MATTERS ARISING

EULAR recommendations for the management of knee osteoarthritis

I welcome the publication of the EULAR recommendations for the management of knee osteoarthritis by the EULAR Standing Committee for International Clinical Studies Including Therapeutic Trials (ESCISIT).1 However, it is out of date and has been superseded by other more up to date publications on the recommendations for the management of osteoarthritis of the hip and knee by the American College of Rheumatology (ACR) Subcommittee on Osteoarthritis Guidelines2 and another review of the medical management of osteoarthritis.3 The newer COX-1 sparing non-steroidal anti-inflammatory drugs, rofecoxib and celecoxib are not discussed in the EULAR recommendations, but they have been included in the other reviews.2,3

My second criticism of the report is that it contains statements such as “SYSADOAs may possess structure modification properties” which is misleading and may lead to widespread use of these drugs in the absence of good evidence.

Although the Annals of the Rheumatic Diseases is the official EULAR journal, EULAR reports should be peer reviewed before publication. The ACR recommendations for the management of osteoarthritis of the hip and knee were only accepted in a revised format.4 Two of the authors of the recent review5 are also coauthors of the EULAR report.

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Authors’ reply

We thank Dr Jawad for his interest in the EULAR recommendations.1 The fact that the literature search ended at December 1998, excluding evidence based discussion of COX-2 inhibitors, is clearly highlighted and discussed in the paper. Unfortunately, a finite end point needs to be set on any thorough evidence based review, and even the ACR document1 was of necessity four months out of date by the time it was published. However, the important strengths of the EULAR document compared with the ACR report are:

1 It used a structured evidence based format rather than just consensus statements.

2 It had input from 25 experts from 12 countries, as opposed to four expert Americans.

3 It undertook additional analyses on published trials to calculate effect sizes and numbers needed to treat where sufficient data were available (that is, it provided new data).

4 The search included surgery and treatments such as topical non-steroidal anti-inflammatory drugs, which are not licensed in the USA and which were excluded from the ACR paper.

5 It provided data on the “epidemiology” and quality of clinical trials in knee osteoarthritis (OA) and highlighted important questions that need to be addressed by future research.

Unlike the review by Walker-Bone and three other coauthors from Southampt,1 the EULAR report encompassed both medical and surgical treatments, and carefully examined the literature in response to specific clinically relevant questions rather than in an attempt to produce a pragmatic algorithm. We also calculated effect sizes rather than estimates of efficacy, and did not extrapolate treatment data for the knee (the site to which most data relate) to other joint sites. The difference in outcome from these two contrasting approaches is shown, for example, by the recommendations in the EULAR report to consider intra-articular steroid injection only if an effusion is present, whereas the EULAR committee concluded there is no clear evidence to justify a knee effusion as a predictor of response to such treatment.

The qualified statement that “SYSADOAs may possess structure modification properties, but more studies, using standardised methodologies, are required” was based on available evidence and made without prejudice. To introduce bias by ignoring published reports on such treatments would indeed be misleading and against all the ethos of evidence based practice.

Although this was a paper that was drafted, edited, and remodified by 21 European experts in OA, the manuscript was peer reviewed by two additional referees after its submission to the Annals.

The committee are delighted with the growing interest in the management of OA and that three contrasting reviews were published on the topic in 2000. We, of course, realise the current interest in highly selective COX-2 inhibitors. Importantly, however, the three reports fully concur with respect to the important elements of OA management—the non-pharmacological interventions. They also all agree that paracetamol remains the oral analgesic to try first and, if successful, is the preferred long term oral analgesic.

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Coffee consumption, RF, and the risk of RA

We read with interest the paper entitled “Coffee consumption, rheumatoid factor, and the risk of rheumatoid arthritis”,1 in which an association between coffee consumption and the presence of a positive rheumatoid factor (RF) in healthy subjects was reported. The authors also observed an increased risk of developing seropositive rheumatoid arthritis (RA), particularly in people who were lifelong non-smokers and drink four cups of coffee a day. Cigarette smoking seemed to be the single most important risk factor for the development of a positive RF in healthy subjects and was strongly associated with coffee consumption. In view of this, it is unsurprising that there is no additive risk noted for the development of seropositive RA in people who were heavy smokers and also habitual coffee drinkers.

