Leader

Mechanisms of viral pathogenesis in rheumatic disease

Considerable evidence indicates that viruses may be important environmental factors in the pathogenesis of autoimmune rheumatic diseases. A concordance rate of 25% for the most common illnesses, such as rheumatoid arthritis (RA) or systemic lupus erythematosus (SLE) in monozygotic twins shows that genetic factors influence susceptibility to autoimmune diseases. Alternatively, a 70% discordance rate emphasises the importance of environmental factors. Forensic studies of archeological sites revealed the presence of RA-like erosive bone changes in pre-Columbian New World populations dating back 6500 years and the absence of RA in the Old World before the 18th century. This geographical distribution suggests that RA may have spread from the Americas through environmental factors, possibly by a virus, another microorganism, or an antigen. Viruses can elicit acute or subacute and, less often, chronic forms of arthritis. These viral arthritis syndromes can be diagnosed by recognition of well defined clinical signs and detection of viral antibodies and nucleic acids. Viral elements may also play a part in the pathogenesis of idiopathic autoimmune rheumatic diseases. This editorial will assess mechanisms of viral pathogenesis in rheumatic disease by focusing on known viruses capable of causing inflammatory arthritis syndromes and comparing virally induced immunological aberrations with those noted in rheumatic disease patients.

Well defined virus induced rheumatic diseases

Viral infections often lead to inflammatory syndromes where arthralgias or arthritis may represent a major manifestation. Most cases of viral arthritis, such as rubella or parvovirus B19 arthropathies are short-term and self limited as a result of an efficient elimination of the organism by the immune system. Chronic arthropathies have been associated with persistent or latent viral infections, virus induced autoimmunity, polyclonal B cell activation, and immunodeficiency resulting in opportunistic infections, largely because of an inability of the immune system to eliminate the pathogen. This latter group of viruses include human immunodeficiency virus 1 (HIV-1), human T-cell lymphotropic virus type I (HTLV-I), and hepatitis C virus (HCV).

VIRUS INDUCED TRANSIENT ARTHRITIS SYNDROMES

Parvovirus B19

Parvovirus B19 is one of the most frequent causes of viral arthritis. Joint manifestations are temporally associated with production of anti-B19 IgM antibodies. While involvement of B19 has been repeatedly raised in classic RA, large surveys failed to demonstrate an association between erosive RA and parvovirus B19.

Rubella virus

Rubella is known to cause mild and self limited arthralgias and acute arthritis. Chronic arthropathy was reported in 1–4% of postpartum female recipients of the RA27/3 vaccine strain. Other studies found no increase of chronic arthritis in women receiving the RA27/3 rubella vaccine. Moreover, no rubella virus can be recovered from peripheral blood lymphocytes of persons with chronic arthropathy following rubella infection or vaccination. Therefore, continued vaccination of rubella susceptible women to reduce the risk of congenital malformations seems warranted.

Alphaviruses

These are arthropod borne viruses that include the chikungunya, o’nyong-nyong, Mayaro, Sindbis, Okelbo, Barmah Forest (BF) and Ross River (RR) viruses. Similar to the rubella virus, they belong to the Togavirus family containing a positive strand RNA genome. The viruses are spread by mosquitoes in endemic areas of Australia (Sindbis, BF, and RR), South America (Mayaro), northern Europe (Okelbo), Asia and sub-Saharan Africa (chikungunya). They can cause an acute infectious illness with rash, fever, arthritis, myalgia and/or encephalitis. RR virus is the aetiological agent of the best studied epidemic polyarthritis (EPA) affecting up to 7800 Australians annually. Persistent infections are believed to be responsible for chronic arthritis. CD4+ T cells dominate mononuclear synovial effusions of EPA patients in contrast with CD8+ T cell infiltration in rashes of RR virus infected patients who made early and complete recoveries. EPA involves the small joint of the hand and often causes tendonitis. Symptoms may persist for months. No erosive changes have been reported. The diagnosis is made by demonstrating IgM antibodies to RR virus.

Adenoviruses

These viruses are a common cause of acute respiratory infections. Symmetric polyarthritis of small and large joints may occur within a week of respiratory symptoms. Recurrent chronic oligoarthritis because of adenovirus infection was rarely reported.

Coxsackieviruses

They belong to the group of enteroviruses. More than 90% of coxsackievirus infections are asymptomatic or manifest in undifferentiated febrile illness. Spectrum and severity of disease manifestations vary with age, sex, and immune status of the host. Coxsackievirus arthritis, usually caused by Group B virus, occurs with fever, serositis, pleurodynia, and rash. The arthritis is usually symmetric and polyarticular, involving both small and large joints.

