Conspicuous synovial lymphatic capillaries in juvenile idiopathic arthritis synovitis with rice bodies

E Rovenska, S Stvrtina, O Greguska, L Pravda, J Rovensky


Recently, mature VEGF-C was found in rheumatoid arthritis synovial tissue.10

Figure 1 Synovial LC (asterisks) in semi-thin resin sections stained with toluidine blue. (A) LC in the SM area covered with fibrinous material (f). Walls of the LC are composed of endothelial cells only. (B) Mononuclear phagocytes (arrows) in the vicinity of the LC. One phagocyte (large arrow) and debris (arrowheads) are visualised inside the lumina of the LC. (C) Large LC with an irregular shaped lumen surrounded by numerous mononuclear cells, situated in the sublining connective tissue of the SM under the intimal layer showing hyperplasia. Some blood capillaries (arrows) and a venule (arrowhead) are also visualised. JS, joint space. Scale bars = 100 \( \mu \)m.
Our observation of conspicuous LC suggests that lymphangiogenesis may occur in JIA synovitis. In chronic synovitis, neogenesis of LC seems to be aimed at improving drainage and thus promoting homoeostasis in the JS.

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**REFERENCES**


6 Hauck G. The connective tissue space in view of the lymphatics. Experientia 1982;38:1121–2.


**Periostitis as the initial manifestation of systemic vasculitis**

P M Aries, M Reuter, P Lamprecht, W L Gross


The greatest challenge in diagnosing vasculitis is the diversity of its clinical presentation. Awareness of the heterogeneity of uncommon manifestations can be decisive for the course of the disease.

**CASE REPORT**

We report on a patient presenting with periostitis as the initial manifestation of systemic vasculitis. A 38 year old female patient complained about progressively painful swelling and reddening of the distal right lower leg for several weeks. The patient had been healthy until then and had no history of arterial or venous insufficiency. She presented at hospital with reduced pulses and a severe compartment syndrome of the tibialis anterior compartment. An angiography. According to the nomenclature of the Chapel Hill Consensus Conference, the patient’s disease was diagnosed as polyarteritis nodosa. Treatment was switched to cyclophosphamide (the so-called “NIH standard”: cyclophosphamide 2.0 mg/kg body weight per day with daily prednisolone po1). After induction of remission, treatment was switched to azathioprine. Follow up bone radiography disclosed a moderate reduction of the new periostal bone formation and clinical remission was maintained at a 3 year follow up.

**DISCUSSION**

The patient presented initially with an unusual manifestation of systemic vasculitis. Vasculitis restricted to the local vascular region may be the initial manifestation of systemic vasculitis. In this particular case, vasculitis of the periostium might have induced local hypoxia of the bone, with subsequent release of bone derived growth factors and manifestation of periostitis. Periostitis is seen in many other conditions but is common in necrotising vasculitis. It was described for the first time by Lovell and Scott in 1956. Until now only a few cases of periostitis in patients with polyarteritis nodosa have been reported; remarkably, the lower extremities were affected in all cases. However, periostitis has also been reported in other forms of systemic vasculitis. Most cases responded well to glucocorticoids. In refractory cases other cytotoxic treatment like methotrexate, azathioprine, or cyclophosphamide may be useful.

Thus, as demonstrated by this case, in patients with painful swelling of the lower limb, clinicians should consider periostitis as an unusual manifestation of systemic vasculitis.

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REFERENCES

Figure 1 (A) Bone radiography of the right lower leg with periostal new bone formation (arrow). (B) A magnified view.

Presence of rheumatoid factor and antibodies to citrullinated peptides in systemic lupus erythematosus


Rheumatoid factor (RF) is found commonly in patients with systemic lupus erythematosus (SLE), and has been associated with a more benign disease course.1,2 Anti-citrullinated peptide antibodies (ACPA) are more specific for rheumatoid arthritis (RA).3–5 Several assays for ACPA detection have been developed: among others, an enzyme linked immunosorbent assay (ELISA) for anti-cyclic citrullinated peptide (anti-CCP) antibodies3 and a line immunoassay (LIA) for antibodies to peptide A (pepA) and peptide B (pepB), two synthetic citrullinated peptides.4 Few reports exist about the presence of ACPA in SLE. Although patients with SLE are often part of the control group when determining the specificity of ACPA for RA, SLE alone is seldom studied. Mediwake et al found that 3/66 patients with SLE were positive for anti-CCP1 antibodies; two of them had erosive arthritis.6 We investigated the presence of RF and three different ACPA (anti-CCP, anti-pepA, and anti-pepB antibodies) in SLE.

Two hundred and thirty five patients with SLE, meeting American College of Rheumatology (ACR) revised criteria for classification of SLE,7,8 were prospectively included in four European centres. The study investigated associations between symptoms and specific antinuclear reactivities and has been reported elsewhere.9 Serum was available for further analysis in 201 patients. The male to female ratio was 25:176. The mean age was 40 years. The study was...
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Antibodies were detected by LIA (Innogenetics). A cut-off value of 42 U/ml was used. Anti-pepA and anti-pepB antibodies were scored.

Anti-CCP2 antibodies were detected by ELISA (Immunoscan LIA-ANA Update (Innogenetics, Ghent, Belgium) and by indirect immunofluorescence on Crithidia luciliae. RF was used.

ANC anti-CCP2 antibodies were found in 11/201 (5.5%) patients. Anti-CCP2 was considered positive, which corresponds to a specificity of 95.9%.