We suggest that the observed risk for developing seropositive RA in people who have never smoked and who consume more than four cups of coffee is possibly confounded by exposure to passive cigarette smoke. For example, coffee is often consumed in coffee houses, which are prone to be particularly smoky environments, and coffee drinkers are likely to mix with other coffee drinkers, who are statistically more likely to be smokers.

The risk observed for developing seropositive RA in this study is modest for subjects smoking fewer than 14 cigarettes a day (relative risk 1.28, CI 0.63 to 2.57). At face value, it would seem unlikely that passive smoking could play a role in the development of seropositive RA. However, a Chinese study noted that active cigarette smoking was a moderate risk factor for nasopharyngeal carcinoma (NPC) (OR=1.28, 95%CI 1.02 to 1.61). Despite this, it was observed that women who were lifelong non-smokers had a strong and statistically significant positive association between NPC risk and exposure to substantial passive smoke as a child or an adult.2 Interestingly, those with NPC had an increased risk of a family history of NPC compared with controls. Subjects with RA who are lifelong non-smokers would seem to have a greater genetic predisposition to RA, indeed we have found a strikingly high prevalence of a positive family history of RA in a first or second degree relative in these subjects (94%).1 Possibly, in such genetically “predisposed” subjects passive smoke may induce rheumatoid factor production by a mechanism similar to that in people who are active smokers. The presence of a positive RF in a healthy person is important, as this increases the risk sevenfold for the development of seropositive RA.2

A second potential mechanism by which passive smoke inhalation might predispose subjects to RA is by changing the local environment of the nasopharynx. Both the pneumococcus and meningococcus bacteria are carried in the nasopharynx and both invasive pneumococcal and meningococcal disease are significantly more often in passive smoke.
smokers.  It is interesting to note that NPC is associated with the Epstein-Barr virus, one of a number of viruses which have been implicated in the pathogenesis of RA. Therefore, passive smoke might potentially predispose subjects to RA as a result of changes within the nasopharynx resulting in antigenic stimulus by a virus that triggers RA.

We agree with the authors that new information is urgently needed about any factor associated with the risk of RA. In view of emerging data highlighting smoking as an important environmental risk factor for the development of seropositive RA,1 and also this study by Heliövaara et al, we propose that passive smoking should be considered as a potential candidate factor for the development of RA.

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No significant association was seen between the number of daily cups of coffee and serum cotinine concentration (age adjusted \( r=0.08, p=0.38 \)). Contrary to the assumption of Drs Hutchinson and Moots, there was a negative correlation between serum thyioate level and coffee consumption (age adjusted \( r=-0.21, p=0.03 \)). The findings are in agreement with our impression that in Finland coffee is not consumed to any greater extent in notable smoking environments. Rather, coffee consumption is confined to breakfast at home and coffee breaks both during working hours and leisure time.

On the basis of these preliminary results it seems unlikely that passive smoking would have correlated strongly enough with coffee consumption to explain the close associations between coffee consumption and the occurrence of rheumatoid factor (RF) and the risk of rheumatoid arthritis (RA) in our study. Nevertheless, our results are far from being conclusive. Whatever the links between coffee consumption, RF, and RA, we agree with Drs Hutchinson and Moots that passive smoking should be considered a potential risk factor for RA.

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Authors’ reply

It is not easy to measure passive smoking. Nevertheless, determinations of serum cotinine and thiocyanate recommended for the detection of indirect exposure to tobacco smoke have been shown to indicate exposures of recent occurrence.1

To test the assumption put forward by Drs Hutchinson and Moots we studied coffee consumption for its associations with serum cotinine and thiocyanate concentrations in a sample of men who had participated in the Mini-Finland Health Survey and served as an age matched control group in a nested case-control study of lung cancer.2 A modification of the Nicotine Metabolite RIA kit method (Diagnostic Products Corporation, Los Angeles, USA) was used to determine serum cotinine concentrations. Serum thiocyanate was determined by the spectrophotometric ferric nitrate method.3 Of the total of 158 men in the sample, 39 reported current smoking, and 10 others had serum cotinine \( \geq 200 \) mg/l or thiocyanate \( \geq 20 \) mmol/l, suggesting direct exposure to tobacco. Exclusion of these men left 109 non-smokers for the final analyses.

