Annals of the Rheumatic Diseases, 1985; 44, 316-320

Prostaglandin (PG) E₂ generation by cultured canine synovial fibroblasts exposed to microcrystals containing calcium

DANIEL J McCARTY AND HERMAN S CHEUNG

From the Division of Rheumatology, Department of Medicine, Medical College of Wisconsin, 8700 W Wisconsin Avenue, Milwaukee, Wisconsin 53226, USA

SUMMARY Immunoreactive prostaglandin (PG) E₂ was released into the ambient medium in a dose dependent fashion when either hydroxyapatite (HA) or calcium pyrophosphate dihydrate (CPPD) crystals were added to canine synovial fibroblasts in tissue culture. PGE₂ release peaked 6 to 9 hours after HA or CPPD crystals were added in the presence of serum but at 24 hours if they were added in the presence of lactalbumin hydrolysate. PGE₂ release correlated with crystal endocytosis estimated qualitatively by serial phase contrast microscopy and time lapse photography. As postulated previously by others for monosodium urate crystals, prostaglandin production by synovial cells may also be related to the pathogenesis of the destructive arthropathies associated with HA or CPPD crystals.

Key words: hydroxyapatite, calcium pyrophosphate.

Both acute inflammation and chronic destructive arthropathy have been described in association with basic calcium phosphate (BCP=hydroxyapatite, octacalcium phosphate, tricalcium phosphate) or calcium pyrophosphate dihydrate (CPPD) crystals.¹⁻⁵ Such tissue reactions resemble in some respects the acute inflammatory attack and chronic (tophaceous) destructive arthropathy associated with gout, the hallmark of which is the formation in tissues of monosodium urate (MSU) crystals. Each of these crystal species undergoes endocytosis by neutrophils and to a lesser extent by monocytes in acute inflammatory exudates.¹ ³ ⁶ Products of both the lipoxygenase and cyclo-oxygenase pathways of arachidonic acid metabolism have been identified when MSU crystals are exposed to these cells in vitro (reviewed by McCarty⁷), and products of both pathways have been identified when cultured human synovial fibroblasts⁸ or human neutrophils⁹ were exposed to MSU crystals. Cyclo-oxygenase products, mostly prostaglandin (PG)E₂ were elaborated in impressive quantities when MSU crystals were added to murine macrophages¹⁰ or lapine synovial fibroblasts.¹¹ ¹² Microcrystalline, but not amorphous, calcium oxalate produced effects similar to that of MSU crystals when incubated with cultured lapine synovial fibroblasts.¹³ PGE₂, and to a lesser extent, PGF₁α, generation by cultured human or canine synovial fibroblasts¹⁴ and by lapine chondrocytes in primary culture¹⁵ was augmented when cells were incubated with hydroxyapatite or CPPD crystals in concentrations greater than those actually found in synovial fluid from patients. As prostaglandins have been implicated as mediators in both acute inflammation¹⁶ and in bone resorption,¹⁷ they may relate to the pathogenesis of both the acute and chronic reactions to microcrystals.

The work reported here was performed with highly characterised synthetic hydroxyapatite (HA) or CPPD crystals in concentrations actually found in human joint fluid.

Materials and methods

General. All reagents were prepared in doubly deionised, glass distilled, charcoal filtered water. Cultures. Canine synovial cells were obtained from anaesthetised animals by a modification of the
method of Clarris et al. Visualy confluent second passage cultures from four mongrel dogs were used experimentally. There were about 1-5 to 2 × 10^6 cells/plate. Cells were grown in 100 mm plastic dishes (Scientific Products, McGaw Park, Ill.) containing 10 ml of Dulbecco’s modified Eagle’s essential medium (DMEM) supplemented with 10% horse serum and 1% penicillin, streptomycin, and amphotericin B (all from Gibco, Grand Island, NY). At the end of each experiment the cells were washed three times with Hanks’s balanced salt solution, released with 2 ml of 20 mM EDTA, and precipitated with an equal volume of 20% trichloroacetic acid. After dissolution of the centrifuged precipitate in 2 ml 0.1 N NaOH, the protein content was measured.