Herpesviruses

After initial infection, viruses of the Herpesviridae family persist in the host with lifelong latency. Therefore, several of these viruses have been considered as aetiological agents in autoimmune diseases, such as systemic lupus erythematosus (SLE), RA, or Sjögren’s syndrome (see below). Epstein-Barr virus (EBV) infection causes arthralgias lasting for up to four months in 2% of patients with mononucleosis. Recently, EBV positive lymphomas were described in methotrexate treated RA patients. Interestingly, remission of lymphomas was noted after discontinuation of methotrexate. HSV-1 arthritis rarely lasts longer.
than two weeks. Varicella-zoster virus (VZV) can cause mononucleosis, mostly in the knee, as a rare complication of chickenpox.19 Cytopathic virus (CMV) may be responsible for scleroderma-like changes in patients with chronic graft versus host disease.20 These patients carry CD13 autoantibodies that bind to skin and mucous membranes. CMV incorporates the human CD13 protein into their viral envelope that may be responsible for generation of autoantibodies.

**Hepatitis B virus (HBV)**

HBV can cause arthralgias and arthritis early after infection. Arthritis resolves in 2–6 weeks with the onset of jaundice. Hepatitis B virus has been also associated with polyarteritis nodosa and cryoglobulinaemia.

**VIRUS INDUCED CHRONIC RHEUMATIC DISEASES**

**Hepatitis C virus (HCV)**

HCV has a wide pathogenic potential that is not limited to diseases of the liver.41 Despite high titre antibody concentrations, > 80% of infected people become chronic virus carriers. Cryoglobulinaemia is detectable in up to 40–50% of HCV infected patients.22 Identification of HCV as the causal agent of most (>90%) type II or essential mixed cryoglobulinaemias (EMC) has been a major breakthrough of rheumatology in the past decade.23 Type II cryoglobulins are immune complexes comprised of a monoclonal IgM/k rheumatoid factor and polyclonal IgG. The clinical syndrome of EMC is an immune complex vasculitis characterised by purpura, arthralgias, inflammatory arthritis, peripheral neuropathy, and glomerulonephritis.41 IgM/k bearing B cells are clonally expanded in the peripheral blood of EMC patients.42 HCV infection by HCV may be directly responsible for the clonal expansion of B cells,26 which may lead to development of B-cell non-Hodgkin’s lymphomas.23 HCV infection is associated with production of autoantibodies. Up to 75% of the patients have high titre rheumatoid factors, presumably produced by HCV infected and clonally expanded B lymphocytes. Fifty per cent or more of the patients have anti-smooth muscle antibodies. Low titre antinuclear antibodies and anticardiolipin antibodies were noted in 10–30% of HCV infected patients.50 Five per cent of patients may develop Sjögren’s syndrome, SLE, autoimmune thyroiditis, or scleroderma.27 Erosive/rheumatoid arthritis, and polymyositis/dermatomyositis were rarely documented.21

**Human T cell lymphotropic virus I (HTLV-I)**

Infection by HTLV-I has been associated with adult T cell leukaemia (ATL), mycosis fungoides/Sézary syndrome, HTLV-I associated myelopathy/tropic spastic paraparesis (HAM/TSP), HTLV-I associated arthritis (HAA), polymyositis, and Sjögren’s syndrome.28 Despite very high rates of infection in endemic areas where 30% or more of the population may be infected, relatively few (<1%) infected people show disease manifestations attributable to HTLV-I. The lifetime risk of developing a HTLV-I associated disorder is less than 5%. Polymyositis, Sjögren’s syndrome, and inflammatory arthritis may occur in the absence of leukaemia. They are characterised by infiltration of the skeletal muscle, salivary glands, or synovium with HTLV-I infected T lymphocytes. HAA is characterised by erosive symmetrical polyarthritis most often involving the hands. The patients may have rheumatoid factor or antinuclear antibodies and usually satisfy the diagnostic criteria for RA.29 Arthritis studies revealed proliferative synovitis in HAA.28 T cells infiltrating the joint have indented cerebriform nuclei similar to those seen in ATL. Prevalence of RA is increased in the HTLV-I infected population (0.56%) with respect to the uninfected population of Japan (0.31%).29 Thus, the relatively low disease frequency in virus-infected people strongly advocates for the role of factors other than HTLV-I in the development of RA. Transgenic mice carrying the tax transactivator gene of HTLV-I develop Sjögren’s syndrome and rheumatoid-like arthritis, indicating a pathogenic role for the p40/tax protein.28 HTLV-I was also shown to induce polymyositis, arthritis, and uveitis in rhesus macaque monkeys,30 thus establishing a primate model of viral arthritis.