Anti-CCP2 antibodies were detected by ELISA (Immunoscan RA, mark 2, Eurodiagnostica, Arnhem, Netherlands). A cut-off value of 42 U/ml was used. Anti-pepA and anti-pepB antibodies were scored.

Anti-CCP2 antibodies were detected by a research LIA (Innogenetics). During each run, a strip was developed using a control serum, providing a cut-off intensity for each antigen line. In the control population mentioned earlier, all three ACPA had a specificity of at least 98.5%.

Anti-CCP2 antibodies were found in 11/201 (5.5%) patients, anti-pepA antibodies 3 (1.5%) patients, and anti-pepB antibodies in 5 (2.5%) patients.

Characteristics of ACPA positive patients with SLE

<table>
<thead>
<tr>
<th>Patient No</th>
<th>RF</th>
<th>Anti-CCP</th>
<th>Anti-PepA</th>
<th>Fine antinuclear reactivities</th>
<th>SE</th>
<th>Rx</th>
<th>RA crit</th>
<th>Clinical signs</th>
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<tr>
<td>1</td>
<td>1280</td>
<td>186</td>
<td>3+</td>
<td>3+ SSB, Ro60</td>
<td>0</td>
<td>–</td>
<td>–</td>
<td>Arthritis, proteinuria, leucopenia, lymphopenia</td>
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<tr>
<td>2</td>
<td>640</td>
<td>9</td>
<td>–</td>
<td>1+ RNP-C</td>
<td>0</td>
<td>–</td>
<td>+</td>
<td>Arthritis rash, photosensitivity, arthritis, lymphopenia</td>
</tr>
<tr>
<td>3</td>
<td>0</td>
<td>168</td>
<td>–</td>
<td>– dsDNA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>Arthritis rash, oral ulcers, arthritis, proteinuria, cellular casts</td>
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<tr>
<td>4</td>
<td>0</td>
<td>83</td>
<td>–</td>
<td>– SmB, dsDNA</td>
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<td>NA</td>
<td>NA</td>
<td>Arthritis rash, photosensitivity, oral ulcers, arthritis, proteinuria, cellular casts</td>
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<tr>
<td>5</td>
<td>80</td>
<td>2</td>
<td>1+</td>
<td>Histones, dsDNA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>Arthritis rash, arthritis, proteinuria, cellular casts</td>
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<tr>
<td>6</td>
<td>0</td>
<td>76</td>
<td>–</td>
<td>– SmB, RNP-A, RNP-C, ribosomal P, histones</td>
<td>1</td>
<td>NA</td>
<td>+</td>
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<tr>
<td>7</td>
<td>40</td>
<td>64</td>
<td>–</td>
<td>– SmD, SmB, RNP-C, RNP-1 70k, ribosomal P</td>
<td>NA</td>
<td>–</td>
<td>–</td>
<td>Arthritis rash, photosensitivity, pleuritis, arthritis, leucopenia</td>
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<td>8</td>
<td>0</td>
<td>58</td>
<td>–</td>
<td>– Negative</td>
<td>0</td>
<td>NA</td>
<td>–</td>
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<tr>
<td>9</td>
<td>320</td>
<td>110</td>
<td>–</td>
<td>– SmB, RNP-70k, RNP-A, RNP-C, RNP-1 70k, ribosomal P</td>
<td>0</td>
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<td>+</td>
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<td>10</td>
<td>320</td>
<td>78</td>
<td>1+</td>
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<td>11</td>
<td>80</td>
<td>56</td>
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<td>– RNP-A, histones, ribosomal P</td>
<td>1</td>
<td>–</td>
<td>–</td>
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<tr>
<td>12</td>
<td>640</td>
<td>52</td>
<td>–</td>
<td>– RNP-70k, RNP-A</td>
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<td>+</td>
<td>+</td>
<td>Arthritis rash, oral ulcers, arthritis, cellular casts, proteinuria, lymphopenia, leucopenia</td>
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<tr>
<td>13</td>
<td>320</td>
<td>&gt;1600</td>
<td>2+</td>
<td>2+ R600</td>
<td>0</td>
<td>+</td>
<td>+</td>
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</table>

RF titres and anti-CCP2 concentrations (U/ml, cut-off point 42 U/ml) are given. Anti-pepA and anti-pepB antibodies were scored.

6/10 evaluable patients, with 3/10 carrying an SE allele; radiographic erosions were present in 3/7 evaluable patients.

Our data suggest that the presence of ACPA does not exclude a diagnosis of SLE. It remains to be evaluated whether ACPA in SLE predispose for a chronic RA-like arthritis in this case.

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REFERENCES


Lack of efficacy of rituximab in Felty’s syndrome

C Sordet, J-E Gottenberg, B Hellmich, P Kieffer, X Mariette, J Sibilia

F elty’s syndrome (FS) is defined by the coexistence of rheumatoid arthritis (RA), neutropenia, and splenomegaly. The mechanisms underlying the neutropenia of FS may involve both cellular and humoral immunity, with a possible role of granulocyte-colony stimulating factor (G-CSF) antibodies.1 Various disease modifying antirheumatic drugs have been used to treat FS, but with varying success2 as anti-citrullinated peptide and anti-RA33 antibodies in distinguishing erosive arthritis in patients with systemic lupus erythematosus and rheumatoid arthritis. Ann Rheum Dis 2001; 60: 67–8.

