Severe septicaemia in a patient with polychondritis and Sweet’s syndrome after initiation of treatment with infliximab

F G Matzkies, B Manger, M Schmitt-Haendle, T Nagel, H-G Kraetsch, J R Kalden, H Schulze-Koops


R D Sweet first described an acute febrile neutrophilic dermatosis in 1964 characterised by acute onset, fever, leucocytosis, and erythematous plaques. Skin biopsy specimens show infiltrates consisting of mononuclear cells and neutrophils with leucocytoclasis, but without signs of vasculitis. Sweet’s syndrome is frequently associated with solid malignancies or haemoproliferative disorders, but associations with chronic autoimmune connective tissue disorders have also been reported. The aetiology of Sweet’s syndrome is unknown, but evidence suggests that an immunological reaction of unknown specificity is the underlying mechanism.

CASE REPORT

A 51 year old white man with relapsing polychondritis (first diagnosed in 1997) was admitted to our hospital in June 2001 with a five week history of general malaise, fever, recurrent arthritis, and complaints of morning stiffness. Besides autoimmune polychondritis, he had insulin dependent diabetes mellitus that was diagnosed in 1989.

On admission, he presented with multiple small to medium, sharply demarked, raised erythematous plaques on both forearms and lower legs, multiple acne-like pustules on the face, neck, and chest, two abscesses on both thighs, and paronychia of several fingers. Microbiological examinations of the abscesses showed that they were sterile, and no bacterial or viral DNA was found in the acne-like lesions. *Staphylococcus aureus* was isolated from the finger paronychia. Laboratory testing showed a white blood cell count of 3.1×10^9/l with 40% lymphocytes and 46% neutrophils, a C reactive protein of 0.21 g/l and maximally raised erythrocyte sedimentation rate. Surgical wound debridement was performed on the fingers and, because of immunosuppressive treatment (glucocorticoids, methotrexate, and azathioprine), systemic antibiotic treatment was started even in the absence of detectable systemic infection. Several days after admission, the patient developed an arthritis flare. Multiple skin biopsy samples were taken and showed typical features of Sweet’s syndrome without signs of bacterial or viral infection.

As the patient had developed Sweet’s syndrome while receiving immunosuppression and the underlying immunological activity of his polychondritis appeared to be insufficiently controlled, azathioprine was stopped and, in the absence of detectable infections, infliximab was used in an attempt to suppress the continuous (auto)immune reactivity.

Infliximab was given at 3 mg/kg body weight. Arthritis and morning stiffness rapidly resolved. The skin lesions disappeared and no new skin lesions developed. However, 14 days after the application, the patient developed fever of up to 40°C and new erythematous plaques, similar in appearance and location to the original plaques (fig 1). As at first admission, an infection was ruled out by intensive clinical, laboratory, microbiological, and radiological tests. Consequently, a higher dose of glucocorticoids (80 mg) and a second application of infliximab (3 mg/kg body weight) were given. The erythematous rash rapidly resolved and the patient was discharged from the hospital in apparently good health.

Eleven days after the second treatment with infliximab, the patient presented with myalgias, subfebrile temperatures, and general malaise. A parasternal abscess with connection into the mediastinum and new multiple pulmonal round formations were detected by computed tomography scan. Subsequently, the patient developed multiple abscesses on the right elbow and both feet. Penicillin resistant *Staphylococcus aureus* was isolated from the parasternal abscess. Despite systemic antibiotic treatment and surgical incisions, the patient deteriorated, developed pneumonia and rapidly met the criteria of septicaemia with acute renal and respiratory failure. Despite continuous aggressive wide range antibiotic and antmycotic treatment and maximum intensive care, he died of multiorgan failure as a consequence of progressive septicaemia. Shortly before his death, 11 weeks after the second infusion of infliximab, the typical Sweet’s syndrome skin lesions reappeared.

DISCUSSION

Neutralising tumour necrosis factor α (TNFα) has been employed as a powerful anti-inflammatory principle in patients with rheumatoid arthritis and other rheumatic diseases such as Still’s disease or giant cell arteritis. After several immunosuppressive drugs alone or in combination had failed to control immunological activity in our patient, infliximab was used and the clinical symptoms rapidly improved, leading to complete resolution of the arthritis, morning stiffness, and skin lesions. However, the case of our
patient dramatically underlines the risk of infectious complications after neutralising TNFα that might be particularly important in patients with a compromised immune system as a consequence of immunosuppressive drugs and/or diseases favouring infectious diseases, such as diabetes, as was the case here. The unfortunate course of our patient should alert rheumatologists to employ reagents that neutralise TNFα with extreme caution in patients who are more susceptible to infections because of accompanying diseases and/or concomitant immunosuppressive treatment.

Adrenomedullin in synovial fluids from patients with rheumatoid arthritis inhibits interleukin 6 production from synoviocytes

Y Nanke, S Kotake, K Yonemoto, S Saito, T Tomatsu, N Kamatani

Adrenomedullin (AM) is a hypotensive peptide found in human pheochromocytoma tissue, which comprises 52 amino acids with an intramolecular disulphide bond.1,2 The ring structure and amidated C-terminus of AM are critical for its receptor binding and hypotensive activity. The mature AM is synthesised as glycine extended AM followed by C-terminal amidation to assume a biologically active form in tissues. AM has a vasorelaxant effect, antagonising the vasoconstrictor effect of endothelin-1 (ET-1). Recently, proinflammatory cytokines, such as tumour necrosis factors α (TNFα) and interleukin-1 (IL1), were found to stimulate production and secretion of AM from vascular endothelial cells and vascular smooth muscle cells in vitro, suggesting that AM interacts with the immune system.3 However, AM reduces the production of TNFα from macrophages stimulated with lipopolysaccharide. In addition, AM shows an anti-inflammatory effect that reduces the production of the IL8 family by macrophages.4 We recently reported that the concentration of AM is raised in plasma from patients with systemic sclerosis complicated by pulmonary hypertension.5 Rheumatoid arthritis (RA) is a chronic inflammatory disease of unknown cause. Inflammatory cells and cytokines such as IL1, IL6, TNFα, and IL17 are responsible, at least in part, for the pathological immune response in RA.6 Thus, we suggested that AM may play a part in the pathogenesis of RA.