**Human immunodeficiency virus 1 (HIV-1)**

During the course of HIV-1 infection three major phases can be distinguished. Within a few weeks after infection, extensive viraemia occurs giving rise to an acute mononucleosis-like syndrome. A second and relatively latent period represents an ongoing fierce battle between virus replication and replenishing of the CD4 T cell reservoirs. On average, 10 years after infection, diminished CD4 T cell function gives rise to opportunistic infections, lymphomagenesis, and autoimmunity at the final stages of disease. Polyclonal B cell activation and production of autoantibodies have been attributed to a shift from Th1 type to Th2 type helper T cell predominance (see below). Rheumatic diseases most commonly noted in patients with AIDS include Reiter’s syndrome, psoriatic arthritis, spondylarthropathies, and diffuse infiltrative lymphocytosis syndrome (DILS). Interestingly, all of these syndromes have been associated with relative expansion of CD8 T cells, thus suggesting that HIV-1 infection accelerates HLA class I restricted CD8 T cell mediated autoimmunity.31 In turn, SLE, RA, and polymyositis, which are thought to be mediated by CD4 T cells, remit in some patients after infection by HIV-1.32

**Caprine arthritis-encephalitis virus (CAEV)**

Like HIV-1, CAEV belongs to the lentivirus subfamily of the Retroviridae. These viruses cause multiorgan diseases that are apparent only months or years after infection. Visna virus, the prototype non-primate lentivirus, causes pneumonitis and progressive demyelinating neurological disease in the sheep.33 CAEV, which is closely related to visna virus, causes neurological diseases in young animals and chronic arthritis and mastitis in adult goats.34 35 Primary host cells of CAEV are monocytes and macrophages. Up to 40% of infected goats develop chronic arthritis characterised by infiltration of the synovium by macrophages, B lymphocytes, plasma cells, CD4+ and CD8+ T cells. CAEV induced arthritis leads to erosions of articular surfaces. CD4+ T cells infiltrating the arthritis joints are predominantly of T helper type 2.36 Similar to RA, tumour necrosis factor α (TNFα) concentrations are increased in synovial fluid of CAEV infected arthritis goats.37 Many naturally or experimentally infected goats are long term non-progressors, characterised by relatively low virus loads and a dominance of viral envelope protein specific Th1 type CD4+ T helper cells in the peripheral blood.38 (Table 1).

**Viral pathogenesis in idiopathic autoimmune diseases**

Independent lines of evidence suggest a viral aetiology in autoimmune rheumatic diseases. The possibility of a viral aetiology was raised by findings of virion-like tubuloreticular structures in endothelial cells and lymphocytes as well as demonstration of increased serum concentrations of type I
interferon (IFN) in lupus patients.38 Virus-like particles were also noted in RA synovium.39 Many viral infections are accompanied by production of autoantibodies and viral proteins have profound effects on both antigen presentation and effector functions of the immune system. Disregulation of programmed cell death has been reported in HIV infected40 VZV, varicella-zoster virus; EBV, Epstein-Barr virus = human herpesvirus 4; HBV, hepatitis B virus; HCV, hepatitis C virus; HTLV-I, human T cell lymphotropic virus immune patients have not been successful.43 Nevertheless, it is ecton functions of the immune system. Dysregulation of proteins have profound eaccompanied by production of autoantibodies and viral pro-

When normal conditions, the immune system develops a|potential virus specific immune response that rapidly eliminates the virus with only minimal tissue injury. Only minimal amounts of self antigens are released, which are insufficient to induce autoreactive B and T lymphocytes and autoimmune disease will not ensue. However, in the event that the host and the virus share antigenic determinants, virus infection may result in autoimmunity as virus specific T cells and antibodies are cross reactive with self antigens. This scenario does not preclude the possibility that the infecting virus is eliminated by the immune response. Alternatively, similarities between proteins of the major histocompatibility complex (MHC) and microbial antigens, especially viral antigens, may allow the host to regard an infectious agent as self and, therefore, forego an immune response. The “shared epitope” QKRAA sequence from the third hypervariable region of HLA DRB1*0401, which has been found in numerous human pathogens, is associated with susceptibility to RA.52 The 70K protein of U1snRNP was the first lupus autoantigen shown to contain a region of homology and immunological cross reactivity with a conserved p30 gag protein of most mammalian type C retroviruses (table 2). Based on this observation, Query and Keene proposed that autoimmunity to U1RNP may be triggered by expression of an endogenous retroviral gag protein. Anti-gag antibodies elicited by the ERS could cross react with the 70K protein and, subsequently, recognition could expand to additional 70K epitopes.

