Case report

Polymyositis in Chagas’s disease


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SUMMARY Polymyositis marked the clinical onset of Chagas’s disease in a patient with rheumatoid arthritis. This is unusual, although clinically unimportant muscle involvement in trypanosomiasis has been described. The plasma cell infiltrate and vascular deposition of IgM and C3 suggest that the humoral immune system may play a role in the pathogenesis of chagasic polymyositis. It is not known whether the rheumatoid diseases predisposed to the polymyositis.

There are few reports of myositis in Chagas’s disease, although this is commonly seen in experimental models (Wolf et al., 1952; Andrade and Andrade, 1968). Myositis associated with the presence of parasites was demonstrated by Ponce (1972) in sequential biopsies of the gastrocnemius muscle in patients with chronic and acute Chagas’s disease and cardiopathy. Cengot and Rojas (1959) observed deltoid myositis in 96.3% of 27 chagasic patients, but did not mention parasites in the muscle lesions.

Cossio et al. (1974a, b) described a circulating autoantibody that reacts against endocardium, as well as vessels and interstitium of skeletal muscle and myocardium and interacts with complement (EVI antibody). It was found in 95% of patients with Chagas’s heart disease and in 45% of asymptomatic individuals infected with Trypanosoma cruzi. In the early stage of acute heart disease EVI antibody is of IgM class but one month later both IgM and IgG antibodies are found. We report a patient with definite rheumatoid arthritis who developed chagasic polymyositis.

Case report

A 43-year-old white female had suffered from rheumatoid arthritis from age 17 years. Early in the course of the disease she received gold salts with remarkable improvement. After a 15-year period without symptoms, she then started to complain of joint pain in both hands, shoulders, and knees, with morning stiffness and fever of 38°C. Nonsteroidal anti-inflammatory drugs provided some relief. At this time she had her tonsils removed. Postoperatively she received 500 ml of fresh blood. 10 days after surgery her joint pains increased and were accompanied by muscle weakness and pain in the shoulders, thighs, and legs, fever of 40°C, and tachycardia. She was wholly incapacitated by the muscle weakness. Raised values of transaminases and LDH isoenzymes and positive antinuclear factor were recorded. Prednisone 80 mg bd was started, but 20 days later she was no better and was admitted to the Hospital das Clínicas da Universidade de São Paulo.

Her general condition was fair, blood pressure 110/70 mmHg, pulse rate 100/min, respiratory rate 30/min, and temperature 38°C. No cardiac murmurs or changes in heart sounds or rhythm disturbances were found. The lungs were normal on examination. The spleen was not palpable, but the liver was enlarged about 3 cm. There was pain on joint movement in both hands, shoulders, knees, and ankles. The left ankle was swollen without limited movement. Muscle weakness in a symmetrical, proximal distribution was the prominent feature, and the affected muscles were tender to pressure. Tendon reflexes were well preserved.

The erythrocyte sedimentation rate was 49 mm/h, RA latex fixation test 1:80, negative LE-cell test, antinuclear antibodies (ANA) positive at a 1:5000 dilution (rat liver preparation), total haemolytic complement 15 U (normal: 166–333), C3 0.6 g/l (normal 0.8–1.2 g/l). Total protein measured 102 g/l, gammaglobulin 44 g/l. Serum muscle enzymes were markedly raised: creatine phosphokinase (CPK)
176 IU/l (normal 0–50 IU/l), lactic dehydrogenase (LDH) 746 IU/l (normal 80–240 IU/l), and SGOT 207 Frankel U/ml (normal 8–40 U/ml). Electromyography suggested primary myopathy with some signs of denervation. X-ray showed erosions and joint-space loss in metacarpophalangeal and proximal interphalangeal joints in both hands.

The diagnosis of American trypanosomiasis was suggested by specific serological reactions and confirmed by the histopathological findings. The former included the complement fixation test, positive at 1:64; the immunofluorescent indirect reaction for *T. cruzi* showed IgM and IgG antibodies at 1:160 and 1:320 respectively, and haemagglutination test with red cells coated with protein and polysaccharide extracts of *T. cruzi* were both positive at 1:320.

Rectal biopsy showed leishmania parasites within smooth muscle fibres and ganglion cells (Fig. 1). Two skeletal muscle biopsies were performed, one before and one after the introduction of specific therapy for acute Chagas's disease. Both showed myositis with hyalin focal necrosis in muscle fibres (Figs. 2, 3), but parasites were found only in the former. Lymphocytes, plasma cells, and macrophages were present, predominantly arranged in large perivascular collections (Fig. 4).

In the chagasic muscle, direct immunofluorescent reactions showed granular deposits of IgM and C3 in the walls of small arteries (Fig. 5), while the reaction for IgG had a linear sarcolemmal pattern. IgA and fibrinogen were not found. In the normal control muscle all reactions were negative, including the indirect immunofluorescent reaction for EVI antibody, using the patient's serum.
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The clinical and laboratory evidence of polymyositis was out of proportion to that which has been recorded as occasionally accompanying rheumatoid arthritis. The finding of parasites in the muscle lesions suggests a chagasic aetiology for the polymyositis. The parasite presumably gained access to the body through the blood transfused one month before. No other mode of transmission can be invoked since she had not been in an endemic zone for several years. These data, together with the results of the haemagglutination and immunofluorescent reactions, strongly suggest acute Chagas's disease and rule out the possibility of a chronic subclinical form of Chagas's disease reactivated by the corticosteroid therapy.

In the acute stage of Chagas's disease, the clinical picture is generally unimpressive but there may be general and cardiac manifestations. Polymyositis has not been reported as a clinical manifestation of any stage of Chagas's disease although histopathological studies of skeletal muscle have often shown conspicuous myositis (Cenget and Rojas, 1959; Ponce, 1972). It seems likely that minor degrees of clinical skeletal muscle involvement may pass unnoticed in the presence of more dramatic cardiac or other features.

The occurrence of synovitis in association with dermatomyositis and the presence of myositis in rheumatoid arthritis raises the possibility that the skeletal muscles in a patient with rheumatoid arthritis may be more vulnerable to damage from other causes. As far as the pathogenesis of polymyositis is concerned, there is evidence that cell-mediated reaction to skeletal muscle and concomitant humoral immunodeficiency are involved (Dawkins, 1975). The vascular deposition of IgM and C3 seen in this case suggest that humoral immune factors may have played a pathological role here. Whitaker and Engel (1972) noted granular deposit of IgG, IgM, and C3 in skeletal muscle blood vessel walls and suggested that the vasculitis may be one of the mechanisms of muscle injury, particularly in dermatomyositis of children.

In our case, the significant findings were the deposition of IgM and C3 in arterial wall and the perivascular mononuclear infiltrate. Necrotising vasculitis, which was frequently found in Ponce's series, was not observed and the EVI factor described by Cossio was not found. We believe that the clinical polymyositis resulted from vascular involvement besides the myositis itself. Our observations also suggest that in Chagas's disease the humoral immune system must play an important role in the pathogenesis of polymyositis.

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References


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W Cossermelli, H Friedman, E H Pastor, M R Nobre, A Manzione, M E Camargo and M Shiroma

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