Synovial fluids were obtained from nine patients with RA,7 and from six patients with osteoarthritis (OA). The concentrations of total and mature AM were measured by immunoradiometric assay. The level of ET-1 was measured by radioimmunoassay. For the immunohistochemical studies, synovial tissue was obtained from the knees of three patients with RA and three with OA and stained using antihuman AM antibody and antihuman ET-1 antibody.

To explore the effect of AM on the production of IL6 from RA synoviocytes, the synovial cells obtained from three patients with RA were cultured for eight days and AM was added at various concentrations for three days. The level of IL6 in the supernatant was measured by an enzyme immunoassay.

The concentration of total AM in synovial fluid (mean (SD); pg/ml) was significantly higher in patients with RA (31.4 (14.7) pg/ml) than in patients with OA (5.5 (1.7) pg/ml) (p=0.001) (web extra fig W1). The levels of mature AM were also higher in patients with RA (3.7 (2.1) fmol/l) than in patients with OA (1.1 (0.2) fmol/l) (p=0.01) (fig 1). There was no significant difference between the level of ET-1 in synovial fluids from patients with RA and OA (data not shown).

Figure 1 The concentration of mature AM was higher in patients with RA than in those with OA (p=0.01). Synovial fluids were obtained from nine patients with RA and six patients with OA.
AM and ET-1 were positive around the perivascular and endothelial cells in the synovial tissue from patients with RA (fig 2, web extra fig W2). In contrast, the synovial tissue from patients with OA was negative. AM reduced constitutive production of IL6 from RA synoviocytes dose dependently. A high concentration of AM (>10^−9 mmol/l) significantly reduced constitutive production of IL6 compared with a low concentration of AM (≤10^−9 mmol/l) (p=0.0029). TNFα dose dependently induced production of IL6 from RA synoviocytes (data not shown). AM did not reduce IL6 production induced by TNFα (data not shown).

Our study showed that the concentration of total and mature AM in synovial fluids was significantly higher in patients with RA than in patients with OA. In addition, by immunohistochemical staining, AM and ET-1 were shown to be positive around the perivascular area, the endothelial cells, and synoviocytes in RA synovial tissue. We have reported that osteoclasts are present in synovial tissues from patients with RA and that IL6 and soluble IL6 receptors in synovial fluids from patients with RA and synoviocytes in RA synovial tissue. We have reported that osteoclasts are present in synovial tissues from patients with RA and that IL6 and soluble IL6 receptors in synovial fluids may participate in osteoclast formation. Thus, IL6 is responsible for joint destruction in the presence of soluble IL6 receptor through osteoclastogenesis. In this study we showed that AM reduced constitutive production of IL6 from RA synoviocytes dose dependently. Thus, our results suggest that AM in patients with RA inhibits both synovitis and osteoclastogenesis through the inhibition of IL6 production.

Additional figures can be found on the website at www.annrheumdis.com

Authors’ affiliations
Y Nanke, S Kotake, K Yonemoto, S Saito, T Tomatsu, N Kamatani, Institute of Rheumatology, Tokyo Women’s Medical University, 10-22 Kawada-cho, Shinjuku-ku, Tokyo 162-0054, Japan

Correspondence to: Dr Y Nanke; ynn@ior.twmu.ac.jp

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REFERENCES


Abnormal IgA levels in patients with rheumatoid arthritis

L J Badcock, S Clarke, P W Jones, P T Dawes, D L Mattey

The dominant antibody at mucous membranes and in exocrine secretions is IgA. It has been implicated in the pathogenesis of rheumatoid arthritis (RA), possibly due to immune complex formation. If IgA is important in RA pathogenesis one might predict that patients with abnormal levels would have different characteristics from the “normal” IgA population. Limited work published on patients with high IgA levels has suggested that there is an increase in erythrocyte sedimentation rate (ESR), microscopic haematuria, and both distal interphalangeal joint involvement and unilateral sacroiliitis, even though patients fulfil the American College of Rheumatology (ACR) criteria for RA and have no other evidence of spondyloarthropathy.

Primary selective IgA deficiency is the most common hypogammaglobulinaemia in the general population, with a prevalence of around 1:500. It is associated with increased risk of autoimmune disease and, possibly, with RA. Primary IgA deficiency may result from impaired switching from class IgM to IgA. Secondary IgA deficiency may be caused by drugs such as n-penicillamine, sulfasalazine, and gold. The few descriptions of primary IgA deficiency and RA have been single case studies and a longitudinal study is needed to determine if these cases represent a subgroup.

METHODS AND RESULTS

Serum immunoglobulins were measured in 352 patients (aged 18–75) attending a rheumatology outpatient department over a six year period. All patients fulfilled the ACR criteria for diagnosis of RA. Patients with selective hypogammaglobulinaemia (≥240 IU/ml) or primary selective IgA deficiency (<50 IU/ml) were identified as the two study cohorts. These
were compared with patients with RA (n=277) with normal IgA levels. No patients had been treated with immunosuppres- 
sant drugs at the time of IgA determination. Measurements of 
disease activity and disability were made at 0, 6, 12, and 18 
months. A long term follow up assessment (including Health 
Assessment Questionnaire and joint surgery) was made at 
about 12 years. Mortality was assessed after 15 years. 

Of 352 patients, eight had a primary selective IgA 
deficiency, a point prevalence of 2.3%. A further three had a 
low IgA as part of combined immunoglobulin deficiency. 
Twenty two patients had a selective IgA hypergammaglobuli-
naemia, a point prevalence of 6.3%, with a further 28 having a 
high IgA combined with abnormal levels of IgG or IgM. The 
IgA deficient patients were more likely to have a first degree 
relative with RA than the overall RA population and none of 
this group had RA nodules compared with 29% of RA controls 
(table 1). There was a tendency for the high IgA group to have 
a higher ESR and C reactive protein over the first 18 months 
(table 1). There was a tendency for the high IgA group to have 
a higher ESR and C reactive protein over the first 18 months 
(table 1).

