Detection of anti-dsDNA as a diagnostic tool: a prospective study in 441 non-systemic lupus erythematosus patients with anti-dsDNA antibody (anti-dsDNA)

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SUMMARY The diagnostic significance of anti-double-stranded deoxyribonucleic acid (anti-dsDNA) determination was evaluated in a prospective manner from 1974 to 1982 in a group of 441 patients without systemic lupus erythematosus whose sera were found to contain antibodies to dsDNA on routine screening (Farr assay). Within one year 69% (304) of these patients fulfilled the preliminary American Rheumatism Association (ARA) criteria for systemic lupus erythematosus (SLE). Eighty-two of the remaining 137 patients were followed up for several years. At the end of the study 52% of these patients had also developed systemic lupus erythematosus. Patients who developed systemic lupus erythematosus were characterised by the occurrence of relatively high avidity anti-dsDNA in the circulation compared with patients who did not develop systemic lupus erythematosus. It can be concluded that about 85% of patients without systemic lupus erythematosus with anti-dsDNA in the circulation will develop SLE within a few years. Taking into account the relative avidity of anti-dsDNA, as determined by calculation of Farr/polylethylene glycol (PEG) ratios, we conclude that patients with relatively high avidity anti-dsDNA are more prone to develop systemic lupus erythematosus than patients with relatively low avidity anti-dsDNA.

Key words: SLE, Farr assay, avidity, prognosis.

Antibodies to deoxyribonucleic acid (DNA) occupy a special position in the study of systemic lupus erythematosus (SLE). The appearance of antibodies to DNA has been alleged to predict exacerbations of disease,1 2 and the diagnostic importance of anti-dsDNA has recently been stressed by incorporation of its presence in serum as an ARA criterion.3 4

From early studies on DNA binding it was concluded that the Farr assay provides a sensitive and highly specific test for SLE.5 6 On the other hand high values of anti-DNA have also been described in patients with chronic active hepatitis and Q fever endocarditis.7 Most of the work on the specificity of anti-dsDNA assays has been carried out by testing panels of serum samples from selected patients with well defined clinical disease. In this way we have confirmed the specificity for SLE of anti-dsDNA measured by the Farr assay and by the immunofluorescence test (IFT) on Crithidia luciliae. However, when sera sent to us for routine diagnostic anti-dsDNA determination were tested, the specificity for SLE of both methods was completely lost.8 We found antibodies to dsDNA in many patients who were not diagnosed as cases of SLE and did not fulfil the preliminary ARA criteria. The sera we tested came from patients suspected of having SLE, and were therefore negatively selected for actually having SLE; a major reason for sending the sera to us was to obtain further information and to establish a diagnosis. The finding of anti-dsDNA

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in these non-SLE patients could be explained by the hypothesis that non-SLE patients with anti-dsDNA would develop SLE in the future, or that these patients formed a subset of SLE patients.

In order to evaluate the relevance of the presence of anti-dsDNA in non-SLE patients we performed a prospective study, from 1974 until 1982, in which all non-SLE patients with anti-dsDNA in their serum were followed up to see whether they would develop SLE or whether they had an SLE-like syndrome.

Materials and methods

ANTINUCLEAR ANTIBODY (ANA) DETERMINATION
To detect the presence of ANAs in sera of patients the indirect immunofluorescence technique was used. Sera were diluted 1:10 in phosphate-buffered saline (PBS) and incubated for 30 min at room temperature on acetone-fixed cryostat sections of rat liver tissue. After washing the slides for 30 min at 30°C with PBS (three changes) they were incubated with a 1:60 dilution of a fluorescein isothiocyanate (FITC)-conjugated polyclonal antiserum against human gammaglobulin (lot no. SH 17-01-F05; prepared in this institute). After a final 30 min washing procedure slides were mounted and read in a Leitz Orthoplan immunofluorescence microscope equipped with incident illumination. Nuclear fluorescence was taken as indication of the presence of ANAs.