No significant association was seen between the number of daily cups of coffee and serum cotinine concentration (age adjusted \( r=0.08, p=0.38 \)). Contrary to the assumption of Drs Hutchinson and Moots, there was a negative correlation between serum thyioate level and coffee consumption (age adjusted \( r=-0.21, p=0.03 \)). The findings are in agreement with our impression that in Finland coffee is not consumed to any greater extent in notable smoking environments. Rather, coffee consumption is confined to breakfast at home and coffee breaks both during working hours and leisure time.

On the basis of these preliminary results it seems unlikely that passive smoking would have correlated strongly enough with coffee consumption to explain the close associations between coffee consumption and the occurrence of rheumatoid factor (RF) and the risk of rheumatoid arthritis (RA) in our study. Nevertheless, our results are far from being conclusive. Whatever the links between coffee consumption, RF, and RA, we agree with Drs Hutchinson and Moots that passive smoking should be considered a potential risk factor for RA.

adrenal gland, ileum (fig 1), and ascending colon. Several stages of vasculitis existed, which was a typical finding of classic PAN. We failed to detect vasculitis affecting arterioles, venules, or capillaries. Duodenal bleeding was from peptic ulcers, which is not associated with arteritis.

A few reports have described TTP complicating certain forms of rheumatic diseases, including systemic lupus erythematosus and systemic sclerosis. To our knowledge there are no reports published in English of PAN complicated by TTP.

The necropsy finding was classic PAN because vasculitis affected vessels larger than arterioles and there were no pathological findings of glomerulonephritis. We failed to detect vasculitis affecting arterioles, venules, or capillaries. MPO-ANCA is usually a marker of microscopic polyarteritis or necrotising glomerulonephritis but does not distinguish it from classic PAN with certainty.

Our case suggests the possibility of secondary TTP due to PAN. We speculate that endothelium damage by PAN may enhance the development of TTP, particularly in the presence of inflammatory processes. Our patient unfortunately died from massive bleeding from a duodenal ulcer. However, conventional treatment for TTP, including plasma infusion and plasma exchange, were effective in this case, too. Our case emphasises the need to consider TTP when thrombocytopaenia occurs with vascular disease, because early and correct treatment of TTP may improve morbidity and mortality in these patients.

PlACEMENT OF INTRA-ARTICULAR INJECTIONS 

verified by ultrasonography and injected air as contrast medium

Intra-articular injection of long acting corticosteroid is a corner stone in rheumatological treatment. The injected intra-articular corticoid is more effective when correctly placed. Injection of radiographic contrast material has shown that fewer than half of the injections are correctly placed in the joint space after blind injection.

Generally, the clinical application of ultrasonographic examinations can be enhanced by contrast agents. The most commonly used technique is creation of microbubble contrast agents. Such agents, applied to the bloodstream, have been used for hepatic, nephrologic, cardiologic, and transcranial examinations.

Obviously, the risk of air embolism depends on the anatomical site of the injected air contrast. Transient ischaemic attacks are described after echocardiography with air contrast and in animal models haemodynamic effects during venous air infusion can be measured.

Intra-articular injection of air and subsequent lateral and posterior radiographs have shown that this technique can enhance the precision of the procedure. The disadvantage of this method is that the result can first be seen after the injection, and that a correction can only be made with a new injection. In the joint space the air is separated from the vascular system and only small amounts of sterile air are used. The risk of venous air embolism is negligible. Air is a very effective contrast medium in ultrasonography. Air sonography has been used for the diagnosis of meniscus lesions in knee joints and for rotator cuff lesions in the shoulder.

We expand the applicability of this method to all joints, not only for diagnosis, but also for the correct placement of the needle before the injection of medicine (steroid, osium acid, viscosupplementation). The sterile air that is contained in the cuffed vain with lidocain or steroid is used as contrast medium. The needle is guided into the joint space of the distended capsule by ultrasonography.

When the steroid and lidocain are mixed in the syringe a small volume of air will be in the needle itself (<0.5 ml). The air in the needle is clearly seen when the injection is started and will secure the correct placement of the needle. With this technique, it is not necessary to use two separate syringes and the inclination of the syringe will not cause the air to move from the needle to the bottom of the syringe.