Table 1  Comparison of clinical features in patients with RA with normal and abnormal IgA levels. Value of the clinical 
feature [95% confidence interval] shown where a range is given.

<table>
<thead>
<tr>
<th>Variable</th>
<th>High IgA group (n=22)</th>
<th>Low IgA group (n=8)</th>
<th>RA controls (n=277)</th>
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<tbody>
<tr>
<td>Male female</td>
<td>7.15</td>
<td>2.6</td>
<td>88.18</td>
</tr>
<tr>
<td>Mean age at assessment</td>
<td>56.0 (52 to 60)</td>
<td>56.8 (43 to 71)</td>
<td>52.1 (48 to 57)</td>
</tr>
<tr>
<td>Mean age of onset</td>
<td>50.0 (45 to 55)</td>
<td>49.8 (37 to 60)</td>
<td>46.3 (45 to 49)</td>
</tr>
<tr>
<td>No with 1st degree relative with RA</td>
<td>11 (20)</td>
<td>5 (7)*</td>
<td>99 (277)</td>
</tr>
<tr>
<td>Median Steinbrocker score</td>
<td>2.3 (1.9 to 2.6)</td>
<td>2.0 (1.4 to 2.6)</td>
<td>2.1 (2.0 to 2.2)</td>
</tr>
<tr>
<td>Mean tender joint count</td>
<td>8.9 (6.6 to 11.3)</td>
<td>10.0 (5.0 to 15.0)</td>
<td>9.0 (7.7 to 10.3)</td>
</tr>
<tr>
<td>No with nodules</td>
<td>3.20</td>
<td>0 (7)</td>
<td>53 (275)</td>
</tr>
<tr>
<td>Mean CRP</td>
<td>49.7 (32 to 67.3)</td>
<td>62.9 (31.7 to 94.0)</td>
<td>45.2 (29.7 to 50.7)</td>
</tr>
<tr>
<td>No with positive RA latex (&gt;1/40)</td>
<td>11 (20)</td>
<td>6 (7)</td>
<td>168 (275)</td>
</tr>
<tr>
<td>No with positive ANF (&gt;1/40)</td>
<td>5 (20)</td>
<td>2 (7)</td>
<td>54 (275)</td>
</tr>
<tr>
<td>Mean ESR over 18 months</td>
<td>40.6 (32.1 to 49.0)</td>
<td>33.0 (20.4 to 45.6)</td>
<td>29.9 (26.2 to 32.9)</td>
</tr>
<tr>
<td>Mean CRP over 18 months</td>
<td>40.8 (28.4 to 53.1)</td>
<td>31.5 (15.0 to 48.0)</td>
<td>29.1 (26.4 to 32.7)</td>
</tr>
<tr>
<td>Mean tender joint count over 18 months</td>
<td>7.6 (6.1 to 9.2)</td>
<td>7.3 (2.9 to 11.7)</td>
<td>7.2 (6.0 to 8.4)</td>
</tr>
</tbody>
</table>

Long term follow up                            

| Mean HAG score                                | 1.7 (1.1 to 2.2) [15] | 1.5 (1.4 to 1.6)    |                     |
| No with one or more large joints replaced     | 8 (21)                | 3 (8)               | 87 (276)            |
| Mean no of large joints replaced per patient  | 0.8 (0.2 to 1.3)      | 0.75 (0.2 to 1.7)   | 0.9 (0.7 to 1.1)    |
| No died over 15 years                         | 8                     | 2                   | 61                  |

*OR 4.5, 95% CI 0.8 to 34.1, p=0.053 [compared with RA controls].

DISCUSSION

As far as we know, this study is the first to examine long term 
orality in patients with RA with abnormal IgA levels, and to 
investigate the prevalence of IgA deficiency. The latter has 
been associated with other autoimmune diseases, suggesting 
that it may predispose a person to autoimmune dysfunction. 
Although most cases of primary IgA deficiency are spontane-
umous, familial cases have been described. In our study, IgA defi-
cient patients were more likely to have a history of RA in 
first degree relatives, suggesting inheritance of a predisposing 
factor. Though numbers were small, no similar published study 
was found. The lack of power caused by small sample sizes 
might have prevented us demonstrating more significant dif-
fences. None the less, the findings are of interest. As in pre-
vious work the high IgA group possibly had more active 
disease. However, there was little overall difference between 
the patients with abnormal IgA levels and the controls. These 
findings do not support a role for IgA as a key factor in the 
pathogenesis of RA, or its clinical presentation.

REFERENCES

1 Westedt ML, Daha MR, Baldwin WM, Stijnen T, Cats A. Serum immune 
complexes containing IgA appear to predict erosive arthritis in a 
longitudinal study in rheumatoid arthritis. Ann Rheum Dis 
2 Davis MJ, Dawes PT, Fowler PD, Shadforth AM, Lewin I, Stanworth DR. 
The association and predictive value of the complex immunoglobulin A-
α1 antitypsin in the development of erosions in early rheumatoid 
3 Sfriso P, Lazzarin P, Punzi L, Oustani PA, Iannelli A, Gamari PF. Clinical, 
radiological and laboratory aspects of rheumatoid arthritis associated 
with high serum levels of IgA. Clin Exp Rheumatol 1994;12:690–1.
4 Holt PJ, Tandy NP, Antsee DJ. Screening of blood donors for IgA 
Antineutrophil and related autoantibodies in sera of healthy subjects with 
6 Koskinen S. Long-term followup of health in blood donors with primary 
7 Islam KB, Bashir B, Nilsson L, Hammarstrom L, Sideras P, Smith CI. 
Molecular analysis of IgA deficiency. Evidence for impaired switching to
8 Arnott FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, Cooper NS, et al. The American Rheumatism Association 1987 revised criteria for the 
9 Kirwan JR, Reeback JS. Stanford Health Assessment Questionnaire 
modified to assess disability in British patients with rheumatoid arthritis. 

