Antibodies to histones in systemic lupus erythematosus: prevalence, specificity, and relationship to clinical and laboratory features

Michael G Cohen, K Michael Pollard, John Webb

Abstract
Antibodies to histones (AHA) are commonly found in patients with systemic lupus erythematosus (SLE). However, the full profile of AHA and their clinical associations remains unclear. A total of 111 patients with SLE were studied, including 13 patients in whom multiple serum samples were available over several years. IgM, IgG, and IgA antibodies to total core histones, histone complexes, and individual histones were determined by highly sensitive enzyme linked immunosorbent assays (ELISAs). Antibodies to histones were detected in 74% of serum samples, though only at low levels in half of these. Antibodies to each of the individual histones (H1, H2A, H2B, H3, H4) occurred with similar frequencies except for IgG and IgA antibodies to H4, which were uncommon. In contrast, antibodies to the histone complexes H2A-H2B and H3-H4 were detected in only two serum samples and thus do not appear to be a feature of SLE. All three major isotypes of AHA were common and usually occurred with similar frequencies to one another for the various histone specificities.

Patients and methods
PATIENTS AND SERUM SAMPLES
One hundred and eleven patients with SLE with serum samples stored at the department of rheumatology, Royal North Shore Hospital from 1970 to 1986 formed the basis of this study. All patients fulfilled the American Rheumatism Association revised criteria for SLE. All serum samples had been stored in aliquots at −20°C.

Antibodies to histones (AHA) are among the more common autoantibodies seen in patients with systemic lupus erythematosus (SLE). In addition, the known structures of the histones and the use of solid phase assays have allowed their nature to be defined in greater detail than most other autoantibodies. Nonetheless, many reports have conflicting findings: the prevalence of AHA in patients with SLE has been variously reported to be as low as 21% or up to 81%. Some workers have observed associations between AHA and indices of disease activity, whereas others have failed to show this. The specificity of AHA for individual histone components has also differed from study to study. In this work, we determined all the major isotypes of AHA (including antibodies to individual histones and histone complexes) in a large cohort of patients with SLE in both cross sectional and longitudinal studies. This has allowed the definition of the prevalence and clinical associations of AHA in our patients with SLE as well as addressing some of the factors that may have given rise to the disparities between earlier reports.

ASSAYS
The routine estimation of rheumatoid factor titre was obtained with a commercial sensitised sheep cell agglutination technique (Rheumaton, Denver Laboratories). Antinuclear antibodies were detected using rat liver substrate as pre-
vously described. Antibodies to native DNA were measured as the percentage binding in the Farr assay (normal value <20%). Antibodies to extractable nuclear antigens (SS-A/Ro, SS-B/La, RNP, Sm) were detected by counter-immunoelectrophoresis or immunodiffusion in agarose gel with rabbit thymus extract (Pel-Freeze Biologicals) or guinea pig kidney extract. Routine complement C3 and C4 levels were measured by radial immunodiffusion. Complement levels in the serum samples used for the determination of AHA and the CHS in the cross sectional study were measured again by rate nephelometry using a Beckman ICS nephelometer.

Antibodies to histone were measured by an enzyme linked immunosorbent assay (ELISA) as described previously. Briefly, a preparation that was found to contain core histones (H2A, H2B, H3, H4) and a preparation containing H1 were obtained from Sigma Chemical. Individual histones were purified by the method of Bohm et al. Histone complexes were prepared as previously described. ELISA plates were coated with histones diluted in phosphate buffered saline (PBS), pH 7.3 and then coated with PBS containing Tween 20 (PBST). Samples were diluted in PBST and bound antibody detected with enzyme conjugated antibodies to human IgM, IgG, or IgA (Silanes). Enzyme substrate was added and the colour change monitored with an automated spectrophotometer. Preliminary experiments determined that none of 30 AHA positive serum samples bound to PBST coated plates in the absence of antigen.

All serum samples were tested for antibodies to total core histones (AcHA) and H1. The upper limit of the normal level in all assays was defined as the mean plus three standard deviations of the absorbance of six serum samples from healthy blood donors, which were representative of 80 serum samples used to determine the normal range. These six serum samples and the one positive control were run in all assays. Units for AcHA were determined by ascribing unit values to the positive control serum sample (100 U for all isotypes) and to the upper limit of the normal level (12, 30, and 30 for IgG, IgM and IgA, respectively). Only serum samples with AcHA levels greater than twice the normal level were further tested for antibodies to individual core histones or histone complexes. Antibodies to histone complexes were stated to be present if the absorbance of the test serum sample was greater than twice the normal level and there was no increased binding to either of the component histones.

