabolite); NK, natural killer; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; TYK2, tyrosine kinase 2; UPA, upadacitinib.

Supplemental Figure 4.



JAKinib dose-response curves for STAT phosphorylation in response to JAK1/JAK2 dependent cytokines IFN γ (pSTAT1) and IL-6 (pSTAT1, pSTAT3). Graphs represent percent pSTAT inhibition in whole blood from n=7–10 healthy volunteers. BARI, baricitinib; FIL, filgotinib; IL, interleukin; INF, interferon; JAK, Janus kinase; JAKinib; Janus kinase inhibitor; MET, GS-829845 (filgotinib metabolite); pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; TYK2, tyrosine kinase 2; UPA, upadacitinib.

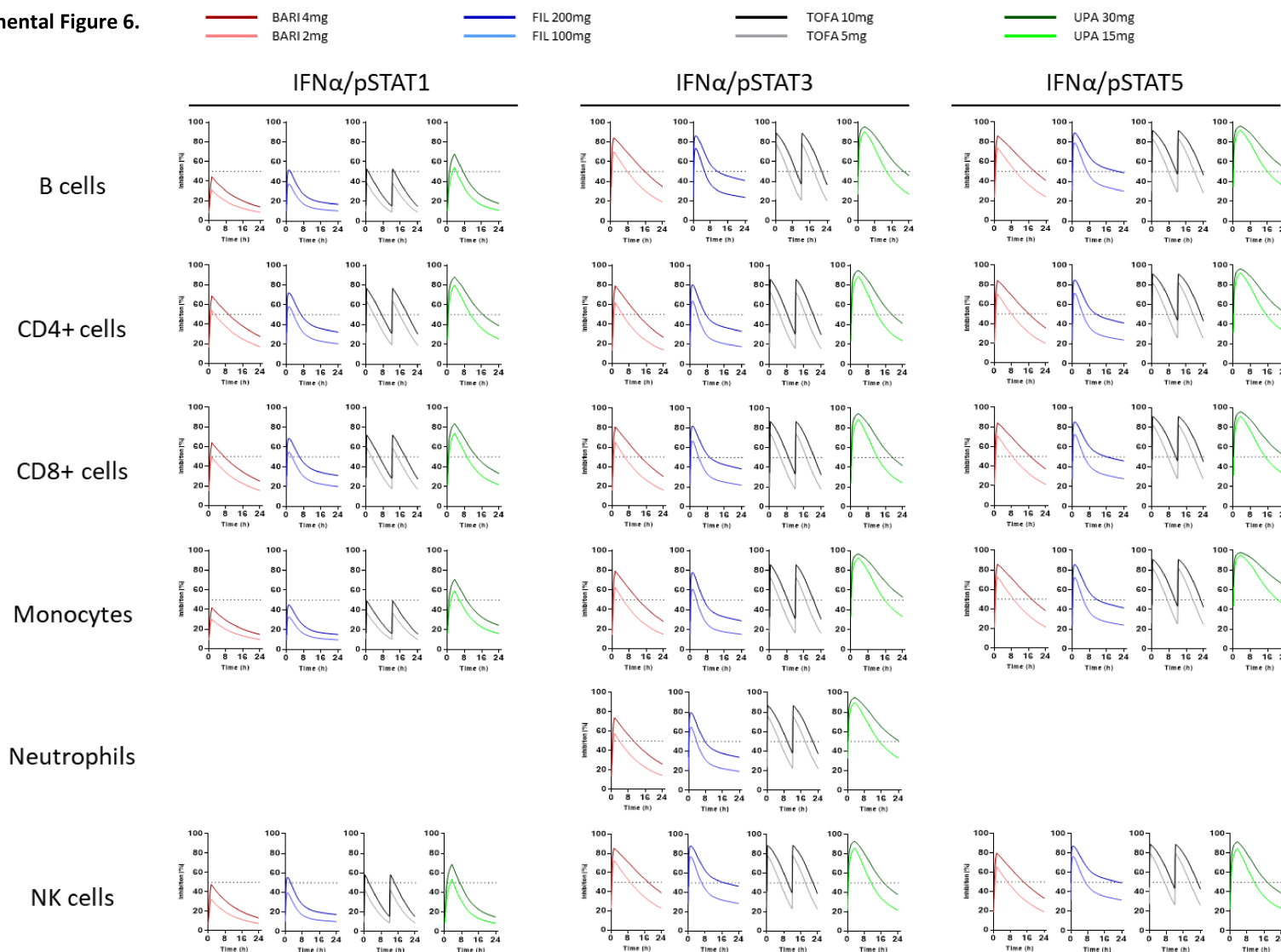
Supplemental Figure 5.