**Crystals.** Hydroxyapatite crystals were a kind gift from M D Francis, PhD, the Proctor and Gamble Co., Cincinnati, Ohio. They had a molar Ca/P of 1:67 and showed only HA adsorption bands by Fourier transform infrared spectrophotometry. Crystal aggregates were sieved to 10–20 μM with Endicott test sieves (London, England) and heated to 200°C for 2 h to assure sterility and to eliminate possible pyrogen contamination. Calcium pyrophosphate dihydrate crystals were synthesised, characterised, and sieved to 10–20 μM and heated as described previously. Monosodium urate monohydrate crystals were prepared and characterised as described previously. They were sonicated to 10–20 μM and then heated to 170°C for 2 h, which does not alter crystal structure.

**Prostaglandin (PG) E₂ assay.** This was done by radioimmunoassay by the method of Dunn et al. From 0:5 to 3 ml of conditioned medium was extracted for each determination to give the desired sensitivity. The higher values required dilution to give results that fell on the standard curve.

**Experiments.** Culture medium was replaced with DMEM alone, DMEM with 10% serum, or 0.2% lactalbumin hydrolysate (LA) with or without HA crystals, 100 μg/ml (mg/l) PG E₂ generation by synovial cells was determined. All cultures were examined at intervals up to 24 hours by phase-contrast microscopy. Several cultures containing crystals were examined continuously by time lapse photography with a microscope equipped with a camera (Nikon).

Samples of conditioned media were removed for PG E₂ assay at periodic intervals up to 24 hours, at which time the cells were released and protein content measured. Similar experiments were performed with MSU or CPPD crystals. Dose response was determined by addition of HA or CPPD crystals in concentrations varying from 0 to 500 μg/ml. Indomethacin was added to at least one plate containing crystals and serum in each experiment at final concentration of 10⁻⁶M. PG E₂, 45 500 pg/ml (ng/l) and 2400 pg/ml (ng/l) were incubated in DMEM–10% horse serum that had been conditioned for 24 hours by incubation with synovial cells, and samples were removed for radioimmunoassay at hourly intervals for the first six hours and then at 24 hours.

**Results**

All data are expressed as pg of PG E₂ per mg of cell protein. Cell protein per plate varied from 350 to 900 μg. Except for occasional clumps, nearly all crystals appeared to have undergone endocytosis within 4 to 6 hours when serum was present. In the presence of DMEM alone or when 0-2% LA was substituted, intracellular crystals were noted in large numbers only at 24 hours. These observations were confirmed by time lapse photography.

PG E₂ concentration in the conditioned medium increased in all cultures (Table 1). The increased

<table>
<thead>
<tr>
<th>Time (h)</th>
<th>DMEM + HA</th>
<th>DMEM + HA²</th>
<th>DMEM + HA³</th>
<th>DMEM 10% HA¹</th>
<th>DMEM 0.2% LA⁴</th>
<th>DMEM 10% HA⁵</th>
<th>DMEM 0.2% LA + HA</th>
<th>DMEM 10% HA + Ind⁶</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>114</td>
<td>0</td>
<td>120</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>3</td>
<td>24</td>
<td>36</td>
<td>19</td>
<td>2480±588</td>
<td>159±77</td>
<td>5.73±1.166</td>
<td>155±67</td>
<td>0</td>
</tr>
<tr>
<td>6</td>
<td>31</td>
<td>87</td>
<td>78</td>
<td>3659±1068</td>
<td>3565±1789</td>
<td>18.90±1.790</td>
<td>2.60±1.564</td>
<td>0</td>
</tr>
<tr>
<td>9</td>
<td>124</td>
<td>471</td>
<td>714</td>
<td>4629±954</td>
<td>3424±1540</td>
<td>23.93±4.057</td>
<td>5.22±2.978</td>
<td>0</td>
</tr>
<tr>
<td>24</td>
<td>821</td>
<td>30.285</td>
<td>88.109</td>
<td>2080±468</td>
<td>3238±1496</td>
<td>46.14±18.911</td>
<td>70.88±49.098</td>
<td>100±33</td>
</tr>
</tbody>
</table>

¹ HA = hydroxyapatite crystals, opsonised in 10% horse serum in DMEM washed 3 times; 100 μg/ml (mg/l) used throughout 10–20 μm aggregate size.
² HA = not opsonised.
³ HS = horse serum.
⁴ LA = lactalbumin hydrolysate.
⁵ Ind = indomethacin 10⁻⁶M.
level in the presence of DMEM alone was just detectable; in the presence of 10% serum or 0-2% LA, PGE$_2$ levels increased after 3 to 6 hours but remained stable. HA crystals added with 10% serum produced progressively greater PGE$_2$ levels up to 24 hours; when incubated with DMEM alone or with 0-2% LA the PGE$_2$ rise was delayed but showed a very large rise at 24 hours, sometimes exceeding that noted with crystals in the presence of serum (Fig. 1). Repeated experiments with cells from different animals gave widely variable results in terms of absolute numbers, but the shapes of the curves representing the time course of PGE$_2$ release were nearly identical.