If the knee is injected, injection directly into the recess of the knee is recommended, which will make the small volume of air momentarily visible.

Figure 1 illustrates the ultrasonography of a metatarsophalangeal joint in a patient before and after injected air. The intra-articular air is clearly seen. We have made over 1000 ultrasonic guided intra-articular injections without any complications. This method is easy, inexpensive, without risk and radiation, and should be used routinely in rheumatology. Chemical synovectomy of the knee is especially, should always be guided by ultrasonography, and with this technique smaller joints can also be considered for chemical synovectomy.

7. Blidhal H. Placement of intra-articular injec-
9. Wei R, Strømsø AE. Air as contrast medium in sonography of the rotator cuff (arthro-sonographic diagnos-

HLA class II alleles and synovial fluid cytoky in RA

Rheumatoid arthritis (RA) is associated with HLA-DR4, which is encoded by the DRB1 gene. This genetic predisposition has been shown to lie within the sequence motif present in the third hypervariable region of the DRB1 gene. This sequence of amino acids has been called the “disease epitope” and can be encoded by DR4 subtypes as well as non-DR4 alleles; DR1 subtype, DR10, and DR14 subtypes. Hence, it is also termed the shared epitope. In addition to imparting susceptibility to RA, HLA-DR4 has also been shown to be associated with the severity of the rheumatoid disease, including destructive erosive joint disease, rheumatoid phe-
toty, and extra-articular manifestations, such as rheumatoid nodules, vasculitis, and Felty’s syndrome. HLA-DQ genes are in linkage disequilibrium with HLA-DR and subjects who are DR4 positive may either be DQB1*0301 or *0302 positive; certain extra-
articular features of RA have been shown to be associated with HLA-DQB1*0303.

Synovial fluid cytoky in RA is heterogeneous both with respect to total white cell

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count and to the proportions of polymorphs, lymphocytes, and ragocytes. Ragocytes are leucocytes containing intracytoplasmic granules and have been shown to be present in higher percentages in the joints of patients with RA. Davis et al showed that when a patient with RA has a relapse affecting the same joint, the proportions of cell types remain similar, though the total white cell count may differ. The same study also found a significantly poorer outcome in patients with persistently high ragocyte counts as assessed by the necessity for subsequent joint replacement or synovectomy.

As both synovial cytology and genetic factors have been shown independently to predict joint damage, this study aimed at assessing whether there is a relation between genetic factors and the pattern of synovial fluid cytology which might also be of prognostic significance.

Fifty nine unrelated patients with RA attending the Department of Rheumatology, Hope Hospital, Salford, were studied. All the patients were white and had classic or definite RA according to the 1987 American Rheumatism Association Criteria. All had synovial fluid analysed on at least one occasion during articular flares of arthritis. Blood was taken for HLA typing from each patient. Clinical details recorded were age, sex, disease duration, and drug treatment.

All synovial fluid samples collected were placed in 2 ml paediatric lithium heparin bottles and processed the same day. Total white cell count was carried out with a modified Fusch Rosenthal counting chamber and expressed as cells/cm³. Polymorphs, lymphocytes, monocytes, and ragocytes were differentiated in a cytocentrifuged and wet preparation using simple morphology and expressed as percentages of all nucleated cells. Genomic DNA was extracted as described previously. DRB, DQA and B were typed by polymerase chain reaction, followed by hybridisation to specific oligonucleotide probes. Rheumatoid factor was measured by the sheep cell agglutination test and titres >1/32 were considered positive. Data for white blood cells, polymorphs, lymphocytes, ragocytes, and monocytes were not normally distributed and were expressed as medians (interquartile range). The Mann-Whitney U test was used to compare these variables between the different patient groups. When more than two groups were compared the Kruskal-Wallis one way analysis of variance was used. Corrections for multiple comparisons were also made.

The 59 patients (45 female, 14 male) had a median age of 63 years (range 27–85) and median duration of disease of 16.5 years (range 2–53). Of these 59 patients, 59 were seropositive for rheumatoid factor, 52 had erosive disease, 27 had extra-articular features (nodules, Felty's syndrome, Sjögren's syndrome, vasculitis, peripheral neuropathy, pulmonary, renal disease, and/or eye disease), and 43 were receiving a disease modifying antirheumatic drug (DMARD) with or without oral steroids at the time of the study.