METHODS AND RESULTS

Two men, were studied, aged 67 (patient 1) and 53 (patient 2) years, with a duration of RA of 6 and 11 years, respectively. FS had been diagnosed respectively 5 and 3 years ago, and RA remained active in both patients despite corticotherapy and respectively one (sulfasalazine) and two (methotrexate) previous disease modifying antirheumatic drugs. Anti-tumour necrosis factor treatment was not used because of neutropenia and the risk of severe infection. The absolute neutrophil count was persistently less than 0.8 × 10^9/L and complicated with recurrent sinopulmonary infections. There was no suggestion of congenital hypogammaglobulinemia and, in particular, no sign of selective IgG2 immunodeficiency. Blood and bone marrow immunophenotyping did not disclose any features of myelodysplasia or lymphoproliferation, or any large granular lymphocytes. No other classical cause of neutropenia, such as toxicity, chronic infection, vitamin deficiency, or liver disease, was present. Anti-G-CSF (IgG) antibodies, which were determined by enzyme linked immunosorbent assay (ELISA),1 were detected in one patient without previous administration of haematopoietic factor (G-CSF).

Owing to the presence of refractory RA associated with severe FS, rituximab was administered as an intravenous infusion at a dose of 375 mg/m² once weekly for 4 weeks. Concomitant treatment consisted of prednisone (15–20 mg/day) for more than 12 months in both patients and methotrexate (20 mg/week) since March 2003 in patient 2. The duration of follow up was 6 months. Rituximab was well tolerated. No major side effects were observed, with the exceptions of transient neutropenia (below 0.8 × 10^9/L) occurring in both patients in the first 3 weeks after the first infusion of rituximab and of severe infection in patient 1 during the first 6 weeks.

Table 1: Clinical and biological features of two patients with FS treated with rituximab

<table>
<thead>
<tr>
<th>Normal range</th>
<th>Neutrophil count</th>
<th>CRP (mg/L)</th>
<th>CD19+ cells</th>
<th>IgG</th>
<th>IgM</th>
<th>RF (IU/mL)</th>
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<tr>
<td>DAS28 &lt; 2.6</td>
<td>1800–7000/µL</td>
<td>4–9 mg/L</td>
<td>200–400/mm³</td>
<td>7.2–14.7 g/L</td>
<td>0.48–3.10 g/L</td>
<td>&lt;11 IU/mL</td>
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<tr>
<td>W0</td>
<td>6.64</td>
<td>460</td>
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<tr>
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<td>300</td>
<td>100</td>
<td>81.6</td>
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<tr>
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<td>360</td>
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<td>W3</td>
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<tr>
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<td>67</td>
<td>54.8</td>
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</tr>
<tr>
<td>W12</td>
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<td>170</td>
<td>65</td>
<td>38.2</td>
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<td>12.1</td>
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<tr>
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<td>6.5</td>
<td>150</td>
<td>55</td>
<td>25</td>
<td>2</td>
<td>11.5</td>
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<table>
<thead>
<tr>
<th>Patient 1</th>
<th>CRP (mg/L)</th>
<th>CD19+ cells</th>
<th>IgG</th>
<th>IgM</th>
<th>RF (IU/mL)</th>
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<tr>
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<td>11.2</td>
<td>2.63</td>
<td>12</td>
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<tr>
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<tr>
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<td>29.8</td>
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<td>ND</td>
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<td>0</td>
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<td>ND</td>
<td>ND</td>
<td>ND</td>
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<tr>
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<td>8.14</td>
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Table 1: Clinical and biological features of two patients with FS treated with rituximab.

DAS28, 28 joint count Disease Activity Score; ESR, erythrocyte sedimentation rate; CRP, C reactive protein; RF, rheumatoid factor, ND, not determined.
tolerated and efficiently controlled the clinical and biological activity of RA in patient 2, who fulfilled the American College of Rheumatology 50 response criteria and showed a marked decrease in serum levels of rheumatoid factor. However, results for FS were disappointing, because no increase in neutrophil count or modification of infection rates could be detected (table 1). In patient 1, a decrease in neutrophil count was observed at week 12, but without any clinical anomaly. Biological controls showed no modification of levels of anti-G-CSF antibodies, no appearance of anti-granulocyte antibodies, and no large granular lymphocyte proliferation.

DISCUSSION
Several factors might account for the lack of efficacy of rituximab in the treatment of FS. Firstly, although different autoreactive B cells may be involved in the pathology of FS, the inability of rituximab to bind to plasma cells, which are CD20 negative, might prevent it from acting on FS. Nevertheless, the efficacy of rituximab in certain conditions associated with autoantibodies is not correlated with a reduction of these antibodies, which would suggest that in addition to autoantibody production, other roles of B cells (immunoglobulins, antigen presentation, T cell cooperation) are important in the pathogenesis of such diseases. Secondly, a subpopulation of T lymphocytes having an antigranulocyte activity may exist independently of B cells in some forms of FS.

In conclusion, the lack of efficacy of rituximab in these two patients with FS raises some important questions about the mechanisms responsible for FS and the best therapeutic strategy to adopt.

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Antinuclear and antiphospholipid autoantibodies in patients with peripheral arterial occlusive disease

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According to the Chapel Hill Consensus Conference, large peripheral arteries are only affected by giant cell vasculitis and, in rare cases, by polyarteritis nodosa.1 2 Vasculitis becomes apparent through involvement of typical organs (lung, kidney, skin) or raised C reactive protein (CRP) level or erythrocyte sedimentation rate (ESR). Thus, a specific diagnostic effort to exclude vasculitides as an underlying disease in patients with peripheral arterial occlusive disease (PAOD) may be unnecessary. On the other hand, there is increasing evidence that humoral immunity may have a role in the pathogenesis of atherosclerosis.1 2 4 Antinuclear antibodies were reported in 70% of patients with severe coronary heart disease (CHD), compared with in only 17% in the control group. Thus, we prospectively studied the importance of autoantibody determination in patients with symptomatic PAOD.