The ERS capable of triggering antibodies cross reactive with the 70K protein may correspond to HRES-1, a human T-cell lymphotropic virus related endogenous retrovirus.53 HRES-1 antibodies may be as high as 52%47 or as low as 21%49 in patients with SLE.50

Below, two possible mechanisms of viral pathogenesis will be discussed. The first scenario involves molecular mimicry causing abnormal self reactivity.51 Naturally, viral infections elicit potent antiviral immunity that may lead to cross reactivity against self antigens. Analysis of molecular mimics that is delineation of autoantigenic epitopes of self antigens may provide clues to the identity of viral antigens responsible for triggering the cross reactive immune responses. Secondly, infection of genetically susceptible hosts by a potentially large number of commonly occurring viruses may lead to T and B cell dysfunction and autoimmunity. Immunoregulatory aberrations triggered by well defined viral proteins at the level of antigen presentation, modulation of cytokine activities, and disruption of cell death pathways, will be discussed.

<p>| Table 1 Virus induced rheumatic diseases |</p>
<table>
<thead>
<tr>
<th>Virus</th>
<th>Arthritis frequency</th>
<th>Arthritis type</th>
<th>Duration</th>
<th>Erosion</th>
<th>Other</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Parvovirus B19</td>
<td>Children: 5–10% Adults: 50–70% Female:male = 2:1</td>
<td>Polynodular, small and large joints, symmetrical</td>
<td>2–8 weeks, rarely chronic</td>
<td>No</td>
<td>Anemia</td>
<td>(3) (90)</td>
</tr>
<tr>
<td>Rubella</td>
<td>10–30%</td>
<td>Multiple small joints</td>
<td>5–10 days</td>
<td>No</td>
<td>Vaccine strain</td>
<td>(4,43)</td>
</tr>
<tr>
<td>VZV</td>
<td>&lt;1%</td>
<td>Mononodular arthritis</td>
<td>1–7 days</td>
<td>No</td>
<td>Life long latency</td>
<td>(19)</td>
</tr>
<tr>
<td>EBV</td>
<td>1–5%</td>
<td>Poly or mononodular arthritis</td>
<td>1–12 weeks</td>
<td>No</td>
<td>Autoantibodies</td>
<td>(16)</td>
</tr>
<tr>
<td>HBV</td>
<td>10–25%</td>
<td>Symmetrical, migratory</td>
<td>1–3 weeks</td>
<td>No</td>
<td>Vasculitis</td>
<td>(43)</td>
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<td>HCV</td>
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<td>(23)</td>
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<td>HTLV-I</td>
<td>&lt;1%</td>
<td>Oligonodular arthritis, large joints</td>
<td>Chronic</td>
<td>Yes</td>
<td>sjögren’s</td>
<td>(28)</td>
</tr>
<tr>
<td>HIV-1</td>
<td>10–50%</td>
<td>Painful joint syndrome</td>
<td>1–2 days</td>
<td>No</td>
<td>promotion of CD8 T cell</td>
<td>(32)</td>
</tr>
<tr>
<td>Alphaviruses</td>
<td>&gt;50%</td>
<td>Reiter’s syndrome</td>
<td>Chronic</td>
<td>Yes</td>
<td>mediated autoimmunity</td>
<td></td>
</tr>
<tr>
<td>Adenovirus</td>
<td>rare</td>
<td>Symmetrical, small joints</td>
<td>1 week to months</td>
<td>No</td>
<td>fever, myalgia encephalitis</td>
<td>(11)</td>
</tr>
<tr>
<td>Coxsackievirus</td>
<td>rare</td>
<td>Small and large joints</td>
<td>transient or chronic</td>
<td>No</td>
<td>Pharyngitis</td>
<td>(12–14)</td>
</tr>
<tr>
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<td>Arthritis type</td>
<td>Duration</td>
<td>Erosion</td>
<td>Other</td>
<td>Reference</td>
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VZV, varicella-zoster virus; EBV, Epstein-Barr virus = human herpesvirus 4; HBV, hepatitis B virus; HCV, hepatitis C virus; HTLV-I, human T cell lymphotropic virus 1; HIV-1, human immunodeficiency virus 1.

Table 2 Molecular mimicry between viral proteins and autoantigens

<table>
<thead>
<tr>
<th>Autoantigen</th>
<th>Prevalence (%)</th>
<th>Viral protein</th>
<th>Virus</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>70kU1 snRNP</td>
<td>30</td>
<td>gag</td>
<td>MoMLV, HRES-1</td>
<td>(47,53)</td>
</tr>
<tr>
<td>HRES-1</td>
<td>21–52</td>
<td>gag, gag24</td>
<td>HRES-1</td>
<td>(46–49)</td>
</tr>
<tr>
<td>Sm B’</td>
<td>30</td>
<td>gag, gag24</td>
<td>MoMLV, HRES-1</td>
<td>(59)</td>
</tr>
<tr>
<td>C/1 snRNP</td>
<td>30</td>
<td>ICP4</td>
<td>HIV-1</td>
<td>(64)</td>
</tr>
<tr>
<td>Sm B</td>
<td>36</td>
<td>EBNA-1, EBV</td>
<td>EBV</td>
<td>(60)</td>
</tr>
<tr>
<td>Sm B’</td>
<td>25–40</td>
<td>EBNA-1</td>
<td>EBV</td>
<td>(63)</td>
</tr>
<tr>
<td>La</td>
<td>15</td>
<td>gag</td>
<td>HSV</td>
<td>(65)</td>
</tr>
<tr>
<td>p542</td>
<td>10–50</td>
<td>EBNA-1</td>
<td>EBV</td>
<td>(61)</td>
</tr>
<tr>
<td>ERV-3</td>
<td>32</td>
<td>env</td>
<td>MoMLV</td>
<td>(50)</td>
</tr>
</tbody>
</table>