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Asymptomatic diffuse pulmonary embolism caused by acrylic cement: an unusual complication of percutaneous vertebroplasty

J Bernhard, P F Heini, P M Villiger

Percutaneous vertebroplasty was first performed in 1984. Galibert et al treated a cervical vertebral angioma by percutaneous puncture and injection of polymethylmethacrylate (PMMA) cement into the vertebral body. Shortly thereafter vertebroplasty was done also in lytic metastatic bone lesions. Vertebroplasty for the treatment of vertebral fractures in osteoporosis has gained fast acceptance in the past two years. Its efficacy is documented in several clinical studies.

However, open questions about indication, technical aspects, and complications remain. We report the first case of extensive but clinically silent cement embolisation into the lungs.

CASE REPORT
A 67 year old man presented with upper abdominal pain. A chest radiograph suggested multiple vertebral fractures as a possible explanation. Magnetic resonance imaging confirmed...
Percutaneous vertebroplasty has been reported. In up to 90% of cases, immediate pain relief is reported. The risk of cement extravasation into the inferior vena cava and subsequently into the lungs is rare. To date, only three cases of patients with pulmonary embolism caused by percutaneous vertebroplasty have been reported. In most cases, the extravasation of cement into the inferior vena cava and subsequently into the lungs is rare. To date, only three cases of patients with pulmonary embolism caused by percutaneous vertebroplasty have been reported. All the cases showed paravertebral venous opacity, the embolisation was documented by CT scan and two of the patients remained asymptomatic. In addition, one case of lethal pulmonary embolism was reported after percutaneous vertebroplasty in a series of patients with spinal metastasis. It is important to note, however, that no cement was found in the pulmonary arteries. The risk of cement leakage depends on the vascular anatomy and fracture pattern, on the one hand, and technical aspects, on the other. The case presented here shows a typical fracture pattern without involvement of the posterior wall and offers no additional risks for this treatment. The viscosity of PMMA cement is a crucial aspect during the procedure. The occurrence of this diffuse, extensive lung embolisation is only possible when a considerable amount of cement is injected in a very low viscous state. The cases mentioned above showed only globular cement in major pulmonary vessels. Furthermore, sufficient radio-opacity of the cement is mandatory. In addition, the placement of the tip of the needle needs to be controlled by CT scan or fluoroscopy. As the performing radiologist did not detect dislocation of cement, we suspect a lesion of the basivertebral vein or a horizontal subarticular collecting vein draining into the vena cava inferior at the L1 level in our case. The value of prior vertebrography is controversial. Some authors recommend a venography to exclude needle placement directly within the basivertebral venous plexus. Others argue that the contrast media has different chemical and physical properties and nearly always escapes through the venous plexus. Our experience suggests that extravasation cannot be avoided by previous venography and, therefore, meticulous monitoring of the cement flow during the procedure is crucial.

This case illustrates for the first time diffuse pulmonary cement emboli as a complication of percutaneous vertebroplasty. It supports the notion that plugging a small percentage of arterial pulmonary vessels does not result in respiratory symptoms. However, whether the stiffness or the chemical properties of the cement, or both, may lead to secondary pulmonary lesions is unknown.

REFERENCES


No association between human parvovirus B19 infection and Sjögren’s syndrome

R De Stefano, S Manganelli, E Frati, E Selvi, A Azzi, K Zakrzewska, R Marcolongo

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The association of human parvovirus B19 (HPVB19) infection with autoimmune disease, including systemic lupus erythematosus, rheumatoid arthritis, polymyositis, and vasculitis, has been suggested, although the exact relationship between the infection and these disorders has not been fully elucidated. A recent report showed serological evidence of past B19 infection associated with the presence of cytopenia in patients with primary Sjögren’s syndrome (SS). To gain more information about the aetiopathogenetic role of HPVB19 for this disease, we evaluated the presence of the viral genome in minor salivary glands from patients with primary SS.
We studied 10 women with SS (mean (SD) age 45 (9) years) and 10 healthy controls matched for age (43 (6) years) and sex. SS was diagnosed according to European criteria. Each subject taking part in the study underwent minor salivary gland 6 mm punch biopsy under local anaesthesia. Histological evaluation of biopsy samples was carried out according to Chisholm and Mason’s classification. They were also analysed for the presence of DNA sequence coding for the HPVB19 non-structural protein (NS1) amplified by nested polymerase chain reaction (PCR) as a marker of infection. The outer primer pairs were P1 and P6, corresponding to nucleotides 1399–1422 and 1682–1659. In the second amplification, the P2 and P5 inner nested primer pairs, corresponding to nucleotides 1498–1525 and 1660–1576, were used. The 103 base pair (bp) diagnostic fragment was subsequently detected by ethidium bromide staining after agarose gel electrophoresis. Each sample was tested in duplicate. A 10⁻² dilution of a reference serum containing about 10–100 HPVB19 genome copies was used as positive control. Negative water controls were extracted concomitantly with the diagnostic specimens in order to monitor possible contamination during the extraction step. Additional negative controls were included in each PCR run to eliminate the possibility of carryover contamination. A 268 bp fragment of the β-globin gene was amplified using primers PC04 and GH20 as a test for the absence of Taq DNA polymerase inhibitors and to estimate the quantity of DNA extracted from each minor salivary gland. A serial 10-fold dilution of DNA extracted from a known number of Hep-2 cells was used as positive control. Negative controls were simultaneously extracted water samples. PCR products were then analysed by agarose gel electrophoresis.

Blood samples from each patient were tested for the presence of anti-B19 IgM and IgG using a commercially available enzyme linked immunosorbent assay (ELISA) (Pantec, Turin, Italy).

All minor salivary gland samples of patients with SS were rated as grade III or IV according to Chisholm-Mason’s classification. In the control group, only three subjects were rated as grade I or II (subjects 13, 14, and 17, table 1). The DNA sequence coding for NS1 of HPVB19 was found in a salivary gland specimen from one case of SS (patient 2) and from one control subject (subject 19). Both cases showed a high titre of anti-B19 IgG antibodies and the absence of specific IgM antibodies. In the patient with SS (patient 2) the presence of IgG B19 antibodies was associated with more than one focus score, whereas in the control subject (subject 19) the presence of DNA PvB19 was not associated with lymphocytic infiltrate. Anti-B19 IgG antibodies, but no anti-B19 IgM antibodies, were detected in three other subjects, including one with SS (patient 6) and two controls (subjects 12 and 17). The results of the study showed that the prevalence of past B19 infection in patients with primary SS was similar to that of the control group. Furthermore, none of the patients with SS showed serological markers of recent infection from HPVB19.