ANTI-dsDNA DETERMINATION WITH THE FARR ASSAY
Details of the method have already been published.\(^9\)\(^-\)\(^12\) \(^3\)H-labelled PM2-DNA (100 ng) was incubated with 50 µl serum in the presence of 800 µg normal human gammaglobulin (HGG). The incubation volume was 0.2 ml and the medium PBS. After incubation of the mixture for one hour at 37°C 5 ml cold 50% saturated ammonium sulphate was added. The precipitate was allowed to form for 30 min at 4°C and was then centrifuged for 15 minutes at 3000 g in a refrigerated centrifuge, washed twice with 50% saturated ammonium sulphate, and counted. Mean binding of 135 control sera was 5%±3% (2×SD) of the input. Anti-dsDNA activity is expressed in terms of units/ml, one unit being defined as the amount of anti-dsDNA precipitating 30% of the added DNA under the above conditions.\(^12\)

ANTI-dsDNA DETERMINATION WITH THE PEG ASSAY
Details of this method have already been published.\(^13\)\(^-\)\(^14\) To 50 µl of serum was added 50 µl of a solution of 1.6 g/l HGG, 50 µl of a solution of 0.2 g/l dextran sulphate (DXS, Pharmacia Fine Chemicals AB, Sweden, lot no. 5259), 50 µl of a solution of 2 mg/l \(^3\)H-labelled DNA, and 200 µl of a solution of 7% (w/v) polyethylene glycol (PEG 6000, Koch-Light Laboratories, Colnbrook, UK). All components were dissolved or diluted in PBS. Incubation was carried out for one hour at 37°C followed by two hours at 4°C. After centrifugation for 15 min at 3000×g 200 µl of the supernatant was taken, dissolved in 8 ml NE-260sp (New England Nuclear Corp.), and counted for radioactivity. A 0% binding control, in which serum was omitted, was included in each experiment. Mean binding of 200 normal control sera was 6%±2% (2×SD). The anti-dsDNA activity was expressed in (PEG) units/ml, according to the definition used in the Farr assay.

PATIENTS' SERA
From all patients whose sera were sent to our laboratory in the period from 1974 to 1982 for routine ANA screening those patients found to have ANA (detected by the indirect IFT on rat liver

<table>
<thead>
<tr>
<th>Year</th>
<th>Total number of subjects</th>
<th>Number of patients followed up</th>
<th>SLE developed</th>
<th>During the follow up</th>
<th>Non-SLE patients</th>
<th>Lost during the study</th>
<th>Died</th>
<th>Other reasons*</th>
</tr>
</thead>
<tbody>
<tr>
<td>1974</td>
<td>52</td>
<td>42 (81%)</td>
<td>3</td>
<td>1</td>
<td>—</td>
<td>6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1975</td>
<td>66</td>
<td>48 (73%)</td>
<td>2</td>
<td>5</td>
<td>3</td>
<td>8</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1976</td>
<td>88</td>
<td>60 (68%)</td>
<td>4</td>
<td>14</td>
<td>2</td>
<td>8</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1977</td>
<td>61</td>
<td>40 (66%)</td>
<td>2</td>
<td>9</td>
<td>3</td>
<td>7</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1978</td>
<td>68</td>
<td>47 (69%)</td>
<td>8</td>
<td>7</td>
<td>—</td>
<td>6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1979</td>
<td>50</td>
<td>29 (58%)</td>
<td>6</td>
<td>4</td>
<td>1</td>
<td>10</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1980</td>
<td>56</td>
<td>38 (68%)</td>
<td>6</td>
<td>11</td>
<td>1</td>
<td>—</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>441</td>
<td>304 (69%)</td>
<td>31</td>
<td>51</td>
<td>10</td>
<td>45</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* Mostly caused by transfer to physicians other than those who participated in this study.
sections) and anti-dsDNA (detected by the Farr assay) were selected for study. Clinical features were obtained by means of a standard questionnaire. Once a year, until the end of our investigation in 1982, an evaluation was made of the course of disease in each patient, with special attention to symptoms incorporated in the preliminary ARA criteria. If a patient fulfilled at least four of the ARA criteria, the investigation was terminated. Only patients who did not fulfil the ARA criteria were followed up. In this way 441 patients, positive in the Farr assay and negative for the diagnosis of SLE, were studied.