Prevalence of antibodies in patients with SLE.
of scleroderma, 44% (8 of 18) of primary SJS, 19% (3 of 16) of polymyositis/dermatomyositis patients also had HRES-1 antibodies. By contrast 3.6% (4 of 111) of normal donors and none of 42 patients with AIDS or 50 asymptomatic HIV infected patients had HRES-1 antibodies. The retroviral gag related region of the 70K protein shares three consecutive highly charged amino acids, Arg-Arg-Glu (RRE), an additional Arg and functionally similar Arg/Lys residues with HRES-1/p28 that represent cross reactive epitopes between the two proteins. Interestingly, the RRE triplet is repeated three times in the 70K protein at residues 248–250, 418–420, and 477–479, respectively (GenEmbl accession number X04654). This suggests that recognition of the retroviral domain may lead to epitope spreading through binding to RRE triplets within the 70K protein. It is well known that highly charged polypeptides can elicit high titre antibodies. Therefore, the presence of charged amino acids in the mimicking epitopes may have important implications in triggering cross reactive antibodies of high affinity. HRES-1 is represented as a single copy element per haploid genome that has been mapped to a common fragile site of chromosome 1 at q42.56. The presence or absence of a polymorphic HindIII site defines two different allelic forms of the HRES-1 genomic locus. In comparison with normal blood donors, a differential segregation of polymorphic genotypes of the HRES-1 locus—that is, a relative decrease of genotype I and increase of genotype III were noted among patients with SLE. The q41→q42 region of chromosome 1 was found to contain susceptibility genes that confer risk for SLE in multiple ethnic groups, further supporting the notion that HRES-1 or another gene(s) closely linked to HRES-1 may influence susceptibility to SLE.

A mimicking epitope between another lupus autoantigen, Sm, and HIV-1 p24 gag was defined based on cross reactivity with monoclonal antibody 4B4. A proline rich domain present in both the Bβ' subunit of Sm and HIV p24 gag was suggested to be the core of cross reactive epitopes. Antibodies binding to HIV-1 p24 gag were found in 22 of 61 patients with SLE. A region of considerable homology, comprised of 11 highly charged residues (GRGGRGRGGRG), was identified as a site of cross reactivity between the D component of Sm and the Epstein-Barr virus nuclear antigen 1 (EBNA-1). Mimicry between EBNA-1 and another self protein, the 71 kDa p542 has been shown in patients with SLE and other autoimmune diseases. The mimicking epitope, a 28-mer glycine-rich sequence was selectively recognised by serum samples from autoimmune patients while it was uncommonly targeted by serum samples from normal donors. The concept that EBV can trigger IgG antibodies that cross react with autoantigens is an attractive one. EBV is a ubiquitous human DNA virus that infects B cells and causes their polyclonal activation and thus polyclonal antibody production. Such polyclonal B cell activation may be an early step in pathogenesis of SLE. Interestingly, prevalence of EBV infection was reported to be as high as 99% in young SLE patients in comparison with a 70% prevalence in controls. Therefore, EBV has the potential to trigger lupus by two mechanisms: polyclonal B cell activation and molecular mimicry. The ICP4 protein of another ubiquitous human DNA virus, human herpesvirus type I (HHV-1) shows cross reactivity with the C component of U1 snRNPs. A region with limited sequence homology to feline sarcoma virus (FSV) gag protein was noted in the La antigen. Antibodies to the env protein of an ERS, ERV-3, were reported in patients with SLE with the highest prevalence in mothers of babies with complete heart block (CHB).