METHODS AND RESULTS
Six hundred and ninety eight patients (mean (SD) age 68 (10) years) referred for treatment of PAOD between 1998 and 1999 were included. In 121 patients with PAOD (aged 61 (12) years) with a low atherosclerotic risk profile, or with rarefied distal arteries without media calcinosis, or with raised ESR or CRP not due to a local infection, the following autoantibodies were determined: antinuclear antibodies (ANA) by an indirect immunofluorescence technique; antibodies against extractable nuclear antigens (Scl-70, RNP, SSA, SSB, Jo-1, SM) by western blot; double stranded DNA antibodies, antineutrophil cytoplasmic antibodies (c- and pANCA), and antiphospholipid antibodies (cardiolipin, phosphatidylserine (APSA), and β2-glycoprotein) by enzyme linked immunoassay. To stratify the importance of autoantibody determination all patients with increased autoantibody concentration were clinically and sonographically followed up for 24 (6) months for evidence of vasculitides or collagen disease. A multivariate logistic regression analysis was performed to evaluate the importance of CRP and ESR in patients with autoantibody concentrations above the appropriate reference value.

Thirty eight of the 121 patients had increased autoantibody concentrations (table 1). ANA were the most common autoantibodies detected in 14 patients followed by APSA in 11, and β2-glycoprotein antibodies in 12. Patients with increased autoantibody concentration did not differ in their PAOD stages and affected segments, but in patients with increased autoantibody concentrations the ESR was higher (p = 0.0043). The ESR at 2 hours was associated with an odds ratio of 7.1 (95% confidence interval 1.5 to 33.8) in determination of increased autoantibody concentrations. During the follow up of 24 (6) months no vasculitides or collagen diseases could be detected by clinical examination or by nailfold capillary microscopy, pulmonary or gastrointestinal imaging in the 38 patients.

DISCUSSION
The group of 121 patients with PAOD analysed is a group selected individually from all the patients, but represents those patients in whom possible vasculitis may be present.
Raised concentrations of autoantibodies were common in the patients investigated. Increased autoantibody concentrations significantly correlated with a raised ESR.

As far as we know, our data are the first to report the results of autoantibody determinations in a large group of patients with PAOD. In contrast with the high rate of ANA in patients with coronary atherosclerosis, the prevalence of ANA in our patients with PAOD was much lower. Whether this difference in the prevalence of ANA was due to different forms of atherosclerosis, or due to specific differences in coronary and peripheral manifestations can only be speculated.

In agreement with the coronary studies, no association of the determined autoantibodies with classical risk factors was found. The higher ESR in the patients with increased autoantibody concentrations might be associated with a higher degree of inflammatory activity of the atherosclerosis. Antiphospholipid and \( \beta_2 \)-glycoprotein antibodies, which are most relevant in association with atherosclerosis, did not seem to lead to a prognosis, but antibody determination in larger groups of non-selected patients is desirable.

### Table 1 Characteristics of patients

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<tr>
<th>Characteristics</th>
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<th>Autoantibody determination</th>
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</thead>
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<td>Number</td>
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<td>83</td>
</tr>
<tr>
<td>Age (years), mean (SD)</td>
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<td>61 (12)</td>
</tr>
<tr>
<td>PAOD:</td>
<td></td>
<td></td>
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<tr>
<td>Stage II</td>
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<td>63</td>
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<tr>
<td>Segment of vascular lesions:</td>
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<td>Crural</td>
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<tr>
<td>Femoral</td>
<td>41</td>
<td>30</td>
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<tr>
<td>Iliacal</td>
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<td>5</td>
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<tr>
<td>Combined</td>
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<td>43</td>
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<tr>
<td>Risk factors:</td>
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<td></td>
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<tr>
<td>Diabetes mellitus</td>
<td>37</td>
<td>26</td>
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<tr>
<td>Hypertension</td>
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<td>39</td>
</tr>
<tr>
<td>Dyslipoproteinaemia</td>
<td>66</td>
<td>41</td>
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<tr>
<td>Nicotine abuse</td>
<td>75</td>
<td>27</td>
</tr>
<tr>
<td>Markers of inflammation</td>
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<tr>
<td>ESR &gt; 20 mm/1st h</td>
<td>31</td>
<td>29</td>
</tr>
<tr>
<td>ESR &gt; 40 mm/2nd h</td>
<td>32</td>
<td>30</td>
</tr>
<tr>
<td>CRP &gt; 10 mg/l</td>
<td>20</td>
<td>37</td>
</tr>
<tr>
<td>History of CHD</td>
<td>47</td>
<td>17</td>
</tr>
</tbody>
</table>

Variables are expressed as a percentage of the number of patients in each group. *p = 0.0043 v “no measured autoantibodies”.