<table>
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<th>Subjects</th>
<th>Lymphocytic infiltrate (grade)</th>
<th>IgG</th>
<th>IgM</th>
<th>DNA PvB19</th>
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B19 DNA can also be found, and can persist, in the salivary glands without inevitably inducing a lymphocytic infiltrate in this tissue. Our results suggest that B19 constituents may also be found in salivary gland tissue. However, the presence of viral DNA in the salivary glands of patients with SS appears to be incidental, and it does not support an association between SS and HPVB19 infection.

REFERENCES
Recurrent uveitis in a patient with juvenile spondyloarthropathy associated with tumour necrosis factor α inhibitors

O Kaipiainen-Seppänen, M Leino

**DISCUSSION**

Chronic uveitis is an important complication of JRA. Uveitis is usually asymptomatic and often bilateral. It becomes manifest usually within seven years from the onset of arthritis.¹ In epidemiological studies the incidence of uveitis has varied from 4 to 16% among patients with JRA in population based series.² Up to 27% of patients with juvenile onset ankylosing spondylitis have one or more attacks of non-granulomatous acute uveitis.³ Among 16 patients who had inflammatory eye disease (uveitis or scleritis), 13/16 also having an associated joint disease, and who were treated with etanercept or infliximab, the joint disease responded excellently to treatment, but the eye disease improved in only 6/16 patients (38%).⁴ Five patients developed an inflammatory eye disease for the first time while taking a TNF inhibitor. Among 10 children with uveitis refractory to long term treatment, 3/14 (21%) eyes achieved remission, 5/14 (36%) eyes improved, 5/14 eyes (36%) remained unchanged, and one eye (7%) worsened during etanercept treatment.⁵ In endotoxin induced uveitis in mice, both pretreatment with TNFα or with anti-TNFα antibody caused the ocular inflammation to worsen significantly.⁶ Thus, TNFα blockade may also stimulate certain aspects of immune defence, exacerbating immune reaction in the tissues which TNF inhibitors do not effectively penetrate, such as the central nervous system,⁷ or the eye. The peripheral joint disease responded well to both TNF inhibitors in our patient. Although a disease associated manifestation cannot be excluded, recurrences of uveitis, when the joint disease was in remission, may be secondary to TNF inhibition. Both etanercept and infliximab induced similar cutaneous vasculitis in a susceptible patient, which might be due to anti-drug antibody production or perturbation in the TNF/TNF receptor system in the target organ.⁸ It was recently shown that peripheral T cell reactivity was increased after four and eight weeks of etanercept treatment among patients with RA.⁹ Surveillance of large patient groups is needed to reveal the magnitude of immune reactions associated with TNFα blockade.

**Authors’ affiliations**

O Kaipiainen-Seppänen, Department of Medicine, Kuopio University Hospital, Kuopio, Finland

M Leino, Department of Ophthalmology, Kuopio University Hospital, Kuopio, Finland

Correspondence to: Dr O Kaipiainen-Seppänen, Department of Medicine, Kuopio University Hospital, PO Box 1777, 70211 Kuopio, Finland; Oili.Kaipiainen-Seppanen@kuh.fi

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**REFERENCES**

Iron infusion activates synovium and induced joint inflammation in experimental animals and causes the flaring up of arthritis in patients with rheumatoid arthritis (RA). Marked iron deposition in RA synovia has been reported over the past 30 years and has also recently been demonstrated by quantitative photometric assessment and is correlated with exudative and proliferative histological features. It has also been reported that the amount of iron deposition in RA synovial tissue is correlated with disease activity and severity. Iron has an important role in RA synovitis through the formation of radical oxygen species, and the enhancement of collagen synthesis and synovial fibroblast proliferation possibly owing to down regulation of prostaglandin E2 (PGE2) production.1

Synovial fibroblasts produce a number of inflammatory mediators including cytokines such as interleukin (IL)1, IL6, IL8, fibroblast growth factor, vascular endothelial growth factor, tumour necrosis factor, and granulocyte macrophage colony stimulating factor (GM-CSF). GM-CSF produces the progenitor cells of macrophage lineage stem cells and stimulates mature granulocytes and macrophages. GM-CSF is produced by T cells, macrophages, and fibroblasts and has been found in synovial fluid and tissue from patients with RA. GM-CSF has an important role in type II collagen induced arthritis in rats and in the acute methylated bovine serum albumin induced murine arthritis model. A protective effect against collagen induced arthritis was seen in GM-CSF knockout mice. Using those mice, it has been recently shown that GM-CSF plays a part in the IL1 induced arthritis that follows methylated bovine serum albumin injection.2 In this study we showed that iron enhanced GM-CSF but did not enhance IL6 or IL8 production by human synovial fibroblasts on stimulation with IL1β in vitro.

Synovial tissues were obtained from 20 patients, 11 with RA and nine with osteoarthritis. The synovial fibroblasts were isolated according to a method described previously.3 The material containing the synovial fibroblasts during the third to seventh passage was used in the experiments. Synovial fibroblasts were added at a concentration of 1×10⁴ cells/well to each well of a 96 well microtitre plate, and cultured for 18, 24, 48, 72, 96, and 120 hours with or without recombinant human IL1β (R&D Systems, UK). Ferric citrate was used as an iron salt or sodium citrate as a control. The differences in cytokine production by synovial fibroblasts cultured with ferric citrate compared with sodium citrate or medium alone were analysed by paired Student’s t test. A level of p<0.05 was accepted as significant.
To investigate the effects of iron on GM-CSF production by synovial fibroblasts, three different concentrations of ferric or sodium citrate (0.01, 0.1, and 1 mmol/l at final concentration) were added to synovial fibroblast 96 hour culture with three different concentrations of IL1β (1, 10, and 100 ng/ml at final concentration). Ferric citrate (1 mmol/l) but not sodium citrate significantly enhanced GM-CSF production by synovial fibroblasts on stimulation with 1 ng/ml of IL1β (fig 1A). Ferric citrate (0.1 mmol/l) enhanced GM-CSF production by synovial fibroblasts on stimulation with 10 ng/ml of IL1β (fig 1B). Also, concentrations of 0.01 or 0.1 mmol/l ferric citrate enhanced production of GM-CSF by synovial fibroblasts on stimulation with 100 ng/ml of IL1β (fig 1C). However, ferric citrate at any concentration tested did not enhance IL6 or IL8 production by synovial fibroblasts on stimulation with any concentration of IL1β including 100 ng/ml (data not shown).