palC₅₀ values from PBMCs (n=5 donors) were compared to IC₅₀ values in whole blood from the same donors (n=5). Pearson's analysis indicated a coefficient of determination $r^2=0.98$.

BARI, baricitinib; FIL, filgotinib; ; IC₅₀, half maximum inhibitory concentration; MET, GS-829845 (filgotinib metabolite); NK, natural killer; palC₅₀, plasma binding-adjusted IC₅₀; PBMCs, peripheral blood mononuclear cells; TOFA, tofacitinib; UPA, upadacitinib; WB, whole blood.

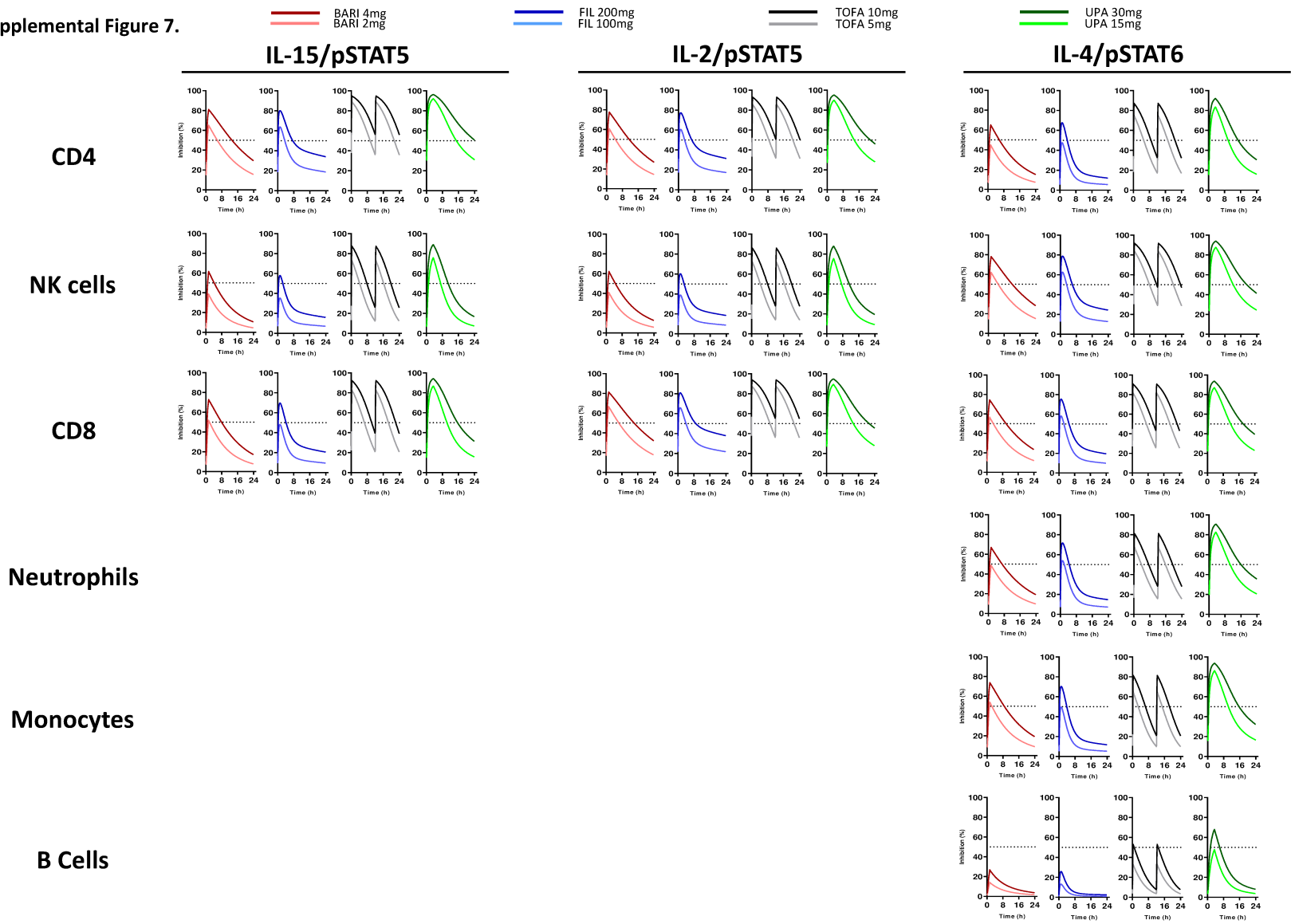
Supplemental Figure 6.