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### REFERENCES


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Incidence of primary systemic vasculitides in Vilnius: a university hospital population based study

J Dadoniene, G Kirdaite, Z Mackiewicz, A Rimkevicius, G Haugeberg


As knowledge of the epidemiology of primary systemic vasculitides (PSV) is fragmentary, we attempted to investigate the incidence of temporal arteritis (TA), Takayasu’s arteritis (TAA), polyarteritis nodosa (PAN), Wegener’s granulomatosis (WG), Churg-Strauss syndrome (CSS), Henoch-Schönlein purpura (HSP), and hypersensitivity vasculitis (HSV) in Vilnius according to the American College of Rheumatology (ACR) 1990 criteria and to compare the data with the results from selected European studies.¹

To be included in this study the patients had to (a) have been diagnosed with systemic vasculitides in the 10 year period from 1990 to 1999 and (b) have been resident in Vilnius at the time of diagnosis. Patients referred to Vilnius University Hospital rheumatology department were prospectively included in the study. Also, the patients’ registration books from tertiary nephrology, dermatology, and internal medicine departments were searched for a diagnosis of PSV retrospectively, but following the same inclusion criteria. Additionally, the data files of the centre of pathology available from 1995 onwards, including renal register, were searched.

We applied ACR 1990 criteria for the classification of PSV; however, for patients classified as HSV, the term cutaneous leukocytoclastic vasculitis (LCV) was equally used.² Patients with microscopic polyangiitis (MPA) were included in the PAN group, and PAN criteria applied to both conditions. The group of PAN and HSV/LCV were reanalysed according the definition for MPA. The denominator population was the adult population over 16 years from Vilnius city, which comprised 468 504 people (53.7% female) in 1999.

### Table 1

Number and annual incidence (per million inhabitants) of primary systemic vasculitides in Vilnius compared with the selected European studies conducted during the past decade

<table>
<thead>
<tr>
<th></th>
<th></th>
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</thead>
<tbody>
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<td>Giant (temporal arteritis) cell arteritis</td>
<td></td>
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<tr>
<td>Number of patients</td>
<td>11</td>
<td>24</td>
<td>–</td>
<td>110</td>
</tr>
<tr>
<td>Median age in years (range)</td>
<td>70 (50–87)</td>
<td>2.3/10⁶*</td>
<td>31.9/10⁶</td>
<td>45.6/10⁶</td>
</tr>
<tr>
<td>Overall annual incidence</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Takayasu arteritis</td>
<td>6</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Number of patients</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Median age in years (range)</td>
<td>40 (16–49)</td>
<td>1.3/10⁶</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Overall annual incidence</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Polyarteritis nodosa</td>
<td>36</td>
<td>5</td>
<td>33†</td>
<td>13</td>
</tr>
<tr>
<td>Number of patients</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Median age in years (range)</td>
<td>48 (17–72)</td>
<td>7.7/10⁶</td>
<td>6.7/10⁶</td>
<td>8.0/10⁶</td>
</tr>
<tr>
<td>Overall annual incidence</td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hypersensitivity vasculitis/cutaneous leukocytoclastic angiitis</td>
<td>122</td>
<td>45 (17–83)</td>
<td>26.0/10⁶</td>
<td>2.7/10⁹</td>
</tr>
<tr>
<td>Number of patients</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Median age in years (range)</td>
<td>26.0/10⁶</td>
<td>2.7/10⁹</td>
<td>17.8/10⁶</td>
<td>29.7/10⁶</td>
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<tr>
<td>Overall annual incidence</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Henoch-Schönlein purpura</td>
<td>14</td>
<td>3</td>
<td>3</td>
<td>27</td>
</tr>
<tr>
<td>Number of patients</td>
<td></td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Median age in years (range)</td>
<td>18 (16–21)</td>
<td>3.0/10⁵</td>
<td>6.7/10⁶</td>
<td>1.2/10⁶</td>
</tr>
<tr>
<td>Overall annual incidence</td>
<td></td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Churg-Strauss syndrome</td>
<td>6</td>
<td>2</td>
<td>6</td>
<td>2</td>
</tr>
<tr>
<td>Number of patients</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Median age in years (range)</td>
<td>43 (27–58)</td>
<td>1.3/10⁶</td>
<td>2.7/10⁶</td>
<td>2.4/10⁶</td>
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<tr>
<td>Overall annual incidence</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wegener’s granulomatosis</td>
<td>10</td>
<td>5</td>
<td>21</td>
<td>9</td>
</tr>
<tr>
<td>Number of patients</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Median age in years (range)</td>
<td>57 (22–68)</td>
<td>2.1/10⁶</td>
<td>6.7/10⁶</td>
<td>8.5/10⁶</td>
</tr>
<tr>
<td>Overall annual incidence</td>
<td></td>
<td></td>
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<td></td>
</tr>
</tbody>
</table>

*7.2/million (95% CI 3.8 to 13.3) for the population aged 50 years and older; †overall, 150 000 adult inhabitants in the Kristiansand study, 414 500 in the Norwich study, 250 000 in the Lugo study, 468 504 in Vilnius refer to the study period; ‡polyarteritis nodosa and hypersensitivity vasculitis in this study are reported for the period from 1988 to 1997 and from 1990 to 1994, respectively.
Overall, we identified 205 patients according to inclusion criteria—an annual incidence of 43.8/10^6 (95% confidence interval (CI) 38.1 to 50.3) (table 1). The most common type of vasculitis was HSV/LCV with an annual incidence of 26.0/10^6 (95% CI 21.7 to 31.2). The incidence of PAN was found to be 7.7/10^6 (95% CI 5.5 to 10.7), HSP 3.0/10^6 (95% CI 1.7 to 5.1), TA 2.3/10^6 (95% CI 1.2 to 4.3), WG 2.1/10^6 (95% CI 1.1 to 4.1), TAA 1.3/10^6 (95% CI 0.5 to 2.9), and CSS 1.3/10^6 (95% CI 0.5 to 2.9) annually. Six patients in the PAN group and eight in the HCV/LCV group responded to the definition of MPA being antineutrophil cytoplasmic antibody (ANCA) positive and/or having nephritis in addition to other system involvement. Therefore, the annual incidence of presumed MPA was 3.0/10^6 (95% CI 2.0 to 5.7) in total. The diagnoses of 66/205 patients were supported by biopsy data. Five of 36 patients with PAN, 8/10 patients with WG, and 4/5 with CSS were found to be ANCA positive.