To test whether iron regulates the transcriptional level of GM-CSF production by synovial fibroblasts on stimulation with IL1β, GM-CSF, IL6 and IL8 mRNA expression in synovial fibroblasts were examined semiquantitatively by reverse transcriptase-polymerase chain reaction (RT-PCR). The method used for RT-PCR using specific primers for cytokines and glyceraldehyde-3-phosphate dehydrogenase as a control has been described in detail previously.1 The expression of mRNA encoding these cytokines in synovial fibroblasts was undetectable without IL1, and was dose dependent on IL1β reaching a plateau after three hours in culture. Ferric citrate (0.1 mmol/l) significantly enhanced IL1 induced GM-CSF mRNA expression in synovial fibroblasts but not that of IL6 or IL8 (data not shown).

GM-CSF is produced by T cells, macrophages, mast cells, endothelial cells, and fibroblasts in response to specific activating signals. GM-CSF gene expression is controlled by binding of transcription factors such as NF-GMa, NF-GMb, NF-κB, IFN-γ and Weiss recently showed that iron regulates the transcriptional level of IL8 mRNA following IL-1 induced GMCSF production by synovial fibroblasts. Dlaska and Weiss recently showed that iron regulates the transcription of inducible nitric oxide synthase (iNOS) of macrophage-like cells stimulated with interferon γ (IFNγ) and/or lipopolysaccharide, and binding of NF-κB to its consensus motif within the iNOS promoter was reduced by iron and enhanced by an iron chelator.7 We found that iron enhanced IL1 induced GM-CSF production by synovial fibroblasts. Taken together, the intracellular iron levels might control GM-CSF production induced by IL1 by mechanism(s) similar to the action of iron-responsive element binding proteins on ferritin or transferrin receptor synthesis or iNOS induction.6

We found that the effects of iron on GM-CSF production induced by IL1 were different from those on IL6 or IL8 production. Agro et al reported that PGE, enhanced IL6 and IL8 but inhibited GM-CSF production by IL1 stimulated synovial fibroblasts.8 Previously, we showed that iron decreased PGE production by synovial fibroblasts.1 Iron might enhance GM-CSF production through down regulation of PGE production. Yoshida et al found that gold compounds and divalent metal ions inhibited induction of IL6 and IL8 but not production of GM-CSF by IL1 induced synovial fibroblasts through inhibition of NF-κB binding to DNA.9 Thus, there may be different signal transduction pathways among these three cytokines in IL1 stimulated human synovial fibroblasts.

Clinical analysis of gouty patients with normouricaemia at diagnosis

Y-B Park, Y-S Park, S-C Lee, S-J Yoon, S-K Lee

Gouty attacks usually occur in patients with hyperuricaemia, but patients with gout who have normouricaemia are believed to be in the minority.1 2 This is a common conception about gout because monosodium urate crystals are formed at blood concentrations of uric acid over 420 μmol/l at a body temperature of 37°C.3 Recently, two studies showed that the incidence of gouty attacks in normouricaemia was as high as 39–43%,4 1 which is much higher than our traditional view.1 2 However, few data on normouricaemia in gout have been reported so far.5 The prevalence of patients with gout who have normouricaemia at diagnosis has not yet been determined, and even their clinical characteristics and laboratory findings have not been reported.

REFERENCES


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We designed this study to determine the prevalence of normouricaemia in patients with gout at diagnosis, and to determine the natural course of normouricaemic gout and the differences between the clinical characteristics of patients with normouricaemic and hyperuricaemic gout at diagnosis.

We retrospectively reviewed 226 Korean patients who were newly diagnosed as having gout at the Severance Hospital, Yonsei University College of Medicine, Seoul, Korea, between January 1996 and May 2000. The diagnosis of gout was made during an acute attack of gouty arthritis and confirmed by either the presence of negatively birefringent needle shaped crystals or by the uricase enzyme method.

Among the 226 gouty patients, 27 (12%) male patients showed normouricaemia at diagnosis. Table 1 summarises the differences between the clinical characteristics of patients with normouricaemia in patients with gout at diagnosis, and to analyse the incidence at the time of diagnosis.

**Table 1** Comparison of the clinical and laboratory variables of normouricaemic gout and hyperuricaemic gout at diagnosis