Inhibition curves of IFN α -induced STAT phosphorylation by JAKinibs (24-hour dose interval) at clinical doses (calculated based on in vitro WB measurements) from healthy volunteers) n=9–10 donors. In this figure, as well as Supplemental Figures 7-9, filgotinib takes into consideration the presence of the active human metabolite, GS-829845, and its potency. Curves above the dashed line (at 50%) indicates the time above 50% target coverage of each JAKinib during the dose interval.

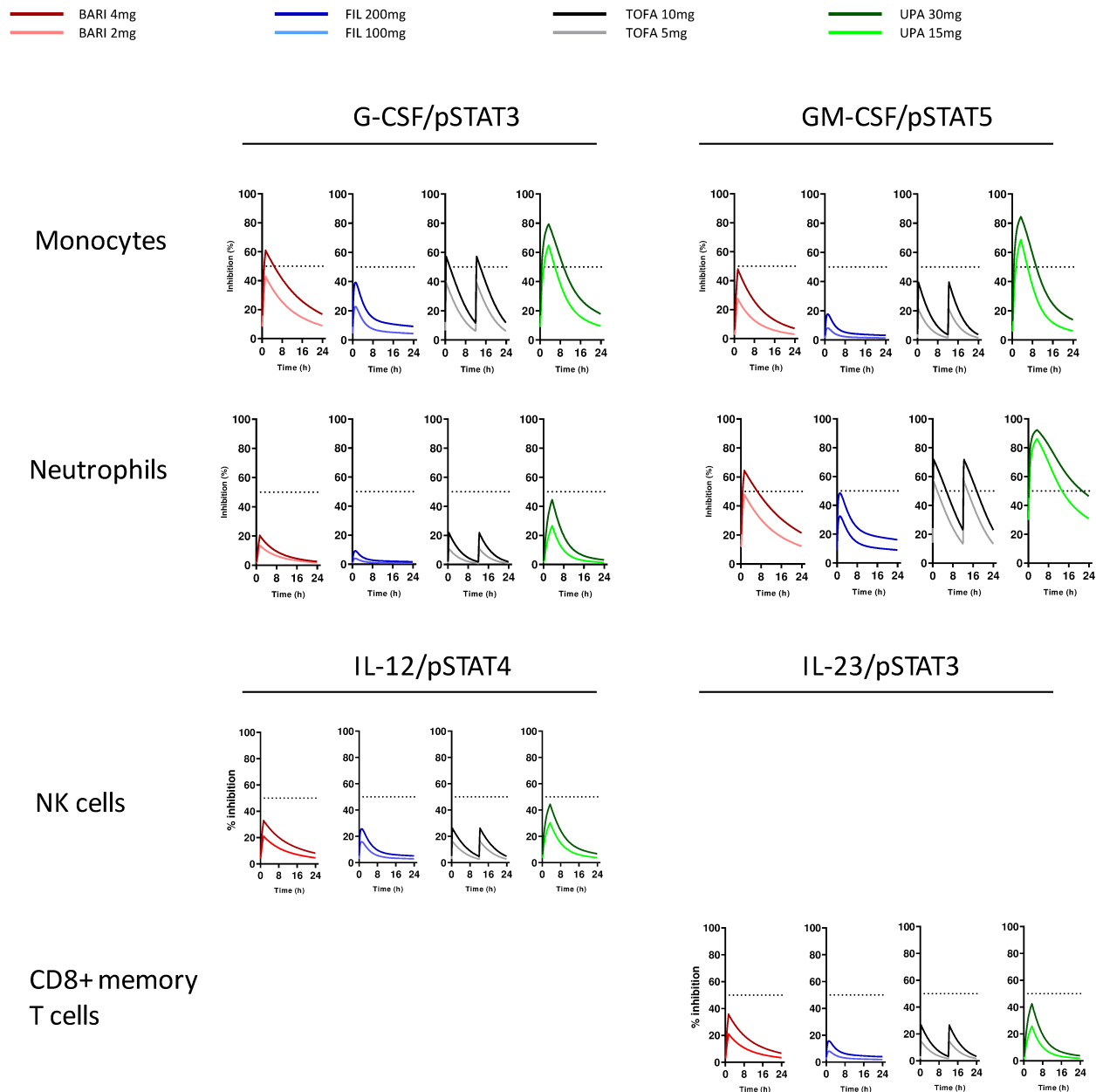
BARI, baricitinib; FIL, filgotinib; IFN, interferon; JAK, Janus kinase; JAKinib; Janus kinase inhibitor; NK, natural killer; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; UPA, upadacitinib; WB, whole blood.

Supplemental Figure 7.