Three studies, Kristiansand (Norway), 1 Norwich (Norfolk, England), 2 and Lugo (Spain) 3 were selected for comparison with our study (table 1). The annual incidence of PSV in Vilnius seems to fall in between the figures of annual incidence reported in Norwich (38.6/10^6, TA excluded), Kristiansand (54.5/10^6), and Lugo (115.0/10^6). However, the distribution of the annual incidence of distinct vasculitides differs from those of other European studies. The most important difference was noted for TA and less notably for WG (table 1). The annual incidence of MPA was in accordance with the lower figures reported in the European studies and less than half that quoted in the study by Watts et al.4

The shorter life expectancy of Lithuanian people, which in 1999 was 71 years and lower than that of the European population, might be a potential explanatory factor for the lower incidence of TA in Vilnius. Possibly, because a histological examination was rarely carried out, and the ANCA test was introduced only after 1995,5 WG and other ANCA associated vasculitides cases are underrepresented, especially in the first 5 years of this study.

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REFERENCES


Bone mineral density in patients with rheumatoid arthritis treated with infliximab

M Vis, A E Voskuyl, G J Wolbink, B A C Dijkmans, W F Lems for the OSTRA study group

Methods and results

This open cohort study consisted of consecutive patients with RA, who were treated with infliximab in the Slotervaart Hospital and the VU University Medical Centre. All patients fulfilled the ACR 1987 criteria of RA and had active disease (defined by the modified 28 joint count Disease Activity Score (DAS28) of at least 3.2). Infliximab was given intravenously at 0, 2, 6, 14, weeks and from the fourth infusion every 8 weeks in a dose of 3 mg/kg. At each visit the DAS28 was calculated and changes in drug treatment were recorded. BMD measurements (g/cm²) of the hip (total hip) and lumbar spine (L1–4) were performed at baseline and after 1 year on a Hologic 4500.

OSTRA: The Dutch Study Group for Observational Risk Assessment in Rheumatoid Arthritis.
In total, 36 patients (29 (81%) female) were included into the study. Patients had a mean (SD) age of 53 (12) years, with a median (range) disease duration of 9.5 years (0–49). Methotrexate, prednisone, and bisphophonates were used by 100%, 50%, and 25% of the patients, respectively. The mean disease activity (DAS28) decreased from 5.6 at baseline to 3.8 at 6 weeks and stabilised around 3.6 for the rest of the studied period. In 36 patients dual x-ray absorptiometry (DXA) measurements of lumbar spine (L1–4) and in 30 patients DXA measurements of the hip were available at baseline and after 1 year. In four patients no DXA hip measurements were available because of bilateral hip replacement, and in two patients only one DXA of the hip was available owing to unknown causes.

Mean (SD) BMD at the lumbar spine increased nonsignificantly from 0.998 (0.205) to 1.001 (0.199) at 1 year (+1.1%, p = 0.117). BMD at the total hip decreased nonsignificantly from 0.857 (0.144) to 0.854 (0.132) at 1 year. (−0.3%, p = 0.683). In a linear regression model, changes in BMD at the hip or the spine were not associated with mean BMD at the lumbar spine increased non-

**DISCUSSION**

This study indicates that BMD of the spine has a tendency to increase and that BMD of the hip slightly decreases during 1 year of treatment with infliximab. This is in contrast with previous longitudinal studies of patients with RA, in which a decrease of BMD was seen during conventional disease modifying antirheumatic drug treatment without tumour necrosis factor blockings agents (table 1).

In our view, these data suggest that treatment with infliximab can arrest generalised osteoporosis in patients with RA. This view is supported by the observation that markers of bone formation increased and markers of bone resorption decreased in the first 6 weeks of treatment with infliximab.7 We do realise that our observations are made in an open cohort study and therefore no definite conclusions can be drawn from our data.

In summary, this study suggests that treatment with infliximab has a positive effect on BMD in patients with RA. Because patients with RA have an increased risk of bone loss and, subsequently, osteoporotic fractures, this might be an additional advantage of infliximab (above the well known favourable effect on disease activity and radiological damage), and warrants further study.

**ACKNOWLEDGEMENTS**

We thank the members of the OSTAR group: TK Kvien, G Haugeberg, T Uhlig, EA Haavardsholm (Oslo, Norway), and A Woolf (Truro, UK), for their valuable comments.