<table>
<thead>
<tr>
<th>Demographics</th>
<th>Normouricaemic gout (n=27)</th>
<th>Hyperuricaemic gout (n=81)*</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age [years]</td>
<td>60.6 (13.9)</td>
<td>54.2 (11.8)</td>
<td>0.02</td>
</tr>
<tr>
<td>First symptom to diagnosis [months]</td>
<td>32.2 (36.3)</td>
<td>54.2 (66.3)</td>
<td>0.03</td>
</tr>
<tr>
<td>BMI [kg/m²]</td>
<td>23.8 (2.9)</td>
<td>23.1 (3.3)</td>
<td>NS</td>
</tr>
<tr>
<td>Associated conditions</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hypertension</td>
<td>15</td>
<td>40</td>
<td>NS</td>
</tr>
<tr>
<td>Renal insufficiency†</td>
<td>5</td>
<td>28</td>
<td>NS</td>
</tr>
<tr>
<td>Heavy alcoholic intake</td>
<td>5</td>
<td>9</td>
<td>NS</td>
</tr>
<tr>
<td>Diabetes mellitus</td>
<td>2</td>
<td>11</td>
<td>NS</td>
</tr>
<tr>
<td>Precipitating events‡</td>
<td>11</td>
<td>35</td>
<td>NS</td>
</tr>
<tr>
<td>Associated diseases§</td>
<td>12</td>
<td>30</td>
<td>NS</td>
</tr>
<tr>
<td>Articular involvement</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>First MTP</td>
<td>19</td>
<td>57</td>
<td>NS</td>
</tr>
<tr>
<td>Upper extremities</td>
<td>1</td>
<td>15</td>
<td>NS</td>
</tr>
<tr>
<td>Lower extremities</td>
<td>27</td>
<td>76</td>
<td>NS</td>
</tr>
<tr>
<td>Polyarticular involvement¶</td>
<td>11</td>
<td>45</td>
<td>NS</td>
</tr>
<tr>
<td>Documented tophi</td>
<td>4</td>
<td>36</td>
<td>0.006</td>
</tr>
<tr>
<td>Serum uric acid (µmol/l)</td>
<td>320 (80)</td>
<td>550 (90)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>24 Hour urinary uric acid excretion (mmol/day)</td>
<td>4 (1)</td>
<td>4 (5)</td>
<td>NS</td>
</tr>
<tr>
<td>BUN (mmol/l)</td>
<td>5.5 (2.0)</td>
<td>7.5 (1.5)</td>
<td>0.02</td>
</tr>
<tr>
<td>Creatinine (µmol/l)</td>
<td>100 (30)</td>
<td>140 (110)</td>
<td>0.002</td>
</tr>
</tbody>
</table>

*Patients with hyperuricaemia were randomly selected from the study group using a table of random sampling. Renal insufficiency was defined as a serum creatinine level above 130 µmol/l or a creatinine clearance below 1.00 ml/s; †precipitating events for an acute attack of gout are binge drinking, meat, surgery, infection, trauma, physical stress, and deterioration of underlying medical disease; §associated diseases are acute renal failure, chronic renal failure, renal transplantation, coronary artery occlusion disease, heart failure, stroke, and pulmonary tuberculosis; ¶polyarticular involvement was defined as more than one joint affected in a gouty attack.

We found that 27/226 (12%) of gouty patients at diagnosis had normouricaemia, which was a lower incidence than found in the two reports mentioned previously. The discrepancy between our results and the results of those studies may be because the previous studies analysed the incidence of gouty patients with normouricaemia at the time of any acute attack, whereas we analysed the incidence at the time of diagnosis.

Seventeen of 21 (81%) gouty patients with normouricaemia at diagnosis subsequently became hyperuricaemic. The median time for progression from normouricaemia to hyperuricaemia was one month. Our results corresponded with those of previous reports which showed that serum uric acid usually fell during an acute attack and rose during the interim. Only four gouty patients with normouricaemia at diagnosis continued to have normouricaemia. This "genuine" normouricaemic gout was present in only a small proportion, and these patients followed a mild disease course without secondary attack.

These observations may help a doctor to decide whether a diagnosis of gout can be made when normouricaemia is present and may help in predicting the disease course of normouricaemic gout at diagnosis.
HLA-DRB1*03 and DQB1*0302 associations in a subset of patients severely affected with systemic lupus erythematosus from western India

U Shankarkumar, K Ghosh, S S Badakere, D Mohanty

Genetic factors are likely to be important both in determining the overall susceptibility to systemic lupus erythematosus (SLE) and in influencing the remarkable clinical heterogeneity in disease expression found in affected subjects. The more common clinical features seen in patients with SLE include, skin and joint diseases, renal disease, neuropsychiatric complications, and some haematological abnormalities. Genetic factors, together with environmental factors, strongly influence the development of SLE. Multiple loci within the major histocompatibility complex (MHC) have been implicated in susceptibility as have HLA class II alleles, complement components, and tumour necrosis factor (TNF) loci.

Currently it is believed that some HLA alleles are in genetic linkage disequilibrium with certain disease related genes and they regulate the immune responses. Since 1969, when the first case of SLE was reported from India, the disease has been extensively studied in different regions of the country—namely, Chennai, Calcutta, Mumbai, and New Delhi. A statistically significant clinical correlation comparing the clinical variables from other racial groups of the world has been reported in Indian patients with SLE. HLA association studies from Indian patients with SLE are considerably limited and, furthermore, varying interethnic differences in the associations have been reported from UK, South African and Icelandic populations.

**PATIENTS AND METHODS**

We studied 53 patients with severe SLE exhibiting the clinical manifestations described by the 1982 revised American Rheumatism Association diagnostic criteria who had one or more organ affected, such as kidney, brain, heart, and lungs. One hundred and ten normal healthy subjects with the same economic status and ethnic background comprised the controls for this study over the same period. The autoantibody profiles among the patients with SLE were studied by immunofluorescence and enzyme linked immunosorbent assay (ELISA). HLA-A and B locus antigens were identified by a National Institute of Health two stage microlymphocytotoxicity assay using indigenous and commercial antisera. The HLA-DRB1 and HLA-DQB1 alleles were determined by a polymerase chain reaction with sequence-specific primers technique from the total genomic DNA extracted from the EDTA blood. The phenotype frequencies, odds ratio, probability value, and confidence intervals were estimated using our database and computer programs. The p value was corrected by the Bonferroni inequality method.