Results

LONGITUDINAL FOLLOW UP DATA
In the period 1974–82 we studied a total of 441 non-SLE patients whose sera were found to contain antibodies to dsDNA. Within one year of entering the study 58% (1979) to 81% (1974) (mean: 69.5%) of these patients developed SLE according to the ARA criteria (Table 1). The remaining non-SLE patients (n=137) were studied for varying periods of time, depending on their year of entry into the study. During this follow-up study we lost 55 patients. Ten of these died of non-SLE-related causes and most of the other 45 transferred to physicians who did not participate in this study. The data in Table 2 show that, apart from the group of patients who developed SLE in the first year after entering the study, there was a tendency either to develop SLE in the following three years, or not to do so. If the lost patients (n=55) are considered not to have developed SLE, then 26% of the patients studied had SLE after five years of follow-up study. If these 55 patients are left out, the conclusion is that 52% of the patients studied had SLE at that time. The cumulative results are depicted graphically in Fig. 1. A steady increase in the percentage of SLE patients was seen during the first years. However, if a diagnosis of SLE was not reached after five years,

<table>
<thead>
<tr>
<th>Follow up years</th>
<th>Number of patients followed in relation to the duration of the follow up</th>
<th>Patients who developed SLE</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0-1</td>
<td>386</td>
<td>304</td>
</tr>
<tr>
<td>1-2</td>
<td>82</td>
<td>1</td>
</tr>
<tr>
<td>2-3</td>
<td>81</td>
<td>10</td>
</tr>
<tr>
<td>3-4</td>
<td>60</td>
<td>15</td>
</tr>
<tr>
<td>4-5</td>
<td>41</td>
<td>2</td>
</tr>
<tr>
<td>5-6</td>
<td>32</td>
<td>3</td>
</tr>
<tr>
<td>6-7</td>
<td>20</td>
<td>--</td>
</tr>
<tr>
<td>7-8</td>
<td>6</td>
<td>--</td>
</tr>
<tr>
<td>8-9</td>
<td>1</td>
<td>--</td>
</tr>
</tbody>
</table>

Fig. 1 Cumulative percentage of anti-dsDNA positive, non-SLE patients who developed SLE, expressed as a function of the duration of the follow-up study. *--* shows the results when the number of patients studied is corrected for patients who died or were lost during the study; *--* shows the results when the number of patients studied is taken as 137.
then such a patient — albeit having raised titres of anti-dsDNA in the circulation — was not prone to develop SLE. At the end of our study 335 out of the 386 anti-dsDNA positive, non-SLE patients we followed had developed SLE (i.e. 87%), mostly during the first year after entering the study (Fig. 2).

AVIDITY IN RELATION TO THE DEVELOPMENT OF SLE

Because all sera gave a positive reaction in the Farr assay, they could not be consigned to a low or high avidity anti-dsDNA group on the basis of this test. Expression of the anti-dsDNA content measured with the Farr and PEG assays in units/ml allowed calculation of the ratio between the Farr and PEG results. A high Farr/PEG ratio indicated a relative preponderance of high avidity anti-dsDNA, whereas the opposite held for a low Farr/PEG ratio. In Fig. 3 the Farr/PEG ratio of patients who developed SLE is compared with the ratio obtained from patients who had still not developed SLE at the end of the study. Patients developing SLE (group I) showed a mean Farr/PEG ratio of 5·4 compared with a Farr/PEG ratio of 2·8 for the non-SLE patients (group II). By drawing a line at a Farr/PEG ratio of

five, we find that 61% of the SLE patients (first group) have an equal or higher Farr/PEG ratio, compared with 18% of the non-SLE patients (second group) (p<0.0901, $\chi^2=3.85$).