ERS, in addition to serving as cross reactive targets of antiviral immunity, may also have a direct role in regulating immune responses. A synthetic heptadecapeptide (CKS-17) corresponding to the transmembrane domain of the env protein conserved among many exogenous and endogenous retroviruses has potent immunosuppressive properties (table 3). ERS and other retrotransposable elements possess a relatively high mobility and thus represent a major factor in shaping and reorganisation of the eukaryotic genome. The ERS HERV-K10 was found to have an integration site in the human complement C2 gene. Variable repeats of this element may have a role in polymorphism and differential expression of C2 loci. Integration of a 5.3 kb ETn retrotransposon in the FasR gene

### Table 3 Viral proteins mimic immunoregulatory abnormalities of rheumatic disease

<table>
<thead>
<tr>
<th>Virus/viral product</th>
<th>Mechanism/effect</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Change in lymphokine milieu</td>
<td>induction of IFNs</td>
<td>(108)</td>
</tr>
<tr>
<td>HTLV-I p40/ tax</td>
<td>increases IL2R expression</td>
<td>(109)</td>
</tr>
<tr>
<td>stimulates apoptosis through bcl2</td>
<td>(98)</td>
<td></td>
</tr>
<tr>
<td>HIV-1 gp41 (aa591-597)</td>
<td>induces apoptosis through Fas and oxidative stress</td>
<td>(99,100)</td>
</tr>
<tr>
<td>inhibits PKC activity and IL2 production</td>
<td>(75)</td>
<td></td>
</tr>
<tr>
<td>HIV-1 tat</td>
<td>binds Ick, inhibits Th1 cytokine production</td>
<td>(110)</td>
</tr>
<tr>
<td>HIV-1 tat</td>
<td>inhibits Th1 cytokine IL12 production</td>
<td>(111)</td>
</tr>
<tr>
<td>CMV US28</td>
<td>binds C-C chemokines</td>
<td>(76)</td>
</tr>
<tr>
<td>CKS-17 env domain</td>
<td>inhibits Th1 cytokine production</td>
<td>(66)</td>
</tr>
<tr>
<td>Altered antigen presentation</td>
<td></td>
<td></td>
</tr>
<tr>
<td>EBV/BZLF2</td>
<td>Inhibits MHC class II binding</td>
<td>(81)</td>
</tr>
<tr>
<td>Adenovirus/E1A</td>
<td>Inhibits MHC class I expression</td>
<td>(80)</td>
</tr>
<tr>
<td>Adenovirus/E3 gp19</td>
<td>Inhibits MHC class I binding to antigen</td>
<td>(79)</td>
</tr>
<tr>
<td>HSV/CIP47</td>
<td>Inhibits TAP (MHC) function</td>
<td>(78)</td>
</tr>
<tr>
<td>Apoptosis pathways</td>
<td></td>
<td></td>
</tr>
<tr>
<td>HTLV-I p40/ tax</td>
<td>stimulation through blocking bcl-2</td>
<td>(98)</td>
</tr>
<tr>
<td>HTLV-I p40/ tax</td>
<td>inhibition of Fas and oxidative stress pathways</td>
<td>(99,100)</td>
</tr>
<tr>
<td>HIV-1 tat</td>
<td>stimulation through Fas and oxidative stress</td>
<td>(84,86)</td>
</tr>
<tr>
<td>HIV-1 protease</td>
<td>Stimulation by cleavage of bcl-2</td>
<td>(87)</td>
</tr>
<tr>
<td>HIV-1 vpr</td>
<td>Stimulation through mitotic arrest</td>
<td>(93,94)</td>
</tr>
<tr>
<td>Papovavirus B19</td>
<td>Stimulation by unknown mechanism</td>
<td>(88)</td>
</tr>
<tr>
<td>Influenza A SN1</td>
<td>Stimulation through Fas and bcl-2 pathways</td>
<td>(89)</td>
</tr>
<tr>
<td>Adenovirus E1B 19K</td>
<td>Bcl-2 homologue, inhibits apoptosis</td>
<td>(95)</td>
</tr>
<tr>
<td>EBV BHRF1</td>
<td>Bcl-2 homologue, inhibits apoptosis</td>
<td>(96)</td>
</tr>
<tr>
<td>HSV 34.5</td>
<td>Inhibition by unknown mechanism</td>
<td>(97)</td>
</tr>
<tr>
<td>HBV pX</td>
<td>Inhibition, p53 antagonist</td>
<td>(92)</td>
</tr>
<tr>
<td>EHV-2 23 K E8 FLIP</td>
<td>Inhibition of Fas pathway</td>
<td>(104,105)</td>
</tr>
<tr>
<td>MCV ORF159L FLIP</td>
<td>Inhibition of Fas pathway</td>
<td>(104,105)</td>
</tr>
<tr>
<td>HVS ORF71+ FLIP</td>
<td>Inhibition of Fas pathway</td>
<td>(104,105)</td>
</tr>
<tr>
<td>HHV8 ORF199+ FLIP</td>
<td>Inhibition of Fas pathway</td>
<td>(104,105)</td>
</tr>
</tbody>
</table>
locus resulted in dysregulation of this apoptosis pathway in lupus prone MRL/lpr mice.69

