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compare the effect of valency on IC-mediated clearance. This FL antibody showed very similar TNF neutralization compared to the conventional PF molecule in a bioassay. To generate ICs, the anti-ID was incubated overnight with either PF or FL CZP, and the size of the resultant ICs determined by analytical ultracentrifugation (AU). The anti-ID complexes and PF and FL CZP alone were then administered IV to the mice, and the elimination of the anti-TNFs from the circulation was monitored by quantitative liquid chromatography-mass spectrometry (LC-MS) in serial plasma samples.

Results: AU analysis of the immune complexes formed between PF CZP and the anti-ID showed the presence of one peak corresponding in size to one anti-ID molecule bound to two PF molecules (~3.5x10⁵ Da). In contrast, the FL CZP/anti-ID mixture showed ICs of various sizes up to very large molecular weights (>1x10 6 Da), with the predominant species corresponding to a complex of two anti-IDs bound to two FL CZP molecules (~6x10⁵ Da). The in vivo studies showed that the FL CZP/anti-ID ICs were eliminated much faster (t_{1/2}=0.3 hours) than the FL CZP alone (t_{1/2}=44.5 hours), whereas the PF CZP/anti-ID ICs were eliminated more slowly ($t_{1/2}$ =60.5 hours) than the PF CZP alone ($t_{1/2}$ =19.7 hours). Conclusions: The FL CZP molecule formed large ICs with the anti-ID, which led to much faster elimination than the FL molecule alone. This result suggests that an ADAb response to a FL antibody could lead to rapid elimination and loss of efficacy of the drug in patients. In contrast, the PF CZP/anti-ID complex had a longer half-life than the PF CZP alone, presumably because this molecule was not seen as an IC due to the presence of only one Fc. These results showed that an ADAb to a univalent biological reagent may not lead to elimination and instead, may actually increase the in vivo half-life of the molecule.

References:

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Acknowledgements: This study was funded by UCB Pharma. Editorial services were provided by Costello Medical Consulting.

Disclosure of Interest: J. Silva Employee of: UCB Pharma, A. Nesbitt Employee of: UCB Pharma

DOI: 10.1136/annrheumdis-2017-eular.2241

FRI0194 USE OF GLORESPONSETM NF-KB-RE-LUC2P HEK293 CELLS TO MONITOR DRUG AND ANTI-DRUG ANTIBODY LEVELS IN

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Background: Rheumatoid Arthritis (RA) is often treated with anti-TNF α such as infliximab (Ifx) which in a long-term treatment can lead to the development of anti-Ifx antibodies (ATI), resulting in an interference with the drug activity. The investigation of the bioactivity of the circulating drug and antibodies present in patients sera with inflammatory diseases will allow to harmonize the different published data using both bioassays as well as immunoassays.

Objectives: To evaluate the ability of the Promega bioassay in the quantification of both Ifx and ATI in serum from RA patients. To compare the bioassay performance between capture- and bridging-ELISA.

Methods: Serum Ifx-trough levels were determined in 50 samples from patients with RA. To measure Ifx, the *bioassay* uses GloResponse™ NF-kB-RE-luc2P HEK293 cell line, which responses to TNF $\!\alpha$ resulting in modulation of NF- $\!\kappa B$ activity (Promega Corp., Madison, USA). This is an ease on performing reporter assay (add-mix-read) that detects the serum Ifx by inhibition of the luminiscent signal due to TNF α . The inhibition of the Ifx-driven luminescent signal is reverted by the presence of ATI in the serum. ATI concentration is determined using a standard curve obtained with a serial dilution of a serum sample known to have ATI. A relative luminescent unit (RLU) is arbitrarily defined. The immunoassay for Ifx (ELISA) captures recombinant TNF α , via a monoclonal antibody that does not interfere with the serum Ifx binding to TNF α . The reaction is developed with a biotinylated anti-idiotype antibody 1 (Cut-off for Ifx: 0.050 μ g/ml). The ELISA to detect ATI takes advantage of the bivalency of most IgG subclasses, allowing the antibodies present in serum to bridge between Ifx-coated onto the plate with biotinylated Ifx in fluid phase1 (Cut-off for ATI 50 AU/ml).

Results: Addition of TNF α on GloResponse cells induces the production of luciferase resulting in an increase of the luminescent signal in a dose-response. Addition of Ifx on GloResponse cells +TNF α decreases the production of luciferase and therefore the luminescent signal falls (r^2 =0.99). Addition of anti-lfx (in relative units) on GloResponse cells +TNF α + Ifx restore the production of luciferase resulting in an increase of the luminescent signal (r²=0.98).