CLINICAL AND LABORATORY FEATURES AT START OF STUDY

Table 3 shows that no clinical differences were detected between patients who developed SLE during the study and those who did not with respect to the number of preliminary ARA criteria they fulfilled when entering the study. With respect to the actual ARA criteria present a slight difference was recorded between the two groups. The incidence of ARA criteria 7 and 8 ranged from 68 to 73% in both groups, but criterion 14 was found in 15% of the patients in group I and in 30% of the

Fig. 2  Percentage of patients who developed SLE (n=335) as a function of the number of years passed since the initial anti-dsDNA detection and the clear development of SLE.

Fig. 3  Relative avidity of anti-dsDNA of two groups of anti-dsDNA positive, non-SLE patients. Group I. Thirty-one patients who had developed SLE at the end of the study; mean Farr/PEG ratio = 5·4 (SD 4.0). (Note: In seven cases no serum was available for the PEG assay; in six cases the PEG assay was negative.) Group II. Fifty-one patients who had not developed SLE at the end of the study; mean Farr/PEG ratio = 2·8 (SD 4.0). (Note: In 10 cases no serum was available for the PEG assay; in eight cases the PEG assay was negative.)
patients in group II. Criteria 10 and 11 were found in only two patients, one in each group.

If Fig. 4 initial anti-dsDNA levels of all patients followed for more than one year are depicted. In group I a mean anti-dsDNA level of 224 U/ml (SD 297) was found and in group II a mean value of 124 U/ml (SD 300). Statistical calculations did not show any correlation between the development of SLE and initial anti-dsDNA values.

Discussion

According to the revised ARA criteria for SLE the detection of antibodies to dsDNA can support the diagnosis of SLE. Since in the study presented here we sought to establish the relevance of anti-dsDNA determination in the diagnosis of SLE, we used the preliminary ARA criteria to differentiate between SLE and non-SLE in order to exclude anti-dsDNA positivity as a criterion for SLE.

During the period 1974–82 we studied 441 patients with serum antibodies to dsDNA who were not diagnosed as having SLE at the time. Nearly 70% of these non-SLE patients with anti-dsDNA in the circulation fulfilled the preliminary ARA criteria within one year after they were admitted to the follow-up study. After five years 52% of the 137 remaining patients had also developed SLE. The figure of 52% may be an overestimation, since 55 patients were lost during the study. If the presumption is made that all of these patients did not develop SLE, then a figure of 26% is obtained. On the other hand the possibility that 52% of these ‘lost’ patients also developed SLE cannot be excluded. If this were the case, then nearly 80% of the 137 non-SLE patients would have developed SLE after five years.

With the revised criteria for SLE another 20 patients would be confined to the SLE group at the end of the study. This would mean that over 70% of the patients who did not have SLE within the first year developed SLE during the five years of follow up study. All our patients were selected on the basis of anti-dsDNA and ANA positivity. As anti-dsDNA and ANAs go hand in hand in these cases, it will – in our view – be an overestimation of the value of these serological findings if both are used as separate criteria for SLE. We think that the diagnosis SLE should be made with caution because of future implications, for example the therapy required and, also the socioeconomic problems which may be raised, e.g., life insurance. With the revised criteria a patient with a quite good prognosis (e.g. mouth ulcers and slight skin rashes combined with a positive ANA test and a positive anti-dsDNA test) may be diagnosed as having SLE.

Our study shows that if anti-dsDNA is detected in a patient (by the Farr assay) the diagnosis SLE – according to the preliminary ARA criteria – can in most cases be made within one year. The remaining patients can be divided into two groups: those who

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**Table 3** Number (percentage) of anti-dsDNA positive, non-SLE patients with a specified number of ARA criteria present at the start of the study

<table>
<thead>
<tr>
<th>Group</th>
<th>No. of patients</th>
<th>Number of positive ARA criteria present</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>31</td>
<td>1 (16%) 5 (16%) 13 (42%) 13 (42%)</td>
</tr>
<tr>
<td>II</td>
<td>51</td>
<td>1 (29%) 15 (29%) 16 (31%) 20 (39%)</td>
</tr>
</tbody>
</table>