HLA-DR typing showed that 44 (75%) patients were DR4 positive. Eight (18%) of the DR4 positive patients were compound heterozygotes (DRB1*0401/0404). Forty five (76%) patients were “RA epitope” positive (DR4/DRB1*0401, DR4/DRB1*0404, DR4/DRB1*0405, or DR1) and 24 (41%) were heterozygous for the “RA epitope”. HLA-DQ typing showed that 25 (42%) patients were positive for DQB1*0301.

When the patients were subdivided into those who were positive or negative for HLA-DR4, there were no differences in the total white cell count or proportion of the cell types between the two groups, and these were also similar between patients who were or were not compound heterozygotes. However, when the patients were subdivided into those who were epitope positive or negative, there was an increased median percentage lymphocyte count in patients who were positive (median 11%) compared with those who were negative (median 3%; p=0.036, pcorr>0.05). There were no differences in the other cell type counts between the two groups.

When the patients were subdivided into those who were positive or negative for HLA-DQB1*0301, patients who were positive for the allele (n=25) had an increased median percentage ragocyte count (50%) compared with those who were negative (n=20, 10%, p=0.033, pcorr>0.05). The percentages of the other cell types were similar in the two groups. When the HLA-DR4 status was also taken into account, there was a higher median percentage of white cells typed as ragocytes (57.5%) in DR4+/DQB1*0301+ patients.

Figure 1 (A) Ultrasonography of a metatarsoophalangeal (MTP) joint before injection. The distended capsule is seen (black arrows). (B) Ultrasonography of MTP joint after injection of sterile air (white arrows) and 2 ml steroid. Only the volume of air that is in the needle itself is injected (~0.05 ml).
Table 1  Synovial fluid cytology in patients with rheumatoid arthritis subdivided into different HLA-DR4 and/or DQB1*0301 groups

<table>
<thead>
<tr>
<th>Patients n=numbers</th>
<th>White blood cells (cm³)</th>
<th>Ragocytes (%)</th>
<th>Polymorphs (%)</th>
<th>Lymphocytes (%)</th>
<th>Monocytes (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>a. DQB1*0301+ (n=25)</td>
<td>8.74</td>
<td>50</td>
<td>73</td>
<td>6</td>
<td>2</td>
</tr>
<tr>
<td>Interquartile range</td>
<td>3.6–16.5</td>
<td>2.5–80</td>
<td>0–87.5</td>
<td>0–19.5</td>
<td>0–8</td>
</tr>
<tr>
<td>b. DQB1*0301– (n=20)</td>
<td>5.75</td>
<td>10</td>
<td>64</td>
<td>15.5</td>
<td>6.5</td>
</tr>
<tr>
<td>Interquartile range</td>
<td>2.9–12.0</td>
<td>0–40</td>
<td>9.3–85.8</td>
<td>3.25–37.8</td>
<td>1.3–13.3</td>
</tr>
<tr>
<td>Comparing a with b (p value)</td>
<td>0.3</td>
<td>0.033*</td>
<td>0.78</td>
<td>0.11</td>
<td>0.10</td>
</tr>
<tr>
<td>c. DR4+/DQB1*0301+ (n=20)</td>
<td>8.5</td>
<td>57.5</td>
<td>71.5</td>
<td>4</td>
<td>1.5</td>
</tr>
<tr>
<td>Interquartile range</td>
<td>3.7–18.1</td>
<td>10–85</td>
<td>0–86</td>
<td>0–19.5</td>
<td>0–6</td>
</tr>
<tr>
<td>d. DR4–/DQB1*0301– (n=6)</td>
<td>6.7</td>
<td>27.5</td>
<td>70.5</td>
<td>3.5</td>
<td>3.5</td>
</tr>
<tr>
<td>Interquartile range</td>
<td>0.3–10.9</td>
<td>0–40</td>
<td>0–79</td>
<td>0–12</td>
<td>0–6.5</td>
</tr>
<tr>
<td>Comparing c with d (p value)</td>
<td>0.66</td>
<td>0.27</td>
<td>0.98</td>
<td>1.00</td>
<td>0.84</td>
</tr>
<tr>
<td>e. DR4+/DQB1*0301– (n=14)</td>
<td>5.8</td>
<td>3.8</td>
<td>59</td>
<td>22.5</td>
<td>8.5</td>
</tr>
<tr>
<td>Interquartile range</td>
<td>3.3–9.2</td>
<td>0–30</td>
<td>13.5–83</td>
<td>4.5–40</td>
<td>3–14.5</td>
</tr>
<tr>
<td>Comparing c with e (p value)</td>
<td>0.34</td>
<td>0.017*</td>
<td>0.99</td>
<td>0.02*</td>
<td>0.027*</td>
</tr>
<tr>
<td>f. DR4–/DQB1*0301+ (n=5)</td>
<td>9.2</td>
<td>5</td>
<td>74</td>
<td>12</td>
<td>4</td>
</tr>
<tr>
<td>Interquartile range</td>
<td>3.6–13.9</td>
<td>0–52.5</td>
<td>21–84.5</td>
<td>3.75–16.5</td>
<td>1–7</td>
</tr>
<tr>
<td>Comparing d with f (p value)</td>
<td>0.66</td>
<td>0.93</td>
<td>0.66</td>
<td>0.43</td>
<td>0.66</td>
</tr>
</tbody>
</table>