STATISTICAL METHODS
Possible associations between the antibodies and the characteristics of the disease for future prospective studies, were sought using the Pearson χ² test. The Spearman rank correlation was used to compare antibody levels. The relationships between each of the AHA, antibodies to native DNA, the erythrocyte sedimentation rate (Westergren) and the lupus activity criteria count in the serially studied patients were assessed for the whole group using logistic regression. In individual patients, the Mann-Whitney test was used.

Results
CLINICAL AND LABORATORY CHARACTERISTICS
The patient sample consisted of 105 women and six men with a median age of 38 years (range 15–70 years). The median duration of disease at the time the serum samples were collected was eight years (range 0–45 years). Table 1 compares the prevalence of each American Rheumatism Association criterion in these 111 patients with those from other studies.

The prevalences of other features recorded in these patients were: Raynaud’s phenomenon (68%), vasculitis (26%), antibodies to extractable nuclear antigen (52%), decreased complement C3 (41%) and C4 (74%), and rheumatoid factor (18%).

In the same serum samples used for the estimation of AHA, raised levels of antibodies to native DNA were observed in 32%, decreased complement C3 in 19%, and C4 in 15% of patients.

ANTIBODIES TO HISTONES
Increased levels of AcHA or antibodies to H1 of one or more isotypes were found in 74% of the 111 serum samples. Although AcHA were seen in 68% of serum samples, only 32% had AcHA of any isotype raised to greater than twice the normal level. Figure 1 illustrates the range of values of AcHA in the 111 serum samples.

Table 1 gives the prevalence of each isotype of AcHA and antibodies to H1. Most serum

<table>
<thead>
<tr>
<th>Table 1</th>
<th>Occurrence of the American Rheumatism Association (ARA) 1982 revised criteria for the classification of systemic lupus erythematosus in various patient groups</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Criterion</strong></td>
<td><strong>Prevalence (%)</strong></td>
</tr>
<tr>
<td><strong>This series</strong></td>
<td><strong>Japan (29)</strong></td>
</tr>
<tr>
<td>Malar rash</td>
<td>79</td>
</tr>
<tr>
<td>Discoid lupus</td>
<td>38</td>
</tr>
<tr>
<td>Photosensitivity</td>
<td>72</td>
</tr>
<tr>
<td>Mouth ulceration</td>
<td>38</td>
</tr>
<tr>
<td>Arthritis</td>
<td>95</td>
</tr>
<tr>
<td>Pleuritis/periarteritis nodosa</td>
<td>51</td>
</tr>
<tr>
<td>Renal disease</td>
<td>63</td>
</tr>
<tr>
<td>Central nervous system disease</td>
<td>46</td>
</tr>
<tr>
<td>Haematological</td>
<td>94</td>
</tr>
<tr>
<td>Immunological</td>
<td>95</td>
</tr>
<tr>
<td>Antinuclear antibodies</td>
<td>100</td>
</tr>
</tbody>
</table>

*Reference from which data were obtained.
†Exact figures could not be stated where the authors have separated individual criteria into their component parts (e.g. stated figures for pleurisy and pericarditis).
‡Renal disease includes patients with biopsy evidence of lupus nephritis. Central nervous system involvement includes all abnormalities (e.g. seizures, psychosis, coma, organic brain syndrome) considered by the doctor to be secondary to SLE and for which no other cause subsequently became evident.

<table>
<thead>
<tr>
<th>Table 2</th>
<th>Antibodies to total core histones and histone H1 in 111 patients with systemic lupus erythematosus</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Isotype(s)</strong></td>
<td><strong>AcHA</strong></td>
</tr>
<tr>
<td></td>
<td>(%)</td>
</tr>
<tr>
<td>IgG</td>
<td>49</td>
</tr>
<tr>
<td>IgM</td>
<td>46</td>
</tr>
<tr>
<td>IgA</td>
<td>37</td>
</tr>
<tr>
<td>IgG, IgM, or IgA</td>
<td>68</td>
</tr>
<tr>
<td>IgG, IgM, and IgA</td>
<td>21</td>
</tr>
</tbody>
</table>

*AcHA=Antibodies to core histones. The figures in brackets represent the percentage of serum samples with that isotype alone.
samples with AcHA or antibodies to H1 had more than one isotype. Additionally, all serum samples with IgG or IgA AcHA alone had only minimally raised levels. Similarly, 12 of the 15 serum samples with only IgM AcHA had low levels of antibody. The remaining three serum samples had moderately high levels.