**VIRAL PROTEINS MODULATE CYTOKINE PRODUCTION**

Functional abnormalities of T and B cells have been correlated with an altered cytokine production profile in patients with rheumatic disease.70 Secretion of T helper type 1 (Th1) cytokines, interleukin 2 (IL2), interferon γ (IFNγ), and IL12, necessary for maintenance of a classic T cell mediated immunity, is diminished while production of Th2 cytokines, in particular, IL4, IL5, IL6 and IL10, promoting mediated immunity, is diminished while production of Th2 cytokines increase. This marked shift in cytokine production may be related to a fundamental biochemical defect manifested in deficiencies of protein kinase A activity, increased phosphatidylinositol turnover and diminished protein kinase C activities in lupus T cells.71

Changes in production of cytokines similar to those in patients with SLE, a shift from a Th1 to a Th2 type cytokine profile, have been described as a result of HIV-1 infection. CD4+ T cell decline is mediated by an increased rate of apoptosis or programmed cell death (PCD).70 Interestingly, Th1 type cytokines protect against apoptosis while Th2 cytokines increase PCD.72 Accelerated apoptosis has also been described in SLE. Moreover, apoptosis has been associated with a compartmentalised release of autoantigens in patients with SLE.73 These observations raise the possibility that increased apoptosis and autoantibody production may be mediated by somewhat similar mechanisms both in AIDS and SLE. The nef and tat genes of HIV-1 are thought to mediate a Th1 to Th2 shift in cytokine production (table 3). A synthetic enve heptadcapeptide, CKS-17 down regulates cell mediated immune responses,74 possibly via suppression of Th1 type cytokine production.75 The CKS-17 motif was found to be highly conserved among infectious and endogenous retroviruses.76 Down regulation of IL2 production in HIV infected cells was also linked with inhibition of protein kinase C (PKC) activity and influx of Ca++.75 US28 of HCMV binds members of the α or C-C family of chemokines.77 Polymorphisms of chemokine receptors are important factors determining sensitivity to infection by HIV-1.78 Development of CAEV induced arthritis is also associated with a Th1 to Th2 shift in cytokine production.79

**VIRAL PROTEINS MODULATE ANTIGEN PRESENTATION**

MHC haplotypes have been associated with susceptibility to rheumatic diseases and recognition of specific autoantigens. This is consistent with a dominant role of the MHC in selection and presentation of antigenic peptides. Interestingly, viral peptides influence expression of MHC class I and II antigens as well as the function of TAP proteins (transporters associated with antigen presentation). Thus, herpes simplex virus (HSV) encodes a cytosolic protein, ICP47, which interferes with the function of the TAP1/TAP2 complex, prevents association of peptide with MHC class I, and leads to degradation of empty class I molecules.80 E3 gp19 of adenovirus type 2 and E1A of adenovirus type 12 also inhibit expression of MHC class I.81 E1A was shown to directly interfere with negative regulatory elements in the MHC class I promoter.82 The BZLF2 protein of EBV recognises the peptide binding pocket of the HLA-DR β chain and interferes with class II directed antigen presentation.83 In addition, expression of MHC proteins is dependent on production of cytokines, such as IFNγ and TNFα, production of which is often inhibited by viral infections.84 85 The above data indicate that viruses commonly infecting the general population, including patients with rheumatic diseases, affect antigen presentation and modulate the cytokine milieu, which may in turn play a part in initiation or perpetuation of autoimmune disease.

**REGULATION OF APOPTOSIS BY VIRAL PROTEINS**

Apolipoprotein or programmed cell death (PCD) represents a physiological mechanism for elimination of autoreactive lymphocytes during development. Viral infections may have a role in dysregulation of apoptosis in autoimmune patients. Many viruses have evolved genes that can selectively inhibit or stimulate PCD. The suicide of an infected cell by internal activation of apoptosis or the killing of an infected cell by a cytotoxic T lymphocyte or NK cell may be viewed as a defence mechanism of the host to prevent viral propagation. In the early stages of infection, viral inhibitors of apoptosis allow for more extensive production of progeny. At later stages, viral inducers of apoptosis facilitate spread of progeny to uninfected cells.