The time course of release of PGE$_2$ from canine synovial cells by triclinic CPPD crystals and by MSU crystals is shown in Table 2. In all cultures MSU crystals resulted in a massive and progressive release of PGE$_2$ into the ambient medium. CPPD behaved like HA crystals, producing an early (3-6 h) rise in the presence of serum. The PGE$_2$ concentrations at 9 and 24 hours were similar with opsonised crystals in DMEM alone or in DMEM with 10% serum.

The PGE$_2$ released into the culture medium 9 hours after addition of varying concentrations of HA or CPPD crystals is shown in Table 3. The levels increased in proportion to the concentration of CPPD crystals and to the concentration of HA crystals at 50, 100, and 500 ng/ml (mg/l). Concentrations of HA crystals of 10 ng/ml (mg/l) or less were not associated with PGE$_2$ generation greater than that found in 10% horse serum alone.

Immunoreactive PGE$_2$ levels decreased gradually over time when incubated in conditioned media. Only 71% and 48% of the higher concentration remained at 6 and 24 hours respectively. At the lower concentration 66% and 48% remained at these times (data not shown).

![Time course of PGE$_2$ release from cultured canine synovial cells after addition of HA crystals (-----) 100 ng/ml (mg/l) in DMEM+10% HS, or (-----) in DMEM+0-2% lactalbumin hydrolysate. Data shown as mean±SEM, n=4.](http://ard.bmj.com/)

### Table 2  
**PGE$_2$ (pg/mg cell protein) released from cultured canine synovial cells by monosodium urate (MSU) or calcium pyrophosphate dihydrate (CPPD) crystals**

<table>
<thead>
<tr>
<th>Time (h)</th>
<th>MSU (500 ng/ml)</th>
<th>CPPD (100 ng/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>DMEM$^\dagger$</td>
<td>DMEM 0-2% LA$^\ddagger$</td>
</tr>
<tr>
<td>0</td>
<td>0</td>
<td>ND$^\S$</td>
</tr>
<tr>
<td>3</td>
<td>20.031</td>
<td>15.535</td>
</tr>
<tr>
<td>6</td>
<td>39.891</td>
<td>28.469</td>
</tr>
<tr>
<td>9</td>
<td>152.036</td>
<td>84.201</td>
</tr>
<tr>
<td>24</td>
<td>304.581</td>
<td>ND</td>
</tr>
</tbody>
</table>

$^\dagger$ DMEM=Dulbecco’s modified Eagle’s medium.

$^\ddagger$ LA=lactalbumin hydrolysate.

$^\S$ HS=horse serum.

$^\S$ OP=opsonised crystals.

$^\S$ Ind=indomethacin 10$^{-6}$M.

$^\S$ ND=not measured.
Table 3  PGE\textsubscript{2} released from cultured canine synovial cells by various concentrations of crystals after nine hours of incubation

<table>
<thead>
<tr>
<th>HA\footnote{HA=hydroxyapatite 10–20 \textmu m aggregate size.} (\textmu g/ml)</th>
<th>PGE\textsubscript{2}\footnote{DMEM=10% horse serum.} (pg/mg protein)</th>
<th>CPPD\footnote{CPPD=calcium pyrophosphate dihydrate—10–20 \textmu m.} (pg/mg protein)</th>
<th>PGE\textsubscript{2} (pg/mg protein)</th>
<th>PGE\textsubscript{2} (pg/mg protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>4.630</td>
<td>0</td>
<td>3.454</td>
<td>333</td>
</tr>
<tr>
<td>1</td>
<td>2.734</td>
<td>5</td>
<td>6.784</td>
<td>1,058</td>
</tr>
<tr>
<td>10</td>
<td>3.834</td>
<td>10</td>
<td>8.494</td>
<td>1,063</td>
</tr>
<tr>
<td>50</td>
<td>13.704</td>
<td>100</td>
<td>10.623</td>
<td>2,100</td>
</tr>
<tr>
<td>100</td>
<td>18.526</td>
<td>500</td>
<td>20.893</td>
<td>14,344</td>
</tr>
<tr>
<td>500</td>
<td>24.457</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Discussion