Serum Ifx concentrations were sorted into 3 groups (low, medium and high following ELISA levels). Similar number of positive Ifx results was found (30 samples, 55% by ELISA vs 28 samples, 52% by bioassay; κ =0.92; p=0.56) along with a significant Pearson correlation (r=0.8) between methods. The bioassay to measure ATI activity showed lower sensitivity than ELISA (9/31, 29% by ELISA vs 4/31, 13% by bioassay, κ=0.53, p=0.22) with a slightly less correlation for ATI levels by both methods (r=0.26).

Conclusions: Bioassay using GloResponse NF-κB cell line can be performed to

monitor therapeutic drug of TNF α blocker and are useful to detect the presence of anti-drug antibody in serum samples treated with anti-TNF α antibodies. Comparison with ELISA method shows in most of the case the same data.

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Disclosure of Interest: None declared DOI: 10.1136/annrheumdis-2017-eular.6352

FRI0195 POINT-OF-CARE MONITORING OF ANTI-INFLIXIMAB ANTIBODIES IN PATIENTS TREATED WITH THE REFERENCE INFLIXIMAB OR CT-P13 IN ROUTINE CLINICAL PRACTICE

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Background: Loss of clinical response and infusion reactions to infliximab (IFX) are associated to the development of antibodies to IFX (ATI). ATI detection is a key step of patient management. However, current techniques may require additional patient appointments for sample collection, processing and batching in centralised facilities. Test reporting usually takes several days or weeks impairing effective decision making.

Objectives: To clinically validate the use of a new rapid test to detect ATI in capillary blood in a real-life point-of-care (POC) setting where patients attend the infusion center for the reference IFX (Remicade®, RMC) or CT-P13 (Inflectra®, IFT, or Remsima®) infusions.

Methods: PQ-EF1 and PQ-EF2 are prospective, observational studies designed to evaluate and compare the performance of a rapid POC test (CE-marked Promonitor® Quick Anti-IFX, Progenika, Spain) to detect ATI in routine clinical practice in rheumatic and gastroenterology patients treated with the reference IFX or the biosimilar CT-P13 attending the infusion center with the ELISA technique as a reference. The POC test is a qualitative immunochromatographic assay based on lateral flow technology to detect ATI (including biosimilar CT-P13) in either fingerprick or serum or venous whole blood. Consecutive patients (initiating or under maintenance therapy) were recruited and tested in La Paz and Basurto University hospitals with the rapid test in capillary and venous whole blood specimens immediately before the infusion. ATI test results were read visually with the POC test in 30 min, just before the patient started the infusion. Trough sera were also collected for subsequent analysis with the rapid test and benchmarked with Promonitor®-Anti-IFX ELISA (Progenika, Spain). Follow-up time was 6 months. ELISA quantitative results were categorized as positive and negative to allow comparisons with the qualitative rapid test.

Results: Ninety consecutive patients were recruited (a total of 137 visits in the 6 months follow-up) accounting for a total of 137 sera, 137 fingerpricks and 71 venous whole blood samples. Overall, 8 (8.9%) patients developed ATI (5 ankylosing spondylitis, 1 Crohn's disease, 1 ulcerative colitis and 1 juvenile idiopathic arthritis). ATI were detected in 5 patients treated with Remicade® and 3 treated with Inflectra®. Overall agreements between fingerprick vs venous whole blood and fingerprick vs serum measured with the rapid POC test were 100% and 99%, respectively. Positive (PPA) and negative (NPA) agreements between the POC test and ELISA were 91% and 99%, respectively. PPA and NPA between the ELISA and the POC test in serum was 100% and 99%, respectively.

Conclusions: ATI can be reliably detected in either venous or capillary circulation. Results show an almost perfect agreement between specimens and with the reference ELISA technique. ATI measurement with the POC test allows the clinician to detect ATI in a quick and fully decentralised mode facilitating immediate POC decision making.

Disclosure of Interest: A. Ametzazurra Employee of: Employee of Progenika Grifols, N. Rivera: None declared, A. Balsa: None declared, M. Arreba: None declared, E. Ruiz: None declared, C. Plasencia: None declared, J. Ortiz: None declared, D. Pascual-Salcedo: None declared, M. Muñoz: None declared, C. De Aysa: None declared, M. Allande: None declared, N. Torres Employee of: Employee of Progenika Grifols, A. Hernández Employee of: Employee of Progenika Grifols, X. Recalde Employee of: Employee of Progenika Grifols, A. Martínez Employee of: Employee of Progenika Grifols, D. Nagore Employee of: Employee of Progenika Grifols

DOI: 10.1136/annrheumdis-2017-eular.5056

FRI0196

BIOSIMILARS IN THE UK: EARLY REAL WORLD DATA FROM THE BRITISH SOCIETY FOR RHEUMATOLOGY BIOLOGICS REGISTERS FOR RHEUMATOID ARTHRITIS

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Background: Biosimilars, biopharmaceuticals assessed by regulatory agencies