Although IgG antibodies to H1 were present in all but one serum sample having any isotype of antibodies to H1, it was uncommon for a serum sample to have all three isotypes of antibodies to H1. In contrast, with IgG antibodies to H1, no serum sample had IgM or IgA antibodies to H1 alone. Strong correlations (p<0.001) were observed between the levels of each isotype of AcHA. Table 3 shows that antibodies to H2A, H2B, and H3 occurred in similar proportions, irrespective of the isotype; however, with the exception of IgM antibodies, antibodies to H4 were detected infrequently. Only a small number of serum samples (2% of the 111 samples) exhibited reactivity with histone complexes in the absence of antibodies to the component histones.

**ASSOCIATIONS OF ANTIBODIES TO HISTONES**

Table 4 shows that very few significant clinical or laboratory associations of AHA were noted. Although an association with psychiatric disturbance was documented, there was no overall association with central nervous system involvement. The associations with ulceration of the mouth and lymphopenia were detected when the three major isotypes were considered together, but no such associations existed with individual isotypes of AcHA. There were no associations with rheumatoid factor, C3, C4, or antibodies to extractable nuclear antigen. Only the association between IgG AcHA and raised levels of antibodies to native DNA had a p value less than 0.01.

**CHANGES IN ANTIBODIES TO HISTONE WITH TIME**

Of the 13 patients tested, only two had IgM AcHA with little or no IgG AcHA in the earliest serum samples tested. These two patients subsequently developed high levels of IgG AcHA (after 25 and 34 months), but high levels of IgM AcHA also persisted. Although antibodies to individual histones varied over time, in no patient was there a lasting change in the profile of antibodies (e.g. a switch from antibodies to H2A to antibodies to H3). Neither was there a profile of antibodies to the individual histones that was associated with active disease. Despite the paucity of IgG antibodies to H4 in the cross sectional study, each of the 13 patients studied over a long time period had IgG antibodies to H4 on one or more occasions.
Table 4  Clinical and laboratory associations of antibodies to histones

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Antibodies</th>
<th>Isotypes</th>
<th>( p ) Value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mouth ulceration</td>
<td>ACHAI</td>
<td>All</td>
<td>0.0188</td>
</tr>
<tr>
<td>Psychiatric5</td>
<td>ACHAI</td>
<td>IgA</td>
<td>0.0416</td>
</tr>
<tr>
<td>Antibodies to native DNA (&gt;20%)</td>
<td>ACHAI</td>
<td>IgA</td>
<td>0.0032</td>
</tr>
<tr>
<td></td>
<td>H1</td>
<td>IgA</td>
<td>0.0174</td>
</tr>
<tr>
<td>Lymphopenia</td>
<td>ACHAI</td>
<td>All</td>
<td>0.0448</td>
</tr>
</tbody>
</table>

* Pearson \( \chi^2 \) test.
† Antibodies to core histones.
‡ Raised IgG, IgM, or IgA antibodies.
§ Organic brain syndrome, psychosis, or severe affective disorder.

**Relationship between AHA, antibodies to native DNA, erythrocyte sedimentation rate, and disease activity in the longitudinal study**

When all the patients were considered, the periods of disease activity were associated with higher levels of IgG ACHA (\( p<0.05 \)), IgA ACHA (\( p<0.05 \)), and antibodies to native DNA (\( p<0.01 \), but not IgM AHA or erythrocyte sedimentation rate. However, there was no association between disease activity and any of these parameters when categorised as normal or abnormal. That is, raised levels of IgG and IgA ACHA as well as antibodies to native DNA were found when the SLE was inactive, but even higher levels were seen during the active phases. Similarly, no associations were noted with any isotype of antibody to H1.

When the patients were considered individually, only one showed a clear association between ACHA and disease activity with \( p<0.001 \) for each of IgG, IgM, and IgA ACHA (fig 2). There was no association with antibodies to native DNA or erythrocyte sedimentation rate in this patient. One other patient had associations between antibodies to native DNA, erythrocyte sedimentation rate, and disease activity.

**Discussion**

The reported prevalence of AHA in unselected patients with SLE has varied from 21% to 81%.

We detected AHA in 74% (ACHAI in 68% and antibodies to H1 in 55%) of patients, but only 32% of the serum samples with ACHA had greater than twice the normal levels. These results are remarkably similar to the findings of Muller et al., who detected AHA in 78% of patients but noted that only 34% had high titres. Consequently, the ability to detect low levels of antibody may explain some of the wide variations in prevalence data. In addition, differences between patient groups, as illustrated in table 1, may go part of the way to explaining disparities in the prevalence and specificities of AHA cited in various reports.