**RESULTS AND DISCUSSION**

A significant increase in the frequency of HLA-A1, A2, B27, DRB1*03, DQB1*0302, and DQB1*0601 was found among patients with SLE. HLA-A19, B15, DRB1*14(6), DRB1*1001, and DQB1*0203 were found to be decreased in the patient group compared with the controls (table 1). The high risk alleles DRB1*03 and DQB1*0302 were then compared with the findings for other populations of the world (table 2). The

**Table 1** HLA alleles associated with severe SLE

<table>
<thead>
<tr>
<th>HLA</th>
<th>Patients (n=53)</th>
<th>Controls (n=110)</th>
<th>OR</th>
<th>p value</th>
<th>CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>A1</td>
<td>39.60</td>
<td>17.80</td>
<td>2.99</td>
<td>&lt;0.001</td>
<td>1.06e2.08</td>
</tr>
<tr>
<td>A2</td>
<td>45.30</td>
<td>29.70</td>
<td>1.95</td>
<td>&lt;0.001</td>
<td>0.52e1.02</td>
</tr>
<tr>
<td>A19</td>
<td>26.50</td>
<td>43.40</td>
<td>0.48</td>
<td>&lt;0.001</td>
<td>9.62e10.72</td>
</tr>
<tr>
<td>B15</td>
<td>2.00</td>
<td>16.30</td>
<td>0.16</td>
<td>&lt;0.001</td>
<td>5.07e7.23</td>
</tr>
<tr>
<td>B27</td>
<td>12.20</td>
<td>2.30</td>
<td>5.40</td>
<td>&lt;0.001</td>
<td>1.42e2.54</td>
</tr>
<tr>
<td>DRB1*03</td>
<td>50.00</td>
<td>6.70</td>
<td>9.67</td>
<td>&lt;0.0001</td>
<td>8.41e13.56</td>
</tr>
<tr>
<td>14(6)</td>
<td>12.50</td>
<td>40.00</td>
<td>0.25</td>
<td>&lt;0.001</td>
<td>1.09e2.56</td>
</tr>
<tr>
<td>1001</td>
<td>6.30</td>
<td>30.00</td>
<td>0.21</td>
<td>&lt;0.001</td>
<td>1.84e2.34</td>
</tr>
<tr>
<td>DQB1*0302</td>
<td>56.30</td>
<td>10.00</td>
<td>8.02</td>
<td>&lt;0.0001</td>
<td>12.88e18.45</td>
</tr>
<tr>
<td>0203</td>
<td>12.50</td>
<td>50.00</td>
<td>0.17</td>
<td>&lt;0.0001</td>
<td>1.32e2.56</td>
</tr>
</tbody>
</table>

*%PF, percentage phenotype frequency; OR, odds ratio; CI, 95% confidence interval.

comparison strongly supported the importance of ethnic background and indicated that the relative importance of different genes may vary in different populations studied. One of the important observations was that the Indian patients with SLE with HLA-DRB1*03 association overlap with the Spanish, Canadian, white, and Mexican populations. However, the Malaysian and Taiwanese population, who are predominantly mongoloid race, showed a different HLA association. All the patients with severe SLE studied presented with a high titre of antinuclear factor, antihistone antibodies, and anti-Sm antibodies. Further, it was found that 54% of the patients were anti-dsDNA positive, 10% had anti-Ro/SS-A antibodies, 22% had anti-La/SS-B antibodies, 14% had both Ro SS-A and La SS-B antibodies.

In SLE multiple loci within the MHC have been implicated in susceptibility-like HLA class II alleles, complement components, and TNF loci. HLA-DR2 and DR3 are both known to be associated with SLE and inheritance of both DR2 and C4B null alleles confers a higher relative risk. HLA-DQ1 and DQ2 are also found to be associated with Ro, La, Sm, and dsDNA autoantibodies. The HLA-DR2, DQ1 haplotype associated with SLE has low TNF levels. A review based on the clinical and laboratory measurements in Indian patients with SLE showed a higher proportion of alopecia, renal lupus, oral ulcers, and neurological involvement, reaching statistically significant levels when compared with other racial groups. HLA studies from patients with SLE from northern India showed an appreciable risk of HLA-DR4 among the patients, and additionally, the haplotype B8-DR4 was often found in the patient group. However, HLA-DR3 and TNF promoter polymorphisms in the white patients with SLE were independently associated. Interethic differences in the associations of TNF promoter polymorphism with SLE have also been reported from the UK, South Africa, and Iceland.

The interesting observations was the increased incidence of renal side effects with disease-modifying drug “azarofin” in patients with rheumatoid arthritis, who incidentally inherited DRB1*0301, an allele linked strongly with SLE in his study as well as in ours. It may be relevant to mention here that the DRB1*0301 allele is present in 6.7% of the normal population, hence they may also become susceptible to the nephrotoxic effect of these kind of drugs when they receive treatment for various disease conditions.

In conclusion, our results show, firstly, a significant twofold increase in the odds ratio for the presence of HLA-DRB1*03, and DQB1*0302, alleles and, secondly, a significant twofold decrease in the odds ratio for the presence of HLA-A19, DRB1*14(6), DRB1*1001, and DQB1*0203 alleles among the western Indian patients with SLE. Finally, the association supports the importance of ethnic background and indicated that the relative importance of different genes may vary in different ethnic populations around the world.

Table 2 Comparison of HLA allele associations in patients with SLE from other populations of the world

<table>
<thead>
<tr>
<th>Population</th>
<th>Number studied</th>
<th>HLA alleles associated</th>
<th>Reference Number</th>
</tr>
</thead>
<tbody>
<tr>
<td>Spanish</td>
<td>85</td>
<td>DR3, DQA1*0501</td>
<td>6</td>
</tr>
<tr>
<td>Canadian</td>
<td>214</td>
<td>DR3</td>
<td>7</td>
</tr>
<tr>
<td>White</td>
<td>99</td>
<td>DRB1*0301</td>
<td>5</td>
</tr>
<tr>
<td>Malaysian</td>
<td>56</td>
<td>DR2, DQB1<em>0501, DQB1</em>0601</td>
<td>11</td>
</tr>
<tr>
<td>Taiwanese</td>
<td>34</td>
<td>DRB1*0301</td>
<td>9</td>
</tr>
<tr>
<td>European</td>
<td>577</td>
<td>DRB1<em>0402, 07, DQB1</em>0302, 0301, DQA1*0201</td>
<td>8</td>
</tr>
<tr>
<td>Mexican</td>
<td>81</td>
<td>DRB1*0301</td>
<td>10</td>
</tr>
<tr>
<td>Western Indian</td>
<td>53</td>
<td>DRB1<em>03, DQB1</em>0302</td>
<td>Present study</td>
</tr>
</tbody>
</table>

**REFERENCES**

Asymptomatic diffuse pulmonary embolism caused by acrylic cement: an unusual complication of percutaneous vertebroplasty

J Bernhard, P F Heini and P M Villiger

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