HIV may use several mechanism to deplete CD4+ T cells at the later stages of disease (table 3). The tat protein induces oxidative stress,86 87 and increases surface expression of the Fas ligand resulting in accelerated signaling through the Fas pathway.88 89 In addition, cleavage of bcl-2 by HIV protease may expose the cell to a variety of apoptotic signals.90 Parovirus B19 depletes erythroid progenitor cells by apoptosis.91 Cells infected by influenza virus undergo PCD that can be inhibited by bcl-2 and facilitated through the Fas pathway.92 It is intriguing to consider the possibility that viruses causing common cold may stimulate antinuclear autoantibody production through periodic release of nucleosomes from apoptotic cells. Thus, chronic parvovirus B19 infection is recently associated with production of a wide array of autoantibodies.93 Replication of CAEV is also associated with induction of apoptosis.94

![Diagram of Regulation of Apoptosis](https://example.com/diagram.png)

Figure 1 Regulation of apoptosis pathways by viral proteins. Oxidants, ultra violet light, and corticosteroids trigger apoptosis by mitochondrial damage, which, in turn, leads to the release of caspase activating factors. This process is inhibited by bcl-2 and its viral homologues. Release of reactive oxygen intermediates causes increased production of FasL and DNA fragmentation. Fas ligand (FasL) crosslinks the Fas receptor (Fas/Apo1/CD95), which recruits an adapter protein with a Fas associated death domain (FADD). Virus FLIP (vFLIP) possesses a viral FLIPs (vFLIPs) may also block TNF receptor mediated signalling through FADD shared by both the Fas and TNF pathways. Upon recruitment of caspase 8, its oligomerisation causes self cleavage and activation of downstream effector caspases. A caspase activated DNase cleaves chromosomal DNA. Oxidants and ultra violet light as well as ROI released from damaged mitochondria cause DNA fragmentation, which in turn activates p53. p53 induces oxidative stress and augments surface expression of the Fas receptor. HIV-1 tat increases mitochondrial ROI production thus increasing apoptosis. By contrast HBV e protein interferes with the proapoptotic effects of p53.
Inhibition of apoptosis by viral proteins help infected cells to evade inflammatory responses, such as killing by cytotoxic T cells through the Fas and TNF pathways (fig 1).10,11 X inactivating hepatitis B virus (HBV) inhibits binding of p53 to DNA.12 The γp protein of HIV-1 causes cells to arrest in the G2 phase of the cell cycle when virus expression is highest.13,14 Viral homologues of bcl-2 can functionally substitute for bcl-2 in binding to the apoptosis accelerator in HTLV-I/tax transgenic mice.103 A NADPH dependent antioxidant99 and inhibition of Fas signalling have been implicated in the anti-apoptotic effect of HTLV-I tax protein.120 These two mechanisms are not mutually exclusive as Fas induced cell death is accompanied by the formation of reactive oxygen intermediates (ROI) and is subject to regulation by enzymes of the pentose phosphate pathway providing NADPH as a source of reducing equivalent for intracellular antioxidants.100 p40/tax may mediate autoimmune arthropathy122 and Sjögren’s syndrome by blocking Fas dependent cell death in HTLV-I/tax transgenic mice.119 A new family of viral inhibitors, designated as vFLIPs (viral FLICE inhibitory proteins), has recently been discovered.123-125 vFLIPs are produced by several γ-herpesviruses, including the Kaposi-sarcoma associated human herpesvirus 8 (HHV-8), the tumorigenic human mucosa-associated lymphoid tissue (HVS), and equine herpesvirus 2 (EHHV-2). vFLIPs block the early signalling events triggered through the death receptors Fas, TRAMP, TRAIL-R and TNFR1. Thus, herpesviruses evolved a series of genes that allow selective blocking of the Fas and TNF signalling pathways.

Conclusion and future directions
The experimental evidence presented above shows immunological cross reactivities between autoantigens and viruses. The concept that autoimmunity is triggered in genetically susceptible hosts by viral environmental factors, possibly different from patient to patient, is consistent with the general epidemiology—that is, a relatively sporadic occurrence, of the disease.100 Moreover, proteins of commonly occurring viruses have profound effects on the cytokine milieu, antigen recognition, and lymphocyte cell survival. Thus, molecular mimicry and immunomodulation by viral proteins may account for both cross reactivity with autoimmune virus abnormal (HSV) and functions in autoimmune disorders. Causation of HCV and virus type II cryoglobulinemias and of HTLV-I with polymyxositis, Sjögren’s syndrome, and erosive arthritis represent significant discoveries for rheumatology. Patients infected with HCV or HTLV-I invariably have high titre antiviral antibodies. Likewise, serum samples of patients infected with HCV or HTLV-I invariably have high titre antiviral antibodies. Thus, molecular mimicry and immunity in rheumatoid arthritis. Role for the Epstein-Barr virus. Rev Rheum Engl Ed 1998;65:283–6.

Funding: this work was supported in part by grant RO1 DK 49223 from the National Institutes of Health, grant 1446A1 from the National Multiple Sclerosis Society, the Arthritis Foundation, and the Central New York Community Foundation.
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Acknowledgements
We thank Dr Katalin Banki for critical review of the manuscript and Dr Paul Phillips for continued encouragement and support.

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Ann Rheum Dis 1999 58: 454-461
doi: 10.1136/ard.58.8.454

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