Antibodies to core histones of the three major isotypes were common and occurred with similar frequencies. Most patients had more than one isotype of AHA which, in longitudinal studies, could persist over many years.

Although it was not possible to study serum samples from patients before the onset of SLE, in no patient was there evidence that a switch from IgM to IgG AHA, with subsequent loss of IgM AHA, had occurred. IgA AHA have been described previously, but not detailed, in patients with SLE. From this study, it can be seen that IgA AHA occur at a similar frequency to the other isotypes of AHA.

Several studies have reported the prevalence of antibodies to individual core histones but there has been no general agreement as to the prevalence of each fraction. Of the core histones, most have found that antibodies to H2B are common and that antibodies to H4 are uncommon, whereas the reported prevalences of antibodies to H2A and H3 have varied greatly. Antibodies to H1 have been reported to be the most common of the antibodies to individual histones in most studies, but this has not been a universal finding. Although several reports have noted a predominance of antibodies to H1 and H2B, most agree that a considerable diversity in the profiles of AHA may be seen in patients with SLE. Apart from a single study finding IgA antibodies to H1 in 18% of patients with SLE,32 IgA antibodies to individual histones have not been detailed. The data presented here suggest that antibodies to each of the histones, except H4, are common. Furthermore, for the core histones, all three major isotypes occur with similar frequencies, whereas IgM antibodies to H1 were less common than the other isotypes of antibodies to H1.

Antibodies to H2A-H2B complexes are characteristic of procainamide related lupus erythematosus33 and have been noted in a minority of patients with idiopathic SLE.17 33 The data presented here confirm this specificity is uncommon in patients with SLE (less than 2% of patients). Antibodies to H3-H4 complexes were detected in only two patients. This specificity has not been observed previously in patients with SLE, possibly because of its rarity. Nonetheless, a monoclonal antibody to this complex has been described34 and we have found that these antibodies occur in quinidine related lupus erythematosus25 and Felty’s syndrome.26

Very few clinical or laboratory associations were evident, which is consistent with other studies.35 When compared with patients with other autoantibodies, Fritzler et al36 reported that patients with AHA had a lower incidence of several clinical and laboratory features of SLE. However, none of the AHA positive group of patients had raised levels of antibodies to native DNA and are thus likely to represent a subset of patients with milder SLE. The associations in our study with mouth ulceration, psychiatric disturbances, and lymphopenia were all weak and a prospective analysis will be needed to determine whether they are genuine. The association with antibodies to native DNA (seen here only with IgG ACHA) or antibodies to denatured DNA has also been observed by most3 7 8 37 but not all, workers.6

The relationship between disease activity and the presence of AHA is unclear. Although several studies have shown an association, it is important to note that some studies failed to control for stronger associations between antibodies to native DNA and disease activity. In one of these studies,9 there was a negative association with antibodies to H1, despite the association between disease activity and anti-
bodies to total histones. In contrast, other studies have failed to show an association between AHA and disease activity.10–12 35 Another study showed no association between AHA and disease overall, but commented that an association was seen in some patients.3 This study supported the possibility that increasing levels of AcHA may be a marker of disease activity when all patients are considered together. However, no such association could be found in most of the individual patients.

The tissue deposition of AHA may result in only low levels being present in the serum, thus obviating the identification of clinical associations. Histone aggregates have been shown to have an affinity for the basement membrane and are able to bind AHA.38 This is probably related to the highly cationic nature of histones, a property which may allow histone containing immune complexes to be nephritogenic.39–41 In vivo, the source of such histones could be in complexes with DNA in the form of oligonucleosomes which have been detected in serum samples from some patients with SLE.42 Importantly, preliminary data indicate that immune complexes involving histone-DNA complexes are also likely to be cationic.43 Despite these studies offering insights into the possible tissue deposition of AHA and subsequent pathology, caution is warranted in view of the fact that no study has shown an association between renal involvement and either high or low levels of AHA. Moreover, AHA are a prominent feature of drug induced lupus erythematosus in which renal involvement is very unusual.3 44 45 It is reasonable to conjecture that only a subgroup of AHA are deposited, but this will remain hypothetical until AHA are eluted from the tissues and their specificities and properties studied.

In summary, we have shown that all major isotypes of AHA are common in patients with SLE. Whether all isotypes are considered together or separately, there were few clinical or laboratory associations. Moreover, our findings indicate that individual AHA levels are of no assistance in assessing disease activity, whereas sequential levels may be of limited value in a minority of patients.

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34 Laskov R, Muller S, Hochberg M, Giloh H, Van Regenmortel M H V.
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