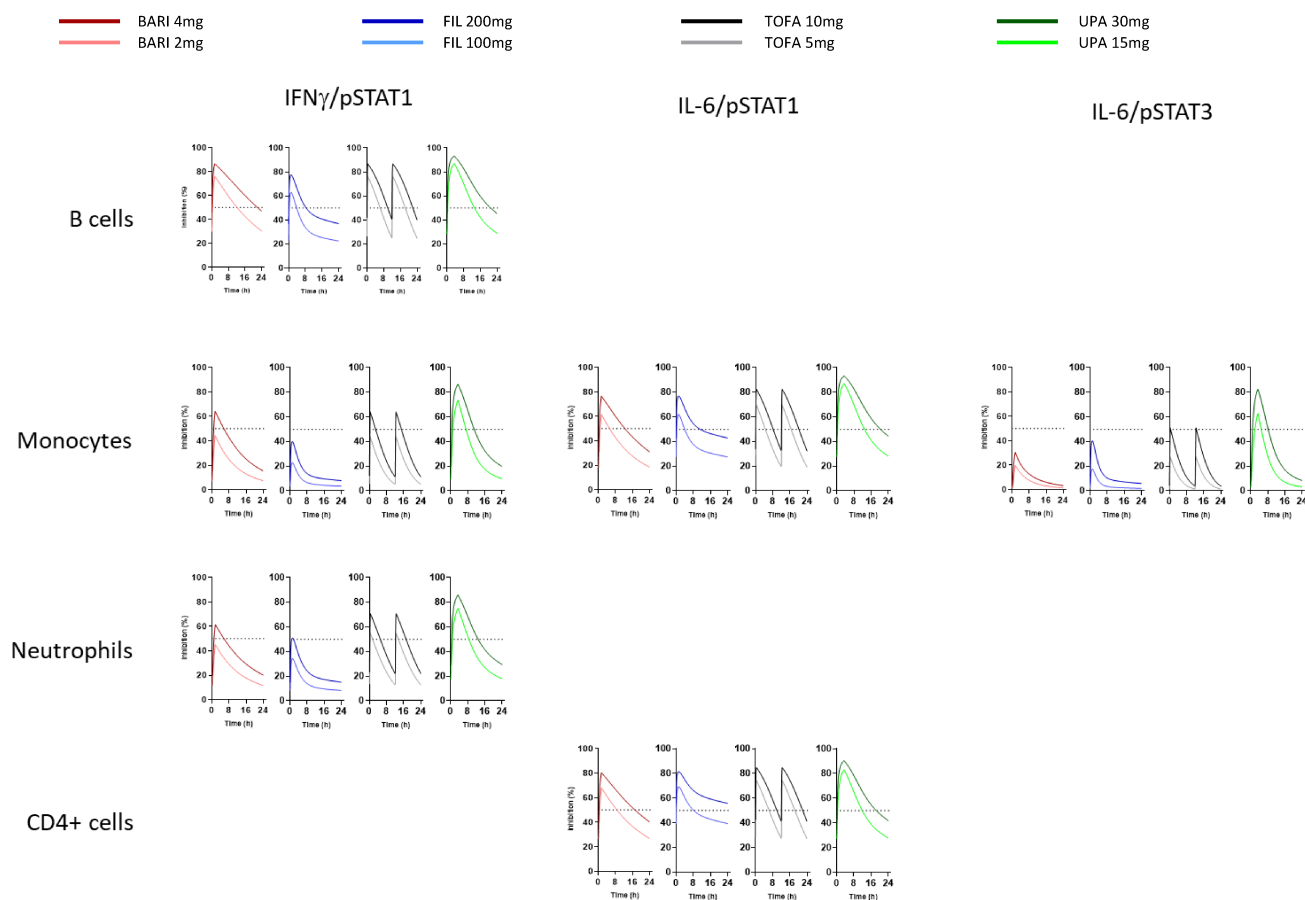
Inhibition curves of IL-2, IL-4 and IL-15-induced STAT phosphorylation by JAKinibs (24 h dose interval) at clinical doses (calculated based on in vitro WB measurements) from healthy volunteers). N=7–10 donors. BARI, baricitinib; FIL, filgotinib; IL, interleukin; JAK, Janus kinase; JAKinib; Janus kinase inhibitor; NK, natural killer; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; UPA, upadacitinib; WB, whole blood.

Supplemental Figure 8.



Inhibition curves of G-CSF, IL-12, IL-23 and GM-CSF-induced STAT phosphorylation by JAKinibs (24-hour dose intervals) at clinical doses (calculated based on in vitro WB measurements from healthy volunteers). N=6–10 donors.
BARI, baricitinib; FIL, filgotinib; IL, interleukin; G-CSF, granulocyte-colony stimulating factor; GM-CSF, granulocyte macrophage-CSF; JAK, Janus kinase; JAKinib; Janus kinase inhibitor; NK, natural killer; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; UPA, upadacitinib; WB, whole blood.

Supplemental Figure 9.



Inhibition curves of IFN γ and IL-6-induced STAT phosphorylation by JAKinibs (24-hour dose intervals) at clinical doses (calculated based on in vitro WB measurements from healthy volunteers). N=7–10 donors.

BARI, baricitinib; FIL, filgotinib; IL, interleukin; INF, interferon; JAKinib; Janus kinase inhibitor; NK, natural killer; pSTAT, phosphorylated signal transducer and activator of transcription; TOFA, tofacitinib; UPA, upadacitinib; WB, whole blood.