*Group I consisted of patients who had developed SLE at the end of the study, in contrast with group II patients who had not.*

**Fig. 4** Initial anti-dsDNA levels of the anti-dsDNA positive, non-SLE patients followed for more than one year.

○ = Patients who had not developed SLE at the end of the study. △ = Patients who developed SLE during the study.
develop SLE, albeit slowly, and those who after five years still do not have SLE. A question we sought to answer was whether it would be possible to differentiate between antibodies to dsDNA in patients who developed SLE from anti-dsDNA in patients who did not. We have shown before that the Farr assay only detects DNA antibodies of high avidity, whereas the PEG assay also detects anti-DNA of low avidity. From the results of both assays we could calculate a relative avidity index for each serum. From these data it became clear that patients with high avidity anti-dsDNA developed SLE in a shorter time than patients with low avidity anti-dsDNA. After five years of follow-up study a large proportion of the latter patients had still not developed SLE. There was no significant difference at the start of the study between the number of positive ARA criteria in patients who developed SLE and those who did not. This implies that if a patient is positive for one ARA criterion and has anti-dsDNA in the circulation, the chance for developing SLE is neither greater nor less than if the patient were positive for three ARA criteria. No obvious differences in the clinical features of patients at the start of the study could be found between groups I and II. ARA criteria 7 (arthritis without deformity) and 8 (LE cells) ranged from 68% to 73%, nearly the same in both groups. A slight difference was found in the frequency of criterion 14 (haemolytic anaemia and/or leucopenia and/or thrombocytopenia); the frequencies in groups I and II were 15% and 30% respectively. Criteria 10 (profuse proteinuria > 3.58 mg/day) and 11 (cellular casts) were found in only two patients in both groups.

No clear difference between the initially detected anti-dsDNA levels of the two groups was observed (group I: 224 U/ml (SD 297); group II: 124 U/ml (SD 300)) as illustrated in Fig. 4. This is in accord with our previous findings and confirms the absence of a correlation between anti-dsDNA level and disease symptoms or activity. The same study also showed that there is no relation between (initial) anti-dsDNA level and prognosis of disease. It is noteworthy that anti-dsDNA levels of group II patients generally remained stable throughout the follow-up study; in only 20% of the patients were anti-dsDNA levels decreased or negative at the end of the study. Patients who developed SLE within the first year after initial anti-dsDNA detection could not be distinguished from group I patients either on the basis of the first signs of disease (ARA criteria 7 and 8) or on the basis of sex, age, duration of complaints before the exacerbation, or type of exacerbation. All patients who developed SLE are still being studied, and data on survival rates are being accumulated.

In conclusion we have found that 69% of a group of 441 non-SLE patients with anti-dsDNA in the circulation developed SLE within one year. Of the remaining patients 53% developed SLE within five years of the follow-up study. Excluding the 55 patients who were lost during follow up, we followed a total of 386 patients during this prospective study. At the end of the study 331 patients (87%) had SLE, which confirms the significance of anti-dsDNA determination. We support the incorporation of anti-dsDNA as an ARA criterion but would emphasise that the method for detection is crucial. Data presented in this paper clearly indicate that patients having high avidity anti-dsDNA in the circulation are more prone to develop SLE than patients with low avidity anti-dsDNA. The IFT on Crithidia lucillae and the PEG assay also detect anti-dsDNA of low avidity and have indeed been found to be less specific for SLE than the Farr assay which was used throughout this study. Therefore the effective incorporation of anti-dsDNA in the criteria for SLE requires that the detection method is selective for high avidity anti-dsDNA. As contamination of the DNA preparation with single-stranded regions also leads to a decrease in specificity of the assay for SLE, care should be taken that the antigen preparation used consists of entirely double-stranded DNA. Longitudinal studies of patients having only anti-dsDNA of low avidity are in progress and will be published at a later date.

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References

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