*Significant on Mann-Whitney U test uncorrected, p corr>0.05 for all four p values.

(n=20) than in those who were DR4+/DQB1*0301– (3.8%, n=14, p=0.017, p corr>0.05). The DR4+/DQB1*0301+ group also had a reduced median lymphocyte percentage (4%) and monocyte count (1.5%) in comparison with DR4+/DQB1*0301– patients (22.5%, p=0.02, 8.5%, p=0.027, p corr>0.05). Table 1 summarises these results.

There were no statistical differences in synovial fluid cytology depending on whether the patients were positive or negative for rheumatoid factor, those with erosive or non-erosive disease, presence or absence of extra-articular features, and treatment with or without DMARDs and/or steroids.

Even though the corrected p values were not significant, this study suggests that there is a genetic influence on the expression of synovial fluid cytology in patients with RA. The presence of HLA-DR4 together with DQB1*0301 leads to an increase in ragocyte cells in the synovial fluid and a reduced lymphocyte and monocyte count. We speculate that the increase in activity of joint disease, and hence severity of the disease, might be due to the presence of excess ragocytes in the synovial fluid or to the loss of dampening effect owing to the reduced number of lymphocytes/monocytes. Ragocytes are leukocytes with intracytoplasmic granules and their function is unknown. In contrast, there was an increase in the percentage of lymphocytes in the synovial fluid of patients who were positive for the rheumatoid epitope. Once again the type of lymphocytes was unknown, but we can speculate that the presence of HLA-DR4 must have been the influencing allele because there were no significant differences in the cell counts when patients who were positive or negative for HLA-DR4 were compared.

The patients in this study were a selected group as they were hospital attenders with joint effusions requiring aspirations and thus had more severe disease than patients without joint effusions who could be managed as outpatients. In this selected group we found no association between synovial fluid cytology and the presence or absence of rheumatoid factor or extra-articular features, erosive or non-erosive disease, and the treatment with DMARDs and/or steroids. This is perhaps expected as patients with negative rheumatoid factor or extra-articular features, erosive or non-erosive disease, and the treatment with DMARDs and/or steroids can develop severe joint disease.

Although the presence of extra-articular features contributes to the overall morbidity of RA, it is not always associated with severe articular disease. Finally, our treatment of patients with DMARDs with and without steroids does not depend on the severity of articular disease alone and is based on the overall activity of the rheumatoid disease. On the other hand, treatment with DMARDs and steroids may modify synovial fluid findings, but we cannot draw any conclusions from this study as it is a cross sectional study and previous treatment was not taken into account.

Thus we have shown that synovial fluid cytology is influenced by genetic factors, but larger studies are needed to confirm these results. Further studies at the molecular level will help to determine the exact mechanism of action by which the presence or absence of certain cell types has an effect on the joint pathology of patients with RA. These studies may throw light on the pathogenesis of the joint disease in these patients.

EULAR recommendations for the management of knee osteoarthritis

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