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**REFERENCES**


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**Table 1** Summary of changes in BMD of five studies in patients with RA

<table>
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<tr>
<th>Study</th>
<th>Patients (n)</th>
<th>RA</th>
<th>Follow up (years)</th>
<th>BMD change (%)</th>
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</thead>
<tbody>
<tr>
<td>Boers*+</td>
<td>62</td>
<td>Early</td>
<td>1</td>
<td>−1.3</td>
</tr>
<tr>
<td>Gough+</td>
<td>50</td>
<td>Early</td>
<td>1</td>
<td>−4.2</td>
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<tr>
<td>Haugeberg+</td>
<td>366</td>
<td>Established</td>
<td>2</td>
<td>−0.77</td>
</tr>
<tr>
<td>Dolan+</td>
<td>21</td>
<td>Established</td>
<td>2</td>
<td>0.0*</td>
</tr>
<tr>
<td>Shibuya+</td>
<td>146</td>
<td>Established</td>
<td>1</td>
<td>No data*</td>
</tr>
<tr>
<td>This study</td>
<td>36</td>
<td>Established</td>
<td>1</td>
<td>−0.3</td>
</tr>
</tbody>
</table>

* Percentage change calculated from data given in the manuscript; BMD of the spine was not measured; data from patients treated with sulphasalazine alone.
An open study of pulse pamidronate treatment in severe ankylosing spondylitis, and its effect on biochemical markers of bone turnover

A P Cairns, S A Wright, A J Taggart, S M Coward, G D Wright

Osteoporosis is a common feature of ankylosing spondylitis (AS), and vertebral fractures are an increasingly recognised complication. A cumulative fracture prevalence of between 9.5% and 18% has been reported, with a six- to eightfold relative increased risk of vertebral fracture.1,2 Osteoporosis and new bone formation (syndesmophytosis) suggest that disordered bone turnover has a role in disease pathogenesis in AS. Bisphosphonates accumulate at sites of increased bone turnover, and inhibit bone resorption by inducing osteoclast apoptosis,3 thereby improving bone density and reducing fracture rates.2 Pulse pamidronate has recently been used with clinical efficacy in the treatment of AS.4 Biochemical markers of bone turnover have been used to monitor response to treatment in postmenopausal osteoporosis.6 The effect of bisphosphonate treatment on biochemical bone turnover markers has not previously been studied in AS. We aimed at studying the efficacy of pulse pamidronate treatment in severe AS, and at determining its effect on biochemical bone turnover markers.

METHODS AND RESULTS

Patients with severe AS were treated with 6 monthly intravenous pulses of pamidronate, receiving 30 mg for the first infusion and 60 mg subsequently. The Bath AS Disease Activity Index (BASDAI) and the Bath AS Metrology Index (BASMI) scores were recorded, and C reactive protein (CRP) and erythrocyte sedimentation rate (ESR) measured at each visit. Fifteen patients participated (13 male, mean age 44). Mean disease duration was 14.8 years. All patients were receiving non-steroidal anti-inflammatory drugs; two were also receiving sulfasalazine, one methotrexate, and one azathioprine. No patients were currently receiving an oral bisphosphonate. Four patients had a history of uveitis, two psoriasis, two inflammatory bowel disease, and eight peripheral arthritis.

Fasting blood samples were taken monthly for measurement of biochemical bone turnover markers, according to the manufacturers’ instructions. Degradation products of type I collagen C-terminal telopeptides were measured with the serum crosslaps enzyme linked immunosorbent assay (ELISA; Nordic Bioscience Diagnostics A/S, Herlev, Denmark). Serum bone GLA protein was measured using the N-MID osteocalcin ELISA kit (Nordic Bioscience Diagnostics A/S, Herlev, Denmark). Bone-specific alkaline phosphatase was measured with the Access Ostase assay (Beckman Coulter Inc, Fullerton, California, USA). Non-parametric analyses (Wilcoxon signed rank tests) on an intention to treat basis were used to analyse the data.

Three patients did not complete the study (arthralgia/headache, back pain/nausea, and myalgia, respectively). The mean total dose of pamidronate received was 277 mg. The initial median serum crosslaps was 1845.0 ng/ml. Mean (SD) population serum crosslaps concentrations are 506 (255) ng/l for postmenopausal women, 321 (155) ng/l for premenopausal women, and 332 (190) ng/l for men. The initial median osteocalcin concentration was 12.4 µg/l. Mean (SD) population osteocalcin concentrations are 123 (43) µg/l for men, 8.7 (2.9) µg/l for premenopausal women, and 13.2 (4.7) µg/l for postmenopausal women. The initial median osteocalcin concentration was 19.5 ng/ml. Mean (SD) population osteocalcin concentrations are 17.9 (6.5) ng/ml for premenopausal women, 28.4 (9.5) ng/ml for postmenopausal women, and 21.4 (9.1) ng/ml for men.

Median serum crosslaps fell from 1845.0 to 556.5 ng/l (Z = −3.29, p = 0.001) (fig 1). Median serum osteocalcin fell from 19.5 to 16.2 ng/ml (Z = −2.34, p = 0.02). Median serum osteoprotegerin fell from 12.4 to 9.6 µg/l (Z = −3.11, p = 0.002). The median BASDAI improved from 6.80 to 5.75 (Z = −1.98, p = 0.048), but there was no significant improvement in the median BASMI (initial 8.00 v final 7.50, Z = −1.64, p = 0.10). There were non-significant trends of reduction in median CRP (initial 36.4 v final 26.6 mg/l, Z = −1.89, p = 0.06) and median ESR (initial 38 v final 29 mm/1h, Z = −0.71, p = 0.48). There were no significant correlations between clinical measures and bone turnover markers.