The results of these experiments with canine cells confirm previous reports that monosodium urate crystals stimulate PGE\textsubscript{2} production by cultured human\footnote{Dias A, Reynolds P, Reynolds J, et al. Exposure of human monocytes to calcium pyrophosphate crystals: stimulation of prostaglandin production. J Immunol 1990; 144: 411-6.} or lapine\footnote{Hayashi M, Hwang SN, Reynolds P, et al. Antigenicity of calcium pyrophosphate. J Rheumatol 1990; 17: 178-83.} synovial fibroblasts. Although precise comparison is not possible because of the variability of response when cells from different animals are used, it seems evident that hydroxyapatite or calcium pyrophosphate dihydrate crystals are much less potent stimulators of PGE\textsubscript{2} production than are MSU crystals (Tables 1 and 2). PGE\textsubscript{2} levels in the ambient culture medium showed a definite lag period after addition of either HA or CPPD crystals in serum, with a marked increase between 3 and 6 hours and a further rise at 9 and/or 24 hours. PGE\textsubscript{2} release correlated with qualitative assessment of endocytosis estimated by serial phase contrast microscopy and time lapse photography, as noted previously with MSU crystals.\footnote{Bouillon R, Deprez RO, McCarthy D, et al. Human monocytes stimulated with calcium pyrophosphate crystals: cytokine, monocyte chemotactic protein, and indoleamine 2,3-dioxygenase production. J Rheumatol 1990; 17: 178-83.} This release was found when synthetic HA crystals were used with properties similar to natural BCP crystals but only in concentrations of 50 \textmu g/ml (mg/l) or greater, levels not generally found in patients joint fluid.\footnote{Hasselbacher and colleagues found no need for serum when MSU crystals were added to murine macrophages or rabbit synovial fibroblasts, but LDH was released at 2 hours and this became massive after 20 hours.\footnote{McCarty D J, Gatter R A. Recurrent acute inflammation associated with focal apatite crystal deposition. Arthritis Rheum 1966; 9: 804-19.}} PGE\textsubscript{2} release by CPPD crystals was noted even at 5 \textmu g/ml (mg/l), far less than the mean of 72 \textmu g/ml (mg/l) found in chronic and 280 \textmu g/ml (mg/l) found in acute effusions.\footnote{McCarty D J, Koh N N, Faires J S. Recurrence of calcium phosphate crystals in the synovial fluid of arthritis patients: the 'pseudogout syndrome'. I. Clinical aspects. Ann Intern Med 1962; 56: 711-37.}

Both endocytosis and PGE\textsubscript{2} generation occurred but were delayed when either opsonised or uncoated crystals were added to cultured cells in the absence of serum. Wigley \textit{et al.} found that serum was required for PGE\textsubscript{2} generation from human synovial cells and that this was not accompanied by cell death as estimated by lactate dehydrogenase (LDH) release into the culture fluid.\footnote{Schumacher H R, Smolyo A P, Tse R L, Maurer K. Arthritis associated with apatite crystals. Arthritis Rheum 1977; 20: 411-6.} Hasselbacher and colleagues found no need for serum when MSU crystals were added to murine macrophages or...
McCarty, Cheung

Prostaglandin (PG) E2 generation by cultured canine synovial fibroblasts exposed to microcrystals containing calcium.

D J McCarty and H S Cheung

*Ann Rheum Dis* 1985 44: 316-320
doi: 10.1136/ard.44.5.316

Updated information and services can be found at:

[http://ard.bmj.com/content/44/5/316](http://ard.bmj.com/content/44/5/316)

**These include:**

**Email alerting service**
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

**Notes**

To request permissions go to:
[http://group.bmj.com/group/rights-licensing/permissions](http://group.bmj.com/group/rights-licensing/permissions)

To order reprints go to:
[http://journals.bmj.com/cgi/reprintform](http://journals.bmj.com/cgi/reprintform)

To subscribe to BMJ go to:
[http://group.bmj.com/subscribe/](http://group.bmj.com/subscribe/)