DISCUSSION

Pulse pamidronate treatment significantly reduced all three biochemical bone turnover markers. This was particularly marked for serum crosslaps, the bone resorptive marker, where there was a 69.8% relative reduction. Such marked reduction of the crosslaps concentration suggests that bisphosphonates have a role in the management of the osteoporosis of AS. Ultimately, studies assessing fracture rates in patients with AS receiving bisphosphonates would be of great interest. One possibility is that by reducing the rate of new bone formation, bisphosphonates may also reduce syndesmophyte formation. However, longer term controlled...
studies in patients with early disease are needed to examine this subject.

Pulse pamidronate treatment also had a small beneficial effect on disease activity as measured by the BASDAI, which has also been demonstrated in a recent randomised controlled study of pamidronate treatment in AS. The patients in our study had established, severe disease. All but one would have met recent ASAS criteria for the use of anti-tumour necrosis factor drugs in AS. Clinical studies in earlier, less severe disease are required to determine if bisphosphonates may be of benefit to these patients.

**REFERENCES**


**Association between interleukin 6 gene polymorphisms and Behçet’s disease in Korean people**

H K Chang, W C Jang, S B Park, S M Han, Y H Nam, S S Lee, J U Kim, H S Lee

Interleukin (IL) 6 is an important mediator of inflammatory and immune responses, and IL6 gene polymorphisms are known to play a part in chronic inflammatory and autoimmune disorders. Increased IL6 plasma levels and enhanced IL6 mRNA expression have been found in patients with active Behçet’s disease. Therefore, this study aimed at investigating the associations between Behçet’s disease in Korean people and two functional IL6 gene polymorphisms—namely, a single nucleotide polymorphism at strand repeat polymorphisms in the 3’ flanking region of the IL6 gene (IL6*vntr*) and a variable number of tandem repeat polymorphisms in the 3’ flanking region of the IL6 gene (IL6*vntr*). The terms IL6*vntr* and IL6*vntr* were designated as in a previous study.

**METHODS AND RESULTS**

The study group included 89 Korean patients with Behçet’s disease (36 men, 53 women; mean (SD) age 39.1 (8.5)), fulfilling the International Study Group criteria, and 123 controls (47 men, 76 women; mean (SD) age 43.1 (13.4)). The cumulative history of severe manifestations was investigated during the disease course. Analyses of IL6*vntr* and IL6*vntr* were carried out in all the subjects by polymerase chain reaction (PCR)-restriction fragment length polymorphism and PCR genotyping, respectively. Significance was evaluated using Fisher’s exact test or t test and defined as p < 0.05: p values with Bonferroni’s correction (Pcorr) were calculated in certain cases. Haplotype and linkage disequilibrium (LD) analyses were assessed using the estimated haplotype (EH) programme.

There was no evidence of genetic association conferred by the IL6*vntr* polymorphism. In the case of the IL6*vntr* genotypes and alleles were identified with the following frequencies in the controls: AB, 2 (1.6%); BB, 117 (95.1%); BC, 3 (2.4%); CC, 1 (0.8); and in the patients with Behçet’s disease: BB, 77 (86.5%); BC, 12 (13.5%). Because the majority (98.6%) of the subjects had one of the two common genotypes (BB and BC), comparisons between the groups were made with these major genotypes and alleles. There were significant differences in the frequencies of the IL6*vntr* and alleles between patients with Behçet’s disease and controls (genotypes: p = 0.005, Pcorr = 0.01; alleles: p = 0.02, Pcorr = 0.044) (table 1). The odds ratio for Behçet’s disease associated with the C allele of IL6*vntr* (IL6*vntr*C) was 3.5 (95% confidence interval 1.2 to 10.0).

When the studied subjects were stratified according to the results of HLA-B51 testing, significant differences in the IL6*vntr* genotype and allele frequencies were found only in the HLA-B51 negative subjects (genotypes: p = 0.005, Pcorr = 0.01; alleles: p = 0.02, Pcorr = 0.04). Using the EH programme, the distribution of haplotypes between patients with Behçet’s disease and controls differed significantly only in those with the IL6*vntr* G/IL6*vntr*C haplotype (p = 0.001, Pcorr = 0.004): the odds ratio for Behçet’s disease in the subjects with this haplotype was 7.3 (95% confidence interval 1.6 to 32.9). In addition, the EH programme revealed a D value of 0.08, suggesting the presence of LD at low level between the two polymorphic sites.
No significant associations were found between the genotypes of the two IL6 polymorphisms (IL6prom and IL6vntr) and clinical variables, including disease duration, mean age at onset, clinical manifestations, severe manifestations, and HLA-B51 positivity, in patients with Behçet’s disease (all p>0.05). However, the distribution of the IL6vntr genotype differed significantly between male and female patients, and the frequency of the BC genotype was much higher in female patients with Behçet’s disease than in male patients (BB: male, 45.5% v female, 54.5%; BC: male, 8.3% v female, 91.7%; p = 0.024, \( p_{corr} = 0.048 \)).

DISCUSSION

Our data are consistent with previous investigations showing considerable interethnic variability in the distribution of the IL6prom and IL6vntr genotypes.\(^1\) There was no evidence for genetic association conferred by the IL6prom polymorphism, whereas significant differences in the IL6vntr genotype and allele frequencies were found between patients with Behçet’s disease and controls. These differences were particularly apparent in the HLA-B51 negative subjects or female patients. In addition, susceptibility to Behçet’s disease was increased significantly in subjects carrying the IL6vntr*C allele and the IL6prom*G/IL6vntr*C haplotype. To confirm these findings, further investigations are required in other ethnic populations.

